

## EFFECT OF CROSSLINKING TO THE MECHANICAL PROPERTY OF APATITE GELATIN HYBRID FOR BONE SUBSTITUTION PURPOSES

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Received May 18, 2011; Accepted November 21, 2011

### ABSTRACT

The clinical success of current generation of synthetic bone substitute relies on bio-inspired design which has a performance level close to that of natural one. In this context, biomedical approaches are considered very important to result bio-functional hybrid for bone substitution purposes. In this study, effect of cross-linking to the mechanical properties of apatite gelatin hybrid has been investigated. Cross-linking was employed by 1-ethyl-3-(3-dimethylaminopropyl carbodiimide (EDC) agent. The EDC agent creates a peptide bond between gelatin molecules inside the hybrid to the cross-linked structure. Cross-linked structure of gelatin increases physical property of the hybrid since it can hold the outer forces longer than that of without cross-linking.

**Keywords:** cross-linking, hybrid, carbonated hydroxyapatite, gelatin, physical property

### INTRODUCTION

Apatite gelatin hybrid has been widely used for bone substituting purposes since it has similar composition with human bone [1-2]. However, the apatite gelatin hybrid has some drawbacks in term of mechanical properties when the hybrid is applied in real physiological conditions. It is because non-cross-linked collagen-based hybrid materials are rapidly degraded *in vivo*. Cross-linking is used to control mechanical properties [3] and the durability of the materials by interconnecting the gelatin network chemically.

Cross-linking process results in several biochemical and structural modifications (such as a decreased antigenicity, increased mechanical property, reduced solubility, and a reduced rate of biodegradability) which are desirable in a surgical prosthesis [4-6]. Two types of cross-linking procedures have been developed: physical treatments, such as heat, ultra-violet, and gamma irradiation as well as chemical treatments [7]. In this study, 1-ethyl-3-(3-dimethylaminopropyl carbodiimide (EDC) was used to cross-link the apatite gelatin hybrid. The EDC is a zero-length cross-linking type agent which creates a peptide bond between carboxyl and an amine group of gelatin molecules without the introduction of 'foreign' molecules in the network [8]. This agent is also known as a non-

toxic and biocompatible cross-linking agent [9]. Therefore, theoretically it is not harmful when implanted into human body.

In order to increase mechanical properties of gelatin, some researchers cross-linked the hybrid by immersing the molded hybrid to the EDC solution [10- 11]. This method will create cross-linked network only in the surface of the hybrid. In view of this, the problem of partial cross-linked network will be solved by modifying the previous cross-linking method with a better method in this research. In the new proposed method done in this research, cross-linking was done by adding EDC solution when the hybrid was still in suspension state to enhance effective cross-linking process of all gelatin molecules.

### EXPERIMENTAL SECTION

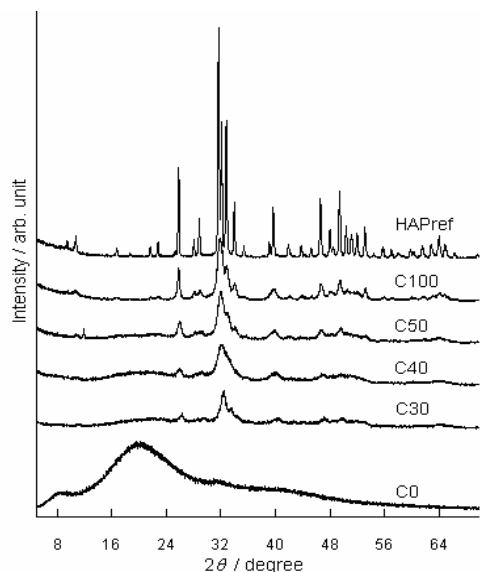
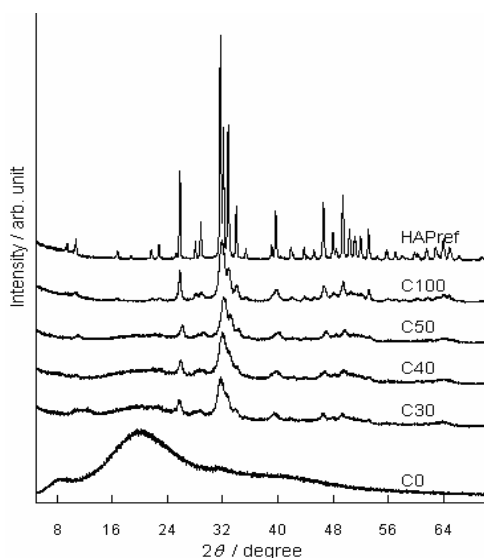
#### Materials

