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# A comparison study of GeneXpert and In-House N1N2 CDC Real-Time RT-PCR for detection of SARS-CoV-2 infection

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#### ABSTRACT

Submitted: 2022-03-01 Accepted : 2022-05-06 COVID-19 is a disease caused by SARS-CoV-2, a new virus from genus  $\beta$ -coronaviruses. This disease has been declared a pandemic by WHO on 11 March 2020 until now. The nucleic acid tests are the most frequently used assays because of their high sensitivity and specificity. One of the tests is the GeneXpert, a real-time reverse transcription polymerase chain reaction (rRT-PCR)-based assay platform. The use of the GeneXpert shows great public health interest because of the rapid (50 min), the minimum number of trained staff, and less infrastructure and equipment. However, there are limited data on the application of the GeneXpert for the detection of SARS-CoV-2. Therefore, we conducted a comparative study between the GeneXpert and in-house N1N2 CDC rRT-PCR assay. Of 86 samples, 17 were rRT-PCR positive while 13 were GeneXpert positive. Of rRT-PCR positive 17 samples, 7 were GeneXpert negative [58.82% (10/17] sensitivity]. We also found that 3 GeneXpert positive samples showed rRT-PCR negative (95.65% [66/69] specificity). It is concluded that negative results by the GeneXpert can not rule out the possibility of SARS-CoV-2 infection, particularly in close-contact individuals and the interpretation of the positive result should be analyzed carefully, particularly amplification with Ct>40.

#### ABSTRAK

COVID-19 adalah penyakit yang disebabkan oleh SARS-CoV-2, virus baru dari genus β-coronaviruses. Penyakit ini telah dinyatakan sebagai pandemi oleh WHO pada 11 Maret 2020 hingga sekarang. Tes asam nukleat adalah tes yang paling sering digunakan karena sensitivitas dan spesifisitasnya yang tinggi. Salah satu tesnya adalah GeneXpert, platform pengujian berbasis realtime reverse transcription polymerase chain reaction (rRT-PCR). Penggunaan GeneXpert menunjukkan minat kesehatan masyarakat yang besar karena kecepatan (50 menit), jumlah staf terlatih yang minimum, dan infrastruktur dan peralatan yang lebih sedikit. Namun, ada keterbatasan data dalam aplikasi GeneXpert untuk deteksi SARS-CoV-2. Oleh karena itu, kami melakukan studi perbandingan antara GeneXpert dan uji rRT-PCR N1N2 CDC in-house. Dari 86 sampel, 17 adalah rRT-PCR positif sementara 13 adalah GeneXpert positif. Dari 17 sampel rRT-PCR positif, 7 adalah GeneXpert negatif (sensitivitas 58,82% [10/17]). Kami juga menemukan bahwa 3 sampel positif GeneXpert menunjukkan rRT-PCR negatif (95,65% [66/69] spesifisitas). Disimpulkan bahwa hasil negatif oleh GeneXpert tidak dapat mengesampingkan kemungkinan infeksi SARS-CoV-2, terutama pada individu yang kontak dekat dan interpretasi hasil positif harus dianalisis dengan cermat, terutama amplifikasi dengan Ct>40.

*Keywords*: COVID-19; SARS-CoV-2; GeneXpert; PCR; nucleic acid tests

# INTRODUCTION

Coronavirus disease 2019 (COVID-19) is a disease caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) from Family *Coronaviridae*.<sup>1,2</sup> Genetic analysis showed that SARS-CoV-2 has similarities with another coronavirus (SARS-CoV) and is grouped within β-coronavirus genus.<sup>2</sup> SARS-CoV-2 is an enveloped virus and has a positive-sense single-stranded RNA genome enclosed in structural nucleocapsid protein.<sup>2,3</sup> (N) Other structural proteins namely E (envelope), M (membrane), and S (spike) proteins form viral envelope.<sup>2,3</sup> Among these structural proteins, S protein facilitates viral entry to the host cell by binding with the ACE receptor.<sup>3</sup>

