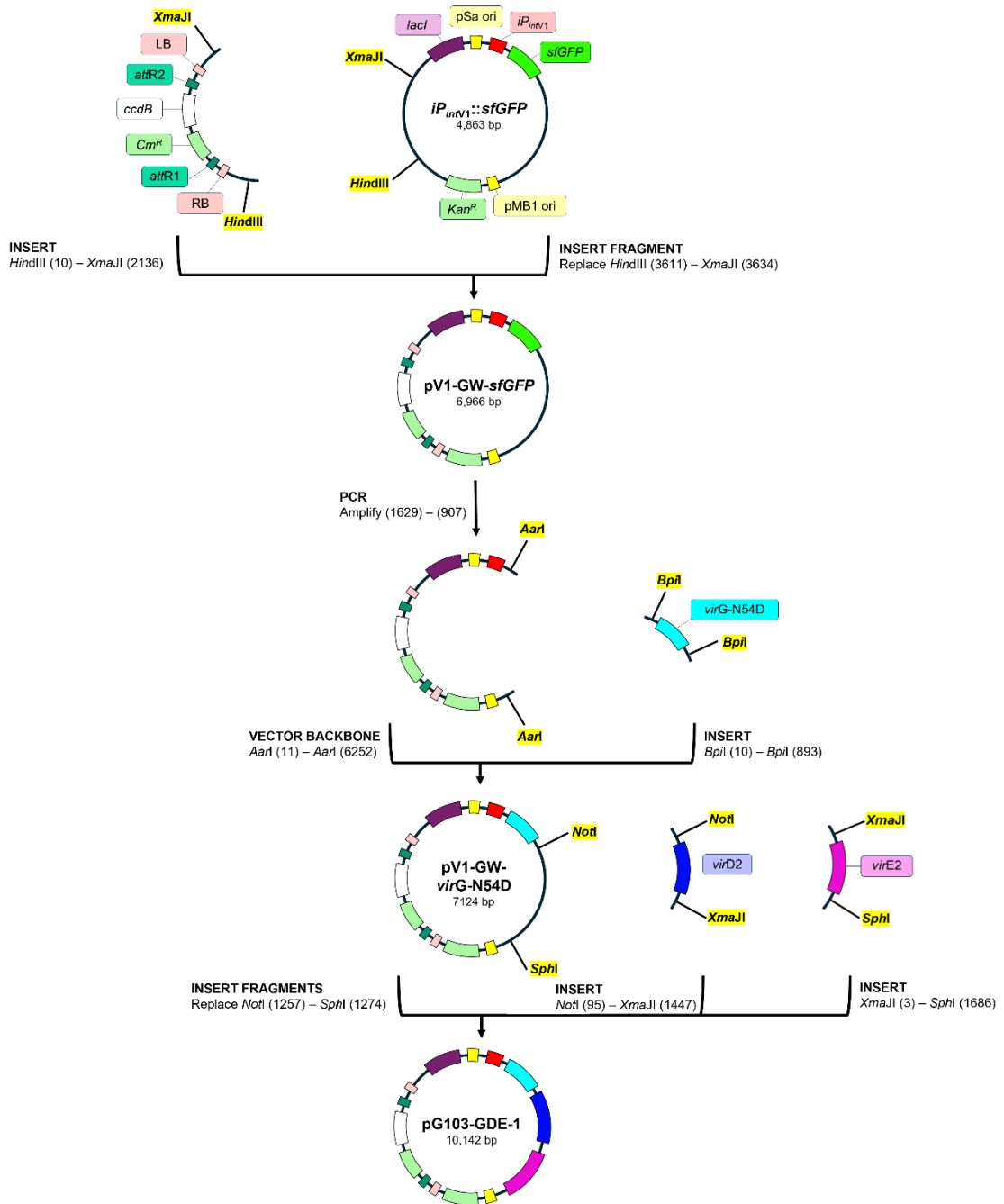


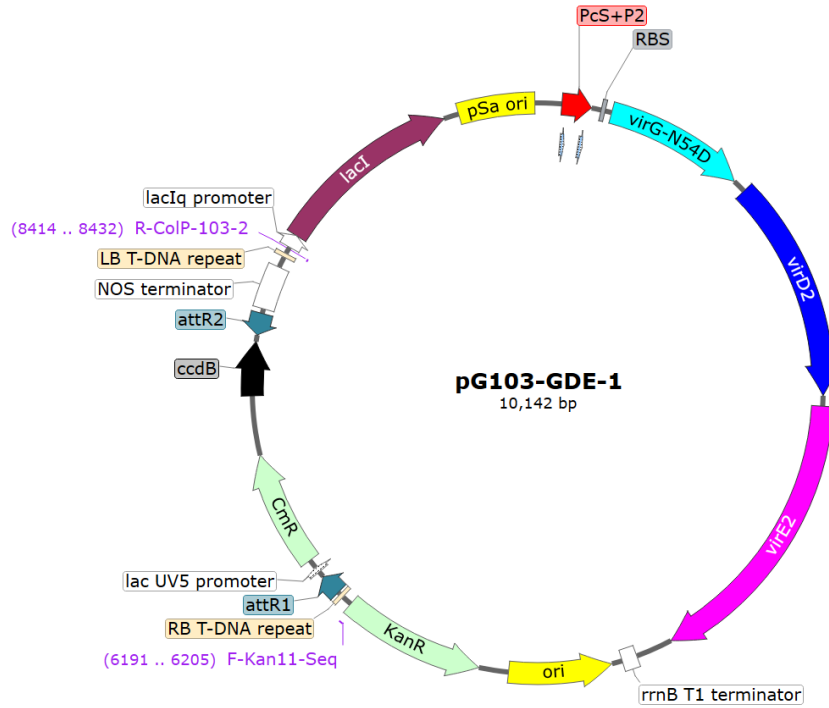
1 SUPPLEMENTARY MATERIALS



2

3 Figure S1: Stepwise construction of the binary vector pG103-GDE-1 from *ip_{intV1}::sfGFP*.

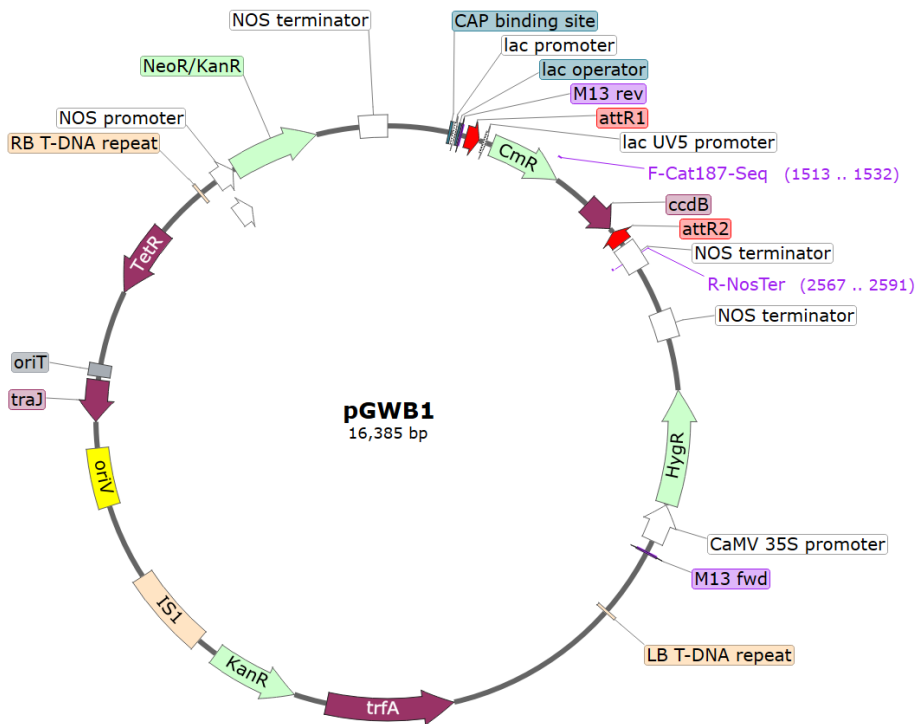
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6 Figure S2: Plasmid map of the unmodified pG103-GDE-1 binary vector. The positions of the
 7 primer pair for PCR analysis to confirm the presence of the plasmid in *A. tumefaciens* (F-
 8 Kan11-Seq and R-ColP-103-2) are indicated.