Starting materials to produce apatite gelatin hybrid were Ca(OH)<sub>2</sub> (Wako Chemical, Japan), 85%w/v of H<sub>3</sub>PO<sub>4</sub> (Wako Chemical, Japan), porcine gelatin with IEP 4.0 (Sigma Aldrich, USA), and EDC or 1-ethyl-3-(3-dimethylaminopropyl carbodiimide (Dojindo, Japan). The ratio of apatite to solid content is described on Table 1.

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**Table 1.** Ratio of apatite to solid content and its preparations

Sample	Gelatin (g)	Ca(OH) <sub>2</sub> (g)	H <sub>3</sub> PO <sub>4</sub> (mL)
Gelatin	7.50	0.00	0.00
30% CO <sub>3</sub> <sup>2-</sup> -HAP	5.25	1.67	0.91
40% CO <sub>3</sub> <sup>2-</sup> -HAP	4.50	2.22	1.21
50% CO <sub>3</sub> <sup>2-</sup> -HAP	3.75	2.78	1.52
CO <sub>3</sub> <sup>2-</sup> -HAP	0.00	5.55	3.03

**Fig 1.** XRD pattern of non-cross-linked apatite gelatin hybrid**Fig 2.** XRD pattern of cross-linked apatite gelatin hybrid

### Instrumentation

All the specimens were characterized using X-ray Diffraction ([XRD] Rigaku Rint 2000V, Japan). X-Ray

diffraction analysis was made using counter-monochromatic CuK $\alpha$  radiation generated at 40kV and 100mA. Fourier transform infrared spectroscopy ([FT-IR] Spectrum 2000 EX Perkin Elmer) and Scanning Electron Microscopy ([SEM] JEOL JSM-5400LV, Japan) were also used to confirm the XRD results. Compressive strength was used to determine mechanical property of the hybrids and evaluated at a constant crosshead speed of 1mm/min on a universal testing machine (SV-301, Imada, Tokyo, Japan).

### Procedure

Apatite gelatin hybrid was prepared by adding Ca(OH)<sub>2</sub> to the gelatin solution (gelatin was dissolved to 50 mL of water) for 45 min and 45 °C to make Ca(OH)<sub>2</sub>-gelatin system in a magnetic stirrer. H<sub>3</sub>PO<sub>4</sub> (dissolved in 50 mL of water) was then dropped wisely to the Ca(OH)<sub>2</sub>-gelatin system and stirred for 24 h respectively. To crosslink the gelatin molecules, 4 mL of 5%w/v EDC was added to the slurry after it was aged for 24 h, then stirred for 24 h at room temperature. The slurry was then molded into the polymer-mould and put into freezer for 24 h at -17 °C. After freezing, all specimens were freeze-dried for 24 h by freeze-drying machine. The weight ratio of apatite to solid content was varied from 30%, 40%, and 50%w/w with 7.5%w/v solid content (in 100 mL of water). These specimens were then denoted by C30, C40, and C50 respectively. Commercially available hydroxyapatite (HAPref), gelatin (C0), and gelatin-free apatite (C100) were used as control materials.