This is a new virus discovered in 2019 in Wuhan, China, after some patients showed symptoms of flu-like illness.<sup>4</sup> In March 2020, COVID-19 was declared a pandemic by WHO because of the number of cases and countries with cases increase.<sup>5</sup> Globally, the numbers COVID-19 patients continuously of increased, with 500 million confirmed cases and over 6 million deaths have been reported from the beginning of the pandemic until the third week of April 2022.<sup>6</sup> In Indonesia, COVID-19 cases have been over 6 million and more than 155,746 deaths have been reported.<sup>7</sup>

At the beginning of the pandemic, there is an urgent need for a highly specific and sensitive method to detect the virus.<sup>2,4,8</sup> Currently, many testing methods are available for the detection of SARS-CoV-2.<sup>2</sup> The most common genes for nucleic acid detection of SARS-CoV-2 are orf1a/b, RdRp, S, N, and E.<sup>1</sup> Some diagnostic methods used to detect SARS-CoV-2 are serological (antigen and antibody detection) and nucleic acid tests such as standard real-time reverse transcription-polymerase chain reaction (rRT-PCR) and rapid tests like RT-LAMP, and GeneXpert assays.<sup>2,9</sup> The rRT-PCR is a gold standard for detection of SARS-CoV-2 because its sensitivity and specificity.<sup>1,2,8</sup> On the other hand, the GeneXpert using single cartridge-based assay is a rapid method for detection of COVID-19 compared to the standard rRT-PCR assay.<sup>5,8</sup> The rapid turnover of the GeneExpert result makes it as an increasingly popular choice for the detection of SARS-CoV-2. However, to our knowledge there is limited data on the GeneExpert performance particularly on its sensitivity and specificity. Therefore, in this study we compared standard rRT-PCR based on the N1N2 CDC protocol and GeneXpert assay.

## **MATERIALS AND METHODS**

## **Clinical specimens**

Eighty-six nasopharyngeal/ oropharyngeal swabs were obtained from suspected COVID-19 individuals in Jakarta from September-December 2020. The swab samples were collected immediately into 1 mL of the viral transport medium (DMEM containing 1% pen-strep and 5% bovine serum albumin) and stored at 2-4°C for not more than 4 h. The viral transport medium was divided for GenXpert and rRT-PCR tests conducted by two separate teams (blind testing). This study was approved by the Ethics Committee, Faculty of Medicine, Universitas Indonesia (KET-395/UN2.F1/ ETIK/PPM.00.02/2020).

### Viral RNA extraction

The viral RNA genome was extracted by using QIAmp Viral RNA Mini Kit (Qiagen, Germany) in accordance with the manufacturer's instructions. The final elute was stored at -80°C for not more than 4h.

#### **Real-time RT-PCR (rRT-PCR) assay**

The primers and probes for N (N1 and N2) and human RNase P (internal control, IC) genes based on the Centers for Disease Control and Prevention (CDC) were used for the detection of SARS-CoV-2.10 The rRT-PCR was performed with the following composition (20  $\mu$ l of total volume): 1x SensiFAST™ Probe No-ROX One-Step mix (Bioline, Cat. No: BIO-76005), 1.5  $\mu$ L each of primer and probe solution (2019-nCoV RUO Kit, IDT Integrated DNA technologies, Cat. no:10006713), 4U of RNase inhibitor, 2 U of reverse transcriptase enzyme, and 7.9 µL of RNA template. The PCR machine, MA-6000 Real-Time PCR System [Molarray, Suzhou, China]), was used under the following conditions: 50°C for 50 min; 95°C for 50 min; 45 cycles of 95°C for 15 sec and 55°C for 30 sec. The rRT-PCR positive was defined if  $Ct \le 40$  for both N1 and N2.10

## **Rapid GeneXpert test**

The GeneXpert used Xpert® Xpress SARS-CoV-2 kit based on N (N2 CDC) and E genes for detection of SARS-CoV-2.11 The procedure and the result interpretation were performed according to the manufacturer's instructions.<sup>11</sup> SARS-CoV-2 positivity was defined if either gene (N2 and E) or only N2 were positive. The presumptive SARS-CoV-2 positive was defined if only the E gene was positive (Ct value  $\leq$  45).

## **Statistical analysis**

The SPSS 16.0 was used for statistical analysis and a fisher test with a 5% (0.05) level of significance was used for hypothesis testing.