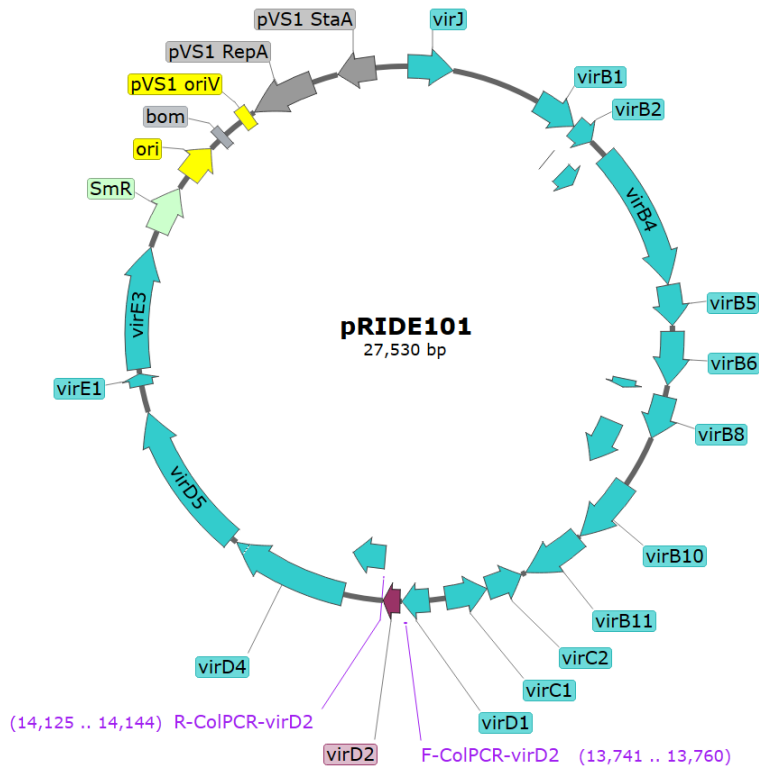
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11 Figure S3: Plasmid map of the unmodified pGWB1 binary vector. The positions of the primer
 12 pair for PCR analysis to confirm the presence of the plasmid in *A. tumefaciens* (F-Cat187-Seq
 13 and R-NosTer) are indicated.

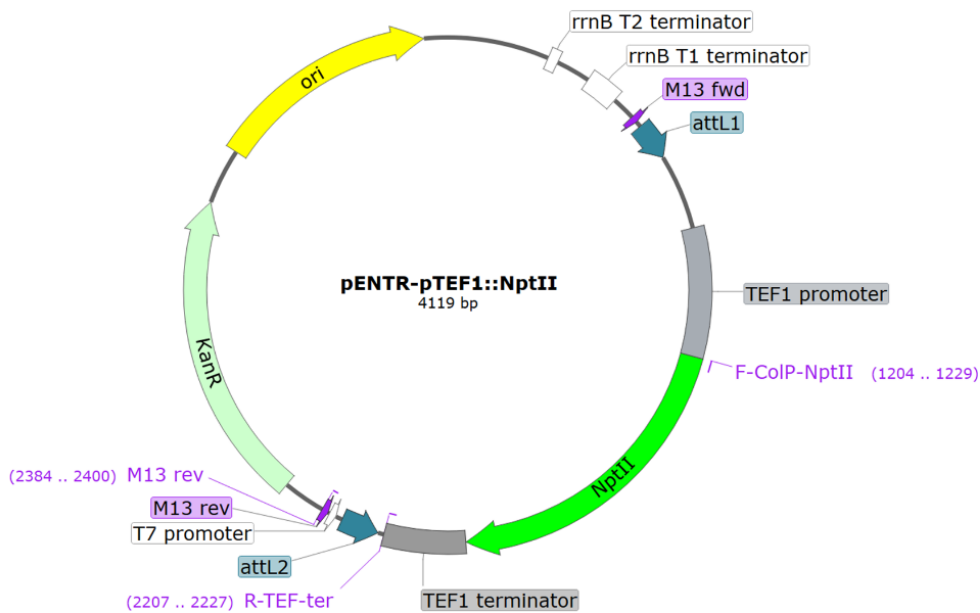
14



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16 Figure S4: Plasmid map of the pRIDE101. The positions of the primer pair for PCR analysis
 17 to confirm the presence of the plasmid in *A. tumefaciens* (F-ColPCR-virD2 and R-ColPCR-
 18 virD2) are indicated.