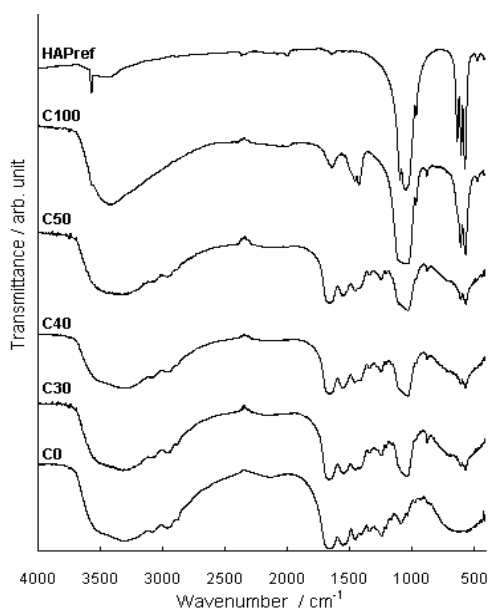
## RESULT AND DISCUSSION

### Characterization of apatite gelatin hybrid

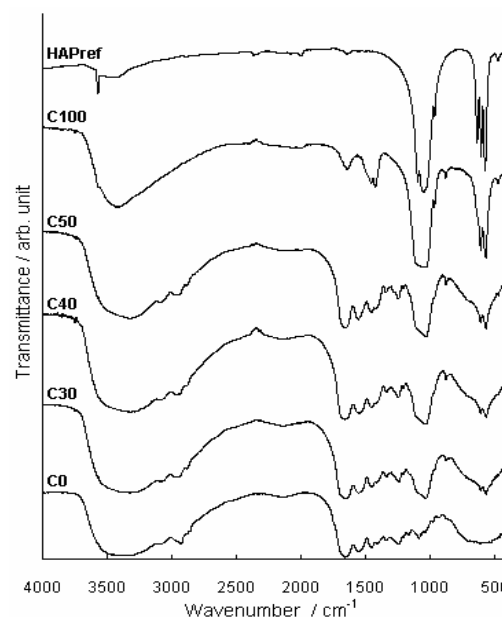
All the specimens, both cross-linked and non-cross-linked hybrids, were characterized using XRD, FTIR, and SEM.

### XRD analysis

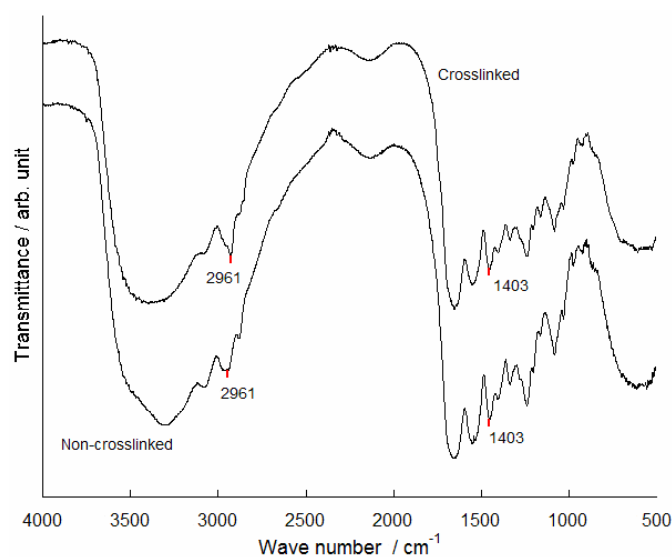
Apatite crystals of hybrid were detected in the XRD pattern of the specimens (Fig. 1 and Fig. 2). Major peaks of apatite were observed at 25.5°, 31°, 32°, and 33°. The X-Ray Diffraction patterns of the hybrids showed low peaks in all specimens (C30, C40, and C50) compared to high crystalline commercially available hydroxyapatite. Therefore, apatite produced by this method has low crystallinity which is similar with human bone. There were no differences between XRD pattern of which produced by both cross-linked hybrid and non-cross-linked hybrid. It has been proven by the results that cross-linking procedure changed only gelatin molecules network, not the apatite crystals.



**Fig 3.** FTIR spectra of non-cross-linked apatite gelatin hybrid



**Fig 4.** FTIR spectra of cross-linked apatite gelatin hybrid



**Fig 5.** The FTIR spectra for gelatin and gelatin after cross-linking. Cross-linked gelatin exhibits similar peaks with non-cross-linked gelatin, but it is observed that after EDC treatment, the symmetric stretching of carboxylate salt ( $1403\text{ cm}^{-1}$ ) was decreased in compared to C-H bond ( $2961\text{ cm}^{-1}$ ), indicating that amidation reaction occurred between carboxyl groups and amino groups. These absorption peaks also confirmed that inter and intra-molecular cross-linking occurred to produce cross-linked gelatin network

#### FTIR analysis

The spectrum of the apatite gelatin hybrid (Fig. 3–5) gave a combination both typical of apatite [12] and

gelatin [13]. The spectra of hydroxyl ion was detected at  $3568\text{ cm}^{-1}$ , while  $\text{CO}_3^{2-}$  ion at  $1455\text{ cm}^{-1}$  and  $1430\text{ cm}^{-1}$ , and  $\text{PO}_4^{3-}$  was detected at  $1300\text{--}900\text{ cm}^{-1}$  and  $570\text{ cm}^{-1}$  respectively [14]. Amide bands from gelatin were observed at  $1500\text{--}1700\text{ cm}^{-1}$  [13].