### **RESULTS**

The comparison results between **RT-PCR** real-time (rRT-PCR) and GeneXpert are shown in TABLE 1. Of 86 samples, 17 were rRT-PCR positive while 13 were GeneXpert positive. Of rRT-PCR positive 17 samples, 7 were GeneXpert negative (41.18% [7/17] discrepancy). The GeneXpert negative samples had Ct values above 34 by rRT-PCR (TABLE 2), indicating that GeneXpert failed to detect SARS-CoV-2 with high Ct values above 34.

TABLE 1. Comparison results between real-time RT-PCR (rRT-PCR) and
GeneXpert methods (n=86)

Positive		GeneXpert		~*	OR
		Negative		— p*	UK
rRT-PCR	Positive	10	7	< 0.001	31.429
IRI-PCK	Negative	3	66	< 0.001	31.429

	Real-time RT-PCR			GeneXpert		
Sample ID	Region target (Ct value )		Result	Region target (Ct value )		Result
	N1	N2		N2	Е	
2809-23	34.22	33.93	+	ND	ND	-
2809-31	37.67	38.15	+	ND	ND	-
2809-20	38.34	37.81	+	ND	ND	-
2809-21	35.91	37.72	+	ND	ND	-
2809-14	ND	ND	-	42.4	ND	+
2909-06	34.75	34.49	+	ND	ND	-
2909-13	36.15	38.28	+	ND	ND	-
0510-05	37.56	36.40	+	ND	ND	-
1008-08	ND	ND	-	44.2	39.2	+
1208-38	ND	ND	-	41.3	ND	+

TABLE 2. Discrepancy results between real-time RT-PCR (rRT-
PCR) and GeneXpert for detection of SARS-CoV-2

Note: All tests were valid with internal control Ct of < 30. N1 and N2: Regions of N gene. E: Envelope gene. ND: Not detected. Ct: Cycle threshold. +: Positive. -: Negative.

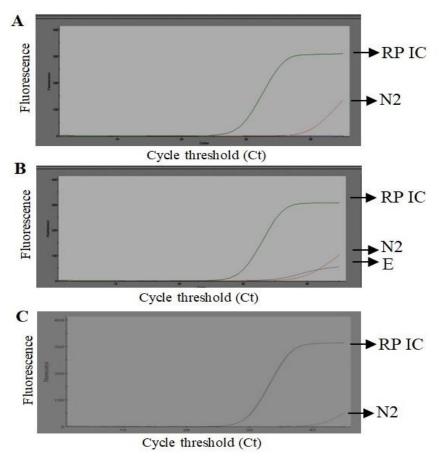


FIGURE 1. Curve Amplification for ID samples 2809-14 (A), 1008-08 (B), and 1208-38 (C). RP IC: Human RNase P gene internal control. N2: Region of N gene. E: E gene.

Test	Value
Sensitivity	0.5882 (58.82 %)
Specificity	0.9565 (95.65 %)
PPV	0.4158
NPV	0.9778

TABLE 3.	Sensitivity, specificity, PPV, and
	NPV of GeneXpert assay (prior
	probability of infection 0.05

We found that 3 GeneXpert positive showed rRT-PCR negative samples (TABLE 2). Of 3 samples, 2 were detected for N2 (Ct>40) and E (Ct=0), while 1 was detected for N2 (Ct>40) and E (Ct=39.2). Based on manual curve analysis, all samples showed sigmoid curves of RP gene internal controls, while N2 and E curves were not sigmoid (FIGURE 1). Because of the questionable results, real-time RT-PCR reaction another (Detection Kit for 2019-nCoV, Cat. no: #DA-930, Da An Gene Co., Ltd. of Sun Yat Sen University), a kit listed by the WHO Emergency Use for detection of SARS-CoV-2 nucleic acid, was performed for clarification. The results showed that all 3 samples were SARS-CoV-2 negative (Data not shown).