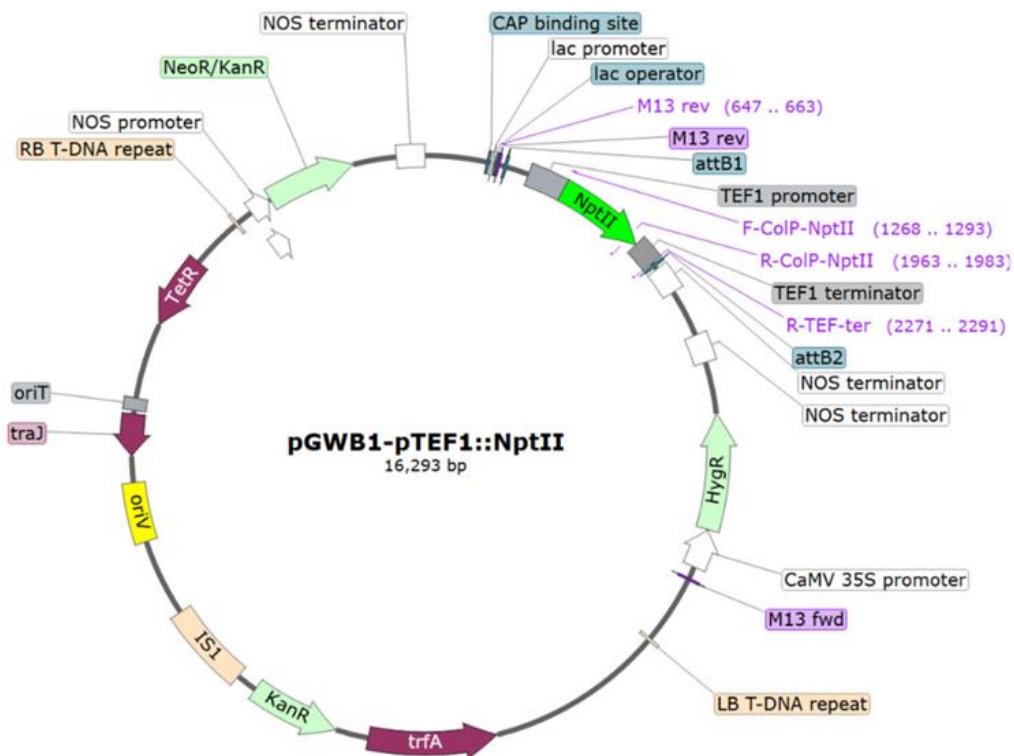
19



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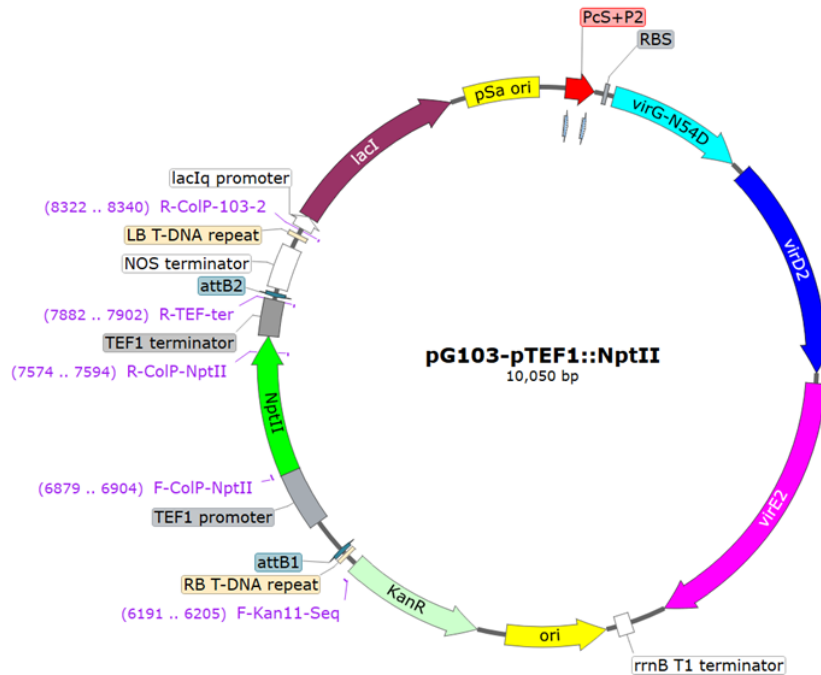
21 Figure S5: Plasmid map of entry vector pENTR- $P_{TEF1}::NptII$. The positions of the gene-
 22 specific primer pair (F-ColP-NptII and R-TEF-ter) and the orientation-specific primer pair
 23 (F-ColP-NptII and M13 rev) used for the colony PCR analyses are indicated.