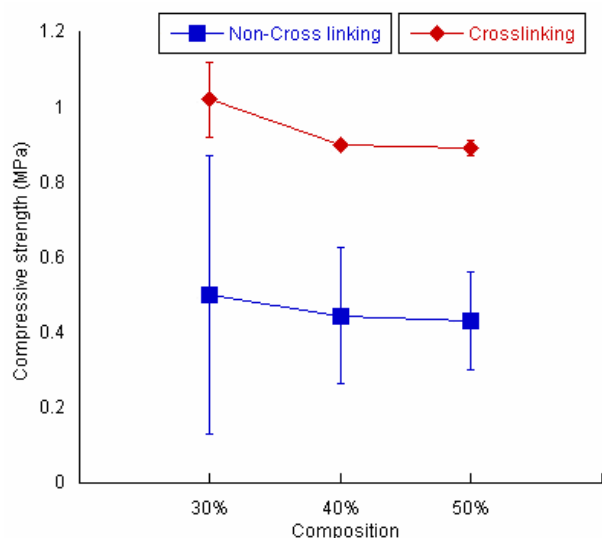
#### Compressive strength

Compressive strength of the cross-linked hybrid was almost two times higher than that of non-cross-linked hybrid (Fig. 5) in all compositions. Specimen C30 which contained the biggest amount of gelatin showed the highest value of compressive strength than the other compositions. Fig. 6 showed that higher amount of gelatin resulted higher value of compressive strength.

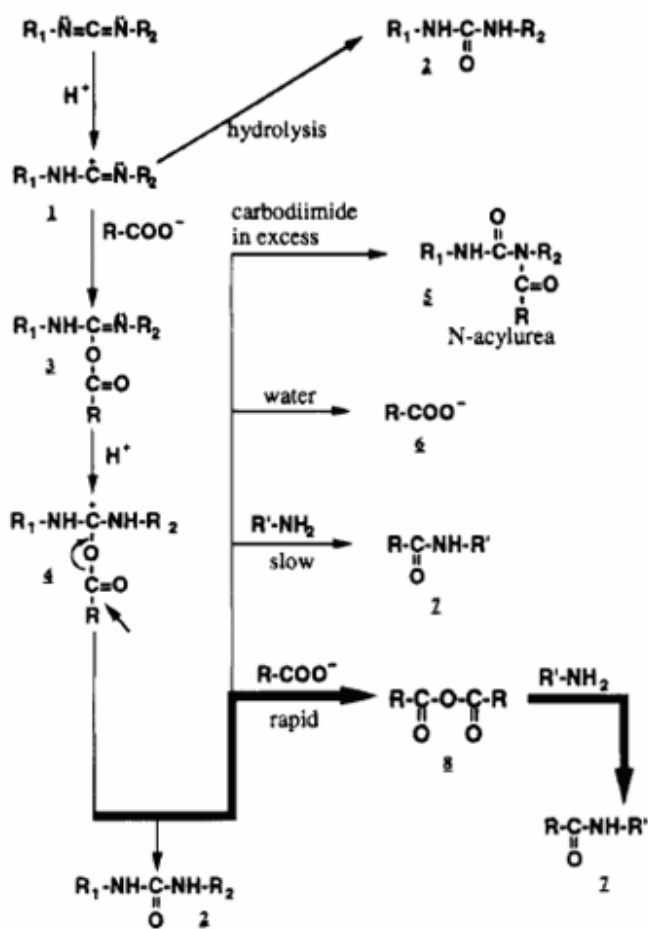
#### Discussion

In this study, compressive strength measurement was used to determine mechanical property of the hybrids. Apatite gelatin hybrid was prepared as load-bearing bone substitute such as skeletal bone etc. As a load bearing bone substitute, it needs high compressive strength in order to be implanted in human body and to physiologically substitute human bone functions.