## **DISCUSSION**

rRT-PCR The and GeneXpert compared in this study have the same gene target [nucleocapsid (N)] for SARS-CoV-2; however, the rRT-PCR detects two regions (N1 and N2) of the N gene, while the GeneXpert detects only one region (N2).<sup>10,11</sup> The GeneXpert detects an additional gene, envelope (E) for all coronaviruses.<sup>11</sup> For detection of specific SARS-CoV-2, regions of N gene including N1 and N2 have been reported as rRT-PCR targets with higher sensitivity than other gene targets.<sup>12-14</sup> The high sensitivity might be caused by a high number of subgenomic mRNA of the N gene produced during the replication of coronaviruses.<sup>15</sup> Comparison between N1 and N2 applied for clinical and environmental samples, most of studies reported N1 having higher sensitivity than N2,<sup>12,16-19</sup> and another study reported an otherwise result.<sup>13</sup> Even though N1 was more sensitive than N2, several valid results were N2 positive and N1 negative.<sup>18</sup> Thus, it is suggested that N1 and N2 primer-probe sets should be used for detection of specific SARS-CoV-2.

The GeneXpert failed to detect SARS-CoV-2 in 7 samples that were positive by rRT-PCR (TABLE 2). Procop et al. reported that the GeneXpert had afalsenegative rate of 2% compared with N1N2 CDC rRT-PCR.<sup>20</sup> Other studies also reported the false-negative results by the GeneXpert.<sup>21</sup> Based on Ct value, the falsenegative occurred in cases with high Ct values above 34 (TABLE 2). The Ct values can be used as surrogate markers for deducing the virus infectivity. For this reason, several studies have reported the association of Ct values with virus infectivity by using cell culture methods. It has been shown that patients with Ct values above 30 or 34 did not excrete infectious viral particles.<sup>22,23</sup>

However, other studies have reported otherwise data.<sup>24-26</sup> Two studies reported that clinical samples with Ct-values above 30 could still be infectious.<sup>24,25</sup> Singanayagam *et al.*<sup>26</sup> reported that 8% of samples with Ct above 35 were still infectious. The different results might be affected by different pre-analytic and post-analytic factors in each laboratory, making Ct values as surrogate markers are unclear and debatable. Thus, Platten *et al.*,<sup>27</sup> suggested that the Ct value cutoffs can be defined as acceptable lowrisk values; higher Ct values as lower infection risks. Based on the data, it is suggested that SARS-CoV-2 negative by the GeneXpert cannot rule out the possibility of SARS-CoV-2 infection.

On the other side, the GeneXpert showed 3 false-positive results (TABLE 1). Based on Ct values, all 3 samples were detected for N with Ct>40 and only 1 sample was detected for E with Ct= 39.2 (TABLE 2). Moreover, N2 and E amplification curves showed nonsigmoid curves (FIGURE 1). The question results have been clarified by another kit and showed SARS-CoV-2 negative (Data not shown). These GeneXpert false-positive results have been reported by Rakotosamimanana *et al.*,<sup>28</sup> in that they found samples, that were no amplification of E gene (Ct=0) and N2 with Ct>40 by GenXpert, are negative by standard rRT-PCR assay. Other studies also reported the same result patterns.<sup>21,29</sup> Das *et al.* reported 16 (34%) of samples with Ct>35 by GenXpert were only 3 (18.8%) positive by standard rRT-PCR assay.<sup>21</sup> Moreover, Moran *et al.*,<sup>29</sup> reported that the GeneXpert results with E gene (Ct=0) and N2 with Ct>40 were SARS-CoV-2 negative when performing the repeated GeneXpert testing. Therefore, we suggested the repeated GeneXpert testing for clarification when the results were N2 with Ct>40.

## **CONCLUSION**

The negative results by GeneXpert cannot rule out the possibility of SARS-CoV-2 infection, particularly for closecontact individuals. Due to automatic interpretation by the GeneXPert software, the interpretation of the positive result should be analyzed carefully, particularly Ct>40. The sensitivity and specificity of the GeneXpert were 58.82% and 95.65% respectively. However, it is important to know that there is a limitation to this study, namely the small number of the samples used.