24



25

26 Figure S6: Plasmid map of binary vector pGWB1- $P_{TEF1}::NptII$. The positions of the gene-
 27 specific primer pair (F-ColP-NptII and R-TEF-ter) and the orientation-specific primer pair
 28 (M13 rev and R-ColP-NptII) used for the colony PCR analyses are indicated.
 29



30

31 Figure S7: Plasmid map of binary vector pG103- $P_{TEF1}::NptII$. The positions of the gene-
 32 specific primer pair (F-ColP-NptII and R-TEF-ter) and the orientation-specific primer pair (F-
 33 ColP-NptII and R-ColP-103-2) used for the colony PCR analyses are indicated.

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35 Table S1: List of primers used in this study.

Primer name	Sequence (5' → 3')	Annotation
F-TEF-NptII	CACCGGTGATGACGGTGAAAACC	Amplification of <i>P_{TEF1}::NptII</i> , 4 base pair sequences (CACC-) necessary for directional cloning were included at the 5' end
R-TEF-ter	CGAATCGACAGCAGTATAGCG	Amplification of <i>P_{TEF1}::NptII</i>
F-ColP-NptII	ATGGGTAAGGAAAAGACTCACGTTTC	Colony PCR and PCR analysis on <i>S. cerevisiae</i> putative transformants; plasmid-specific gene marker
R-ColP-NptII	GGAGAAAACCTACCGAGGCAG	Colony PCR and PCR analysis on <i>S. cerevisiae</i> putative transformants; plasmid-specific gene marker
M13 rev	CAGGAAACAGCTATGAC	Colony PCR; plasmid-specific gene marker
F-Kan11-Seq	CCTCGAGCAAGACGTTTCC	Colony PCR; plasmid-specific gene marker
R-ColP-103-2	GTCGGGAAGATGCGTGATC	Colony PCR; plasmid-specific gene marker
ITS3	GCATCGATGAAGAACGCAGC	Fungal universal primer
ITS4	TCCTCCGCTTATTGATATGC	Fungal universal primer
F-Cat187-Seq	GGCGATTCAGGTTTCATCATG	Plasmid-specific gene marker
R-NosTer	CCATCTCATAAATAACGTCATGCAT	Plasmid-specific gene marker
F-ColPCR-virD2	CGCTCCATTTTGTCCGTATC	Plasmid-specific gene marker
R-ColPCR-virD2	CCATTATTCGGTCCTTCCTG	Plasmid-specific gene marker

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38 Table S2: Recipes for preparation of stock solutions of IM.

Stock solution	Composition
K (pH 4.8)	170.2 g/L Potassium dihydrogen phosphate 217.8 g/L Dipotassium phosphate
M-N	30g/L Magnesium sulfate heptahydrate 15 g/L Sodium chloride
Microspores	100 mg/L Sodium molybdate 100 mg/L Manganese (II) sulfate monohydrate 100 mg/L Zinc sulfate heptahydrate 100 mg/L Copper (II) sulfate pentahydrate 100 mg/L Boric acid
Fe ²⁺	100 mg/L Iron (II) sulfate heptahydrate
Ca ²⁺	10 g/L Calcium chloride dihydrate
NH ₄ ⁺	200 g/L Ammonium nitrate
MES (pH 5.5)	195.2 g/L 2-Morpholinoethanesulphonic acid (MES)
Glycerol	87% (v/v) Glycerol
Glucose	200 g/L Glucose
Adenine	2 g/L Adenine sulfate

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40 Table S3: Recipe for preparation of IM.

Stock solution	Amount required per 1 L of IM (mL)
K (pH 4.8)	0.8
M-N	20
Microspores	0.5
Fe ²⁺	1
Ca ²⁺	1
NH ₄ ⁺	2.5
MES (pH 5.5)	40
Glycerol	5.7
Distilled water	908.5
Glucose	10
Adenine sulfate	10

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