Compressive strength values of apatite gelatin hybrid (Fig. 6) showed the differences between non-cross-linked hybrid and cross-linked hybrid as expected. In this point it has been proven that cross-linking can significantly increase compressive strength



**Fig 6.** Compressive strength of both non-cross-linked apatite gelatin hybrid and cross-linked apatite gelatin hybrid



**Fig 7.** Proposed reaction mechanisms of the amide formation between carboxylic acid and amine in aqueous media in the presence of carbodiimide [15]

of the hybrid. This preliminary data can be used to control not only mechanical property of hybrid but also its degradation rate in the future since cross-linked network of gelatin is known to be insoluble in water [12]. Cross-linking worked only in gelatin molecules so that the larger amount of gelatin in the hybrid, the higher value of compressive strength was observed. Therefore, in Fig. 6, C30 has higher value of compressive strength than both C40 and C50.

Carboxyl groups (-COOH) and amine groups (-NH<sub>2</sub>) are functional groups which are being target of EDC to create a peptide bond. The bonding changed gelatin networks to be interconnected each other (Fig. 8). The interconnected networks make the possibility for the hybrid to hold the forces worked to its surface longer than that of non-interconnected networks, yielded high compressive strength values. Reaction mechanism of both carboxyl and amine groups in the presence of EDC in the aqueous media is considered very complicated (Fig. 7) since it might result cross-linked networks by rapid mechanism as well as slow mechanism, hydrolysis, and or N-acylurea residues.

Gelatin as a protein has variety of functional groups such as -NH<sub>2</sub>, -OH, and -COOH, which will make the pathway of reaction with carbodiimide very complicated [13]. In the study done by Zeeman et al. [17], EDC known to form intra-molecular cross-links within a gelatin molecule or short-range inter-molecular cross-links between two adjacent gelatin molecules as long as the cross-linked carboxylic acid and amino groups are less than 1.0 nm apart. EDC itself can be hydrolyzed in low pH or in the absence of carboxyl groups [13], producing the corresponding urea derivative which can react with ionized carboxyl group to form product O-acylisourea. Reprotonation of O-acylisourea will change O-acylisourea to be a carbocation that followed by transferring carbocation in absence of nucleophile to form urea derivative. The ionized carboxyl group is a very strong base, so that the reaction with carbocation may produce carboxylic anhydride in the case of cyclizable carboxyl group, which quickly forms the corresponding amide when amine is present. Therefore, in the case of non-cyclizable carboxyl group, carbocation will react with a water molecule or an unionized amine to yield carboxylate or amide respectively.

The complexity of the reaction can explain the disadvantage of EDC as a cross-linking agent, wherein N-acylurea will be a by-product that might be yielded during reaction if EDC is in excessive amount. In this point, N-acylurea is potential to be a toxic compound when implanted to human body. To overcome the problems, there are two solutions proposed, those are by using N-hydroxysuccinimide (NHS) to convert N-acylurea into NHS activated carboxylic acid group which

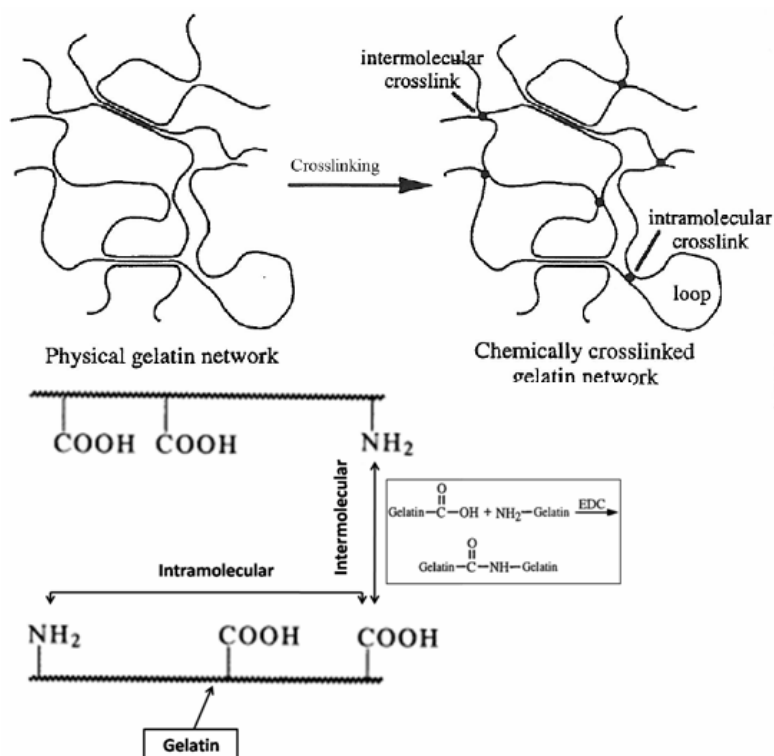


Fig 7. Illustration of the effect of cross-linking on a gelatin network structure [16]

