

## BENZYLIDENE CYCLOPENTANONE DERIVATIVES AS INHIBITORS OF RAT LIVER GLUTATHIONE S-TRANSFERASE ACTIVITIES

Sudibyo Martono

Laboratory of Pharmaceutical Chemistry, Faculty of Pharmacy,  
Gadjah Mada University, Yogyakarta, Indonesia

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### ABSTRACT

The effect of the curcumin analogues, 2,6-bis-(4-hydroxy-3-methoxy benzylidene) cyclopentanone (B1) and two of its derivatives on  $\mu$  class glutathione S-transferases (GSTs) from phenobarbital-induced and uninduced rat liver cytosol has been studied to elucidate their anti-inflammatory activity. GST activity was monitored spectrophotometrically using 1,2-dichloro-4-nitrobenzene. B1 was the most potent inhibitor of GSTs, both in uninduced and in phenobarbital-induced rat liver cytosol. These inhibitory properties might be explained as part of the anti-inflammatory activity of benzylidene cyclopentanone derivatives (B1 and B12).

**Keywords:** curcumin; benzylidene cyclopentanone; inhibitory potency; glutathione S-transferases mesoporous

### INTRODUCTION

Non-steroid anti-inflammatory drugs (NSAIDs) are active through inhibition of glutathione S-transferases (GSTs). Curcumin is an established NSAID [1], but there is potential for improved products with higher potency and stability based on synthetic analogues. Curcumin [*bis*-(4-hydroxy-3-methoxy phenyl)-1,6-diene-3,5-dione] (Figure 1) is a yellow dye derived from the rhizome of *Curcuma longa* L [2] and is relatively unstable in phosphate buffer at pH 7.4 [3]. However, the stability of curcumin can be improved by lowering the pH or by adding glutathione, N-acetyl L-cysteine, ascorbic acid, rat liver microsomes or rat liver cytosol [4]. Photosensitivity of curcumin has been reported by Tonnesen *et al.* [5], and Dahl *et al.* [6].

Glutathione S-transferases are a family of multifunctional isoenzymes that catalyze the conjugation of glutathione (GSH) with electrophilic compounds, thereby neutralizing their electrophilic sites and rendering the products more water-

soluble. This results in the protection of cellular macromolecules from xenobiotics [7]. Another activity of GSTs is the conversion of leukotriene A<sub>4</sub> to leukotriene C<sub>4</sub> [8]. Leukotrienes are oxidation products of arachidonic acid metabolism *via* the lipoxygenase pathway [9].

Inhibition of GSTs affects a large number of endogenous processes. For example, the anti-inflammatory drug sulfasalazine inhibits formation of leukotriene C<sub>4</sub>, by inhibiting both leukotriene C synthase and several GST isoenzymes [10]. Another anti-inflammatory drug, indomethacin, is a moderate inhibitor of  $\alpha$ ,  $\mu$ , and  $\pi$  GSTs [11; 12]. Curcumin strongly inhibits leukotriene B<sub>4</sub> formation in rat peritoneal polymorphonuclear neutrophils [13] due to inhibition of lipoxygenase and cyclooxygenase activity [14] and also inhibition of  $\alpha$ ,  $\mu$ , and  $\pi$  GST activity [4; 15; 16] as measured with 1-chloro-2,4-dinitrobenzene (CDNB). During inflammation, GSTs are also involved in the formation of inflammation mediators such as prostaglandin [17] and leukotriene [8].

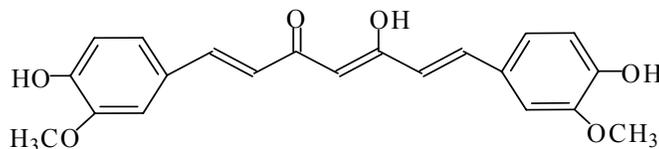
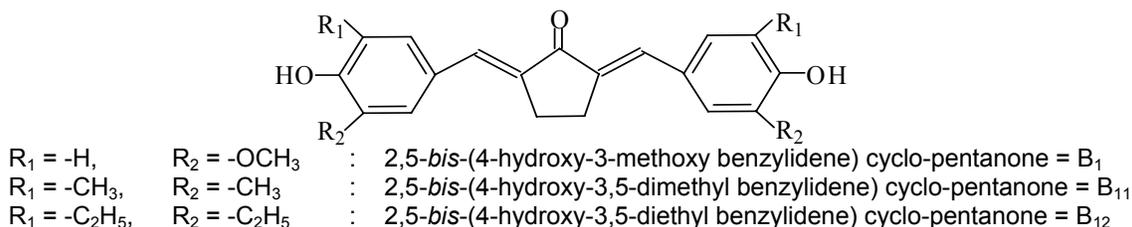