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## REFERENCES

 Chen W, Xiao Q, Fang Z, Lv X, Yao M, Deng M. Correlation analysis between the viral load and the progression of COVID-19. Comput Math Methods Medi 2021; 2021:9926249. https://doi.org/10.1155/2021/0026240

https://doi.org/10.1155/2021/9926249

- Jalandra R, Yadav AK, Verma D, Dalal N, Sharma M, Singh R, *et al.* Strategies and perspectives to develop SARS-CoV-2 detection methods and diagnostics. Biomed Pharmacother 2020; 129:110446. h t t p s : //doi.org/10.1016/j. biopha.2020.110446
- 3. Afzal A. Molecular diagnostic technologies for COVID-19: Limitations and challenges. J Adv Res 2020; 26:149-59. https://doi.org/10.1016/j.jare.2020.08.002
- 4. Uhteg K, Jarrett J, Richards M, Howard C, Morehead E, Geahr M, *et al.* Comparing the analytical performance of three SARS-CoV-2 molecular diagnostic assays. J Clin Virol 2020; 127:104384.

https://doi.org/10.1016/j.jcv.2020.104384

- 5. Goldenberger D, Leuzinger K, Sogaard KK, Gosert R, Roloff T, Naegele K, et al. Brief validation of the novel GeneXpert Xpress SARS-CoV-2 PCR assay. J Virol Methods 2020; 284:113925. https://doi.org/10.1016/j. iviromet.2020.113925
- 6. WHO. COVID-19: epidemiology,

virology, and prevention. Up To Date; 2022.

https://www.uptodate.com/contents/ covid-19-epidemiology-virologyand-prevention

- WHO. Novel Coronavirus; 2022. https://www.who.int/indonesia/ news/novel-coronavirus.
- 8. Vaz SN, de Santana DS, Netto EM, Wang WK, Brites C. Validation of the GeneXpert Xpress SARS-CoV-2 PCR assay using saliva as biological specimen. Braz J Infect Dis 2021; 25(2):101543.

https://doi.org/10.1016/j.bjid.2021.101543

- 9. Eftekhari A, Alipour M, Chodari L, Maleki Dizaj S, Ardalan M, Samiei M, et al. A comprehensive review of detection methods for SARS-CoV-2. Microorganisms 2021; 9(2):232. h t t p s : // d o i . o r g / 1 0 . 3 3 9 0 / microorganisms9020232
- CDC. Research use only 2019-novel coronavirus (2019-nCoV) realtime RT-PCR primers and probes. 2021. Available at https://www.cdc. gov/coronavirus/2019-ncov/lab/ rt-pcr-panel-primer-probes.html). Accessed December , 14, 2021,
- US Food and Drug Administration. Xpert Xpress SARS-CoV-2. (Package insert.) US Food and Drug Administration, Silver Spring, MD. 2021. Available at https://www. cepheid.com/en/coronavirus. Accessed December, 14, 2021.
- 12. Vogels CBF, Brito AF, Wyllie AL, Fauver JR, Ott IM, Kalinich CC, *et al.* Analytical sensitivity and efficiency comparisons of SARS-CoV-2 RT–qPCR primer–probe sets. Nat Microbiol 2020; 5(10):1299-305. https://doi.org/10.1038/s41564-020-0761-6
- 13. Nalla AK, Casto AM, Huang MLW, Perchetti GA, Sampoleo R, Shrestha L, *et al.* Comparative performance of SARS-CoV-2 detection assays using seven different primer-probe sets and one assay kit. J Clin Microbiol 2020; 58(6):e00557-20.

https://doi.org/10.1128/JCM.00557-20

- 14. Chu DKW, Pan Y, Cheng SMS, Hui KPY, Krishnan P, Liu Y, *et al.* Molecular diagnosis of a novel coronavirus (2019-nCoV) causing an outbreak of pneumonia. Clin Chem 2020; 66(4):549-55. https://doi.org/10.1093/clinchem/hvaa029
- 15. Moreno JL, Zúñiga S, Enjuanes L, Sola I. Identification of a coronavirus transcription enhancer. J Virol 2008; 82(8):3882-93.

https://doi.org/10.1128/JVI.02622-07

16. Huang Y, Johnston L, Parra A, Sweeney C, Hayes E, Hansen LT, *et al.* Detection of SARS-CoV-2 in wastewater in Halifax, Nova Scotia, Canada, using four RT-qPCR assays. FACETS 2021; 6:959-65.