is very reactive towards amine groups of (hydroxyl)lysine, producing cross-linking structure product (No. 7, Fig. 7) or making the amount of EDC less than gelatin so that N-acylurea will not be produced during reactions. Therefore, the understanding of cross-linking mechanism and the products by cross-linking reaction are very useful to design cross-linked apatite gelatin hybrid by minimizing the risk when implanted to human body in the future.

## CONCLUSION

Cross-linking method which is done during the suspension state of the hybrid fabrication has feasibility to increase mechanical property of apatite gelatin hybrid significantly. In our laboratory now in vitro studies as well as the animal experiments and initial clinical research related to apatite-gelatin hybrids have been being done to understand both physico-chemical and physiological phenomena with regard to the future applications of the hybrids for biomedical purposes.

## ACKNOWLEDGEMENT

This research was part of continuing collaboration between UGM and Kyushu University funded by First Batch of JSPS-DGHE. Sunarso was running some parts of the joint works in Kyushu University funded by

Kyushu-University Friendship Scholarship for Undergraduate Students Exchange in 2008 under the supervision of Kunio Ishikawa and Ika Dewi Ana.

## REFERENCES

1. Chang, M.C., Ko, C.C., and Douglas, W.H., 2003, *Biomaterials*, 24, 2853–2862.
2. Landi, E., Valentini, F., and Tampieri, A., 2008, *Acta Biomater.*, 4, 1620–1626.
3. Kikuchi, M., Matsumoto, H.N., Yamada, T., Koyama, Y., Takakuda, K., and Tanaka, J., 2004, *Biomaterials*, 25, 63–69.
4. Rault, I., Frei, D., Herbage, N., and Hue, A., 1996, *J. Mater. Sci. - Mater. Med.*, 7, 4, 215–222.
5. Sabelman, E.E., 1985, *Biocompatibility of Tissue Analogs*, ed. D.F. Williams, CRC Press, Boca Raton.
6. Simmons, D.F., and Kearney, J.N., 1993, *Biotechnol. Appl. Biochem.*, 17, 23–29.
7. Weadock, K., Olsen, R.M., and Silver, F.H., 1984, *Biomater. Med. Devices Artif. Organs*, 11, 4, 293–318.
8. Young, S., Wong, M., Tabata, Y., and Mikos, A.G., 2005, *J. Controlled Release*, 109, 256–274.
9. Chang, M.C., and Douglas, W.H., 2007, *J. Mater. Sci. - Mater. Med.*, 18, 2045–2051.

10. Narbat, M.K., Orang, F., Hashtjin, M.S., and Goudarzi, A., 2006, *Iran Biomed. J.*, 10, 4, 215–223.
11. Kim, H.W., Knowles, J.C., and Kim, H.E., 2005, *J. Biomed. Mater. Res. Part B*, 74B, 686–698.
12. Krajewski, A., Mazzocchi, M., Buldini, P.L., Ravaglioli, A., Tinti, A., Taddei, P., and Fagnano, C., 2005, *J. Mol. Struct.*, 744-747, 221–228.
13. Chang, M.C., 2008, *J. Mater. Sci. - Mater. Med.*, 19, 3411–3418.
14. Aizawa, M., Ueno, H., Itatani, K., and Okada, I., 2005, *J. Eur. Ceram. Soc.*, 26, 4-5, 501–507.
15. Nakajima, N., and Ikada, Y., 1995, *Bioconjugate Chem.*, 6, 1, 123–130.
16. Kuijpers, A.J., Engbers, G.H.M., Feijen, J., De Smedt, S.C., Meyvis, T.K.L., Demeester, J., Krijgsveld, J., Zaat, S.A.J., and Dankert, J., 1999, *Macromolecules*, 32, 3325–3333.
17. Zeeman, R., Dijkstra, P.J., Wachem, P.B.V., Luyn, M.J.A.V., Hendriks, M., Cahalan, P.T., and Feijen, J., 1999, *Biomaterials*, 20, 921–931.