https://doi.org/10.1139/facets-2021-0026

17. Etievant S, Bal A, Escuret V, Brengel-Pesce K, Bouscambert M, Cheynet V, *et al.* Performance assessment of SARS-CoV-2 PCR assays developed by WHO referral laboratories. J Clin Med 2020; 9(6):1871. https://doi.org/10.2200/jcm9061871

https://doi.org/10.3390/jcm9061871

- 18. Coryell MP, Iakiviak M, Pereira N, Murugkar PP, Rippe J, Williams DB, *et al.* A method for detection of SARS-CoV-2 RNA in healthy human stool: a validation study. Lancet Microbe 2021; 2(6):e259-e66. https://doi.org/10/1016/S2666-5247(21)00059-8
- 19. Hong PY, Rachmadi AT, Mantilla-Calderon D, Alkahtani M, Bashawri YM, Al Qarni H, *et al.* Estimating the minimum number of SARS-CoV-2 infected cases needed to detect viral RNA in wastewater: To what extent of the outbreak can surveillance of wastewater tell us? Environ Res 2021; 195:110748. https://doi.org/10.1016/j.

envres.2021.110748

20. Procop GW, Brock JE, Reineks EZ, Shrestha NK, Demkowicz R, Cook E, *et al.* A Comparison of five SARS-CoV-2 molecular assays with clinical correlations. Am J Clin Pathol 2021; 155(1):69-78.

https://doi.org/10.1093/ajcp/aqaa181

21. Das R, Joshi S, Pednekar S, Karyakarte R. Comparison of Xpert Xpress SARS-CoV-2 assay and RT-PCR test in diagnosis of COVID-19. IOSR-JDMS 2021; 20(6):12-7.

https://doi.org/10.9790/0853-2006131217

22. La Scola B, Le Bideau M, Andreani J, Hoang VT, Grimaldier C, Colson P, *et al.* Viral RNA load as determined by cell culture as a management tool for discharge of SARS-CoV-2 patients from infectious disease wards. Eur J Clin Microbiol Infect Dis 2020; 39(6):1059-61. https://doi.org/10.1007/s10096-020-

03913-9

- 23. Glenet M, Lebreil AL, Heng L, N'Guyen Y, Meyer I, Andreoletti L. Asymptomatic COVID-19 adult outpatients identified as significant viable SARS-CoV-2 shedders. Sci Rep 2021; 11(1):20615. https://doi.org/10.1038/s41598-021-00142-8
- 24. Arons MM, Hatfield KM, Reddy SC, Kimball A, James A, Jacobs JR, *et al.* Presymptomatic SARS-CoV-2 infections and transmission in a skilled nursing facility. N Engl J Med 2020; 382(22):2081-90. https://doi.org/10.1056/NEJMoa2008457
- 25. Kujawski SA, Wong KK, Collins JP, Epstein L, Killerby ME, Midgley CM, *et al.* Clinical and virologic

characteristics of the first 12 patients with coronavirus disease 2019 (COVID-19) in the United States. Nat Med 2020; 26(6):861-8.

https://doi.org/10.1038/s41591-020-0877-5

- 26. Singanayagam A, Patel M, Charlett A, Lopez Bernal J, Saliba V, Ellis J, *et al.* Duration of infectiousness and correlation with RT-PCR cycle thresholdvalues in cases of COVID-19, England, January to May 2020. Euro Surveill 2020; 25(32):2001483. https://doi.org/10.2807/1560-7917. ES.2020.25.32.2001483
- 27. Platten M, Hoffmann D, Grosser R, Wisplinghoff F, Wisplinghoff H, Wiesmüller G, *et al.* SARS-CoV-2, CTvalues, and infectivity-conclusions to be drawn from side observations. Viruses 2021; 13(8):1459. https://doi.org/10.3390/y13081459

https://doi.org/10.3390/v13081459

- 28. RakotosamimananaN,Randrianirina F, Randremanana R, Raherison MS, Rasolofo V, Solofomalala GD, *et al.* GeneXpert for the diagnosis of COVID-19 in LMICs. Lancet Glob Health 2020; 8(12):e1457-e8. https://doi.org/10.1016/S2214-109X(20)30428-9
- 29. Moran A, Beavis KG, Matushek SM, Ciaglia C, Francois N, Tesic V, *et al.* Detection of SARS-CoV-2 by use of the cepheid Xpert Xpress SARS-CoV-2 and roche cobas SARS-CoV-2 assays. J Clin Microbiol 2020; 58(8):e00772-20. https://doi.org/10.1128/JCM.00772-20