Fig 1 Curcumin [*bis*-(4-hydroxy-3-methoxy phenyl)-1,6-diene-3,5-dione]



**Fig 2** Structures analogues of curcumin

In a preliminary study, analogues of curcumin, such as 2,5-bis-(4-hydroxy-3-methoxy benzylidene) cyclopentanone were about 6 times more potent than curcumin in inhibiting rat liver cytosolic GSTs. In these experiments, 1,2-dichloro-4-nitrobenzene (DCNB) was used as a specific substrate for  $\mu$  class GSTs. Benzylidene cyclopentanone (Figure 2) is a known anti-inflammatory compound [18] and so we synthesized 2,5-bis-(4-hydroxy-3-methoxy benzylidene) cyclopentanone and its derivatives by changing the acetyl acetone group in the centre of the curcumin molecule with a cyclopentanone group [19]. This paper presents the results of inhibition studies of the compounds on GST activity.

## EXPERIMENTAL SECTION

### Materials

1,2-Dichloro-4-nitrobenzene (CDNB) was obtained from Aldrich Chemie (Beerse, Belgium), reduced glutathione (GSH), and bovine serum albumin were purchased from Sigma Chemical Co. (St Louis, MO). All solvents and buffer components used were obtained from E. Merck (Darmstadt, Germany). Curcumin was synthesized according to Pabon [20]. 2,5-bis-(4-hydroxy-3-methoxy benzylidene) cyclopentanone (B<sub>1</sub>), 2,5-bis-(4-hydroxy-3,5-dimethyl benzylidene) cyclopentanone (B<sub>11</sub>) and 2,5-bis-(4-hydroxy-3,5-diethyl benzylidene) cyclopentanone (B<sub>12</sub>) (Figure 2) were kindly given by Dr. Sardjiman, from the Faculty of Pharmacy, Gadjah Mada University, Yogyakarta, Indonesia. Stock solutions of curcumin and its analogues were prepared at 10 mM in dimethylsulfoxide (DMSO). These were diluted in ethanol before use to give the appropriate working concentrations.

### Inhibition Studies

Male Wistar rats (210-230 g) from the laboratory of Pharmacology and Toxicology, Faculty of Pharmacy, Gadjah Mada University, Yogyakarta, Indonesia, were used to obtain GSTs following treatment with or without phenobarbital. GSTs were isolated from uninduced and phenobarbital-induced rat liver cytosol [4] and then stored at -80 °C until

use. Protein concentrations were determined using a bovine serum albumin standard [21].

GST activity was measured spectrophotometrically using DCNB [22]. The reaction mixtures contained 17.5  $\mu$ L of liver cytosol from either rats pretreated with phenobarbital, or from untreated rats, along with 10.0  $\mu$ L of 50 mM ethanolic DCNB, and 75  $\mu$ L of 50 mM GSH. The mixtures was made up to 750  $\mu$ L using 0.1 M potassium phosphate buffer pH 7.5. In inhibition studies, 7.5  $\mu$ L samples of inhibitors in ethanol were preincubated for 4 min before GSH and DCNB were added. In the kinetic studies, the following concentration of DCNB in the final incubation mixtures were used: 1; 0.67; 0.50; 0.40; 0.33; and 0.285 mM. All of the concentrations of inhibitor studied were kept lower than 50  $\mu$ M in the final incubation mixture due to their low water solubility. All treatments were replicated 4 times.

### Analysis

IC<sub>50</sub> values (concentration of inhibitor resulting in a 50 % reduction of GST-activity) were obtained from regression analysis of concentration of inhibitor vs % of inhibition at fixed concentrations of DCNB (0.67 mM) or GSH (5 mM). The values of V<sub>max</sub>, K<sub>m</sub>, K<sub>i</sub> and the type of inhibition were determined according to [23] and [24].

## RESULTS AND DISCUSSION

Inhibitory effects were strongly correlated with concentration of curcumin and the analogues B<sub>1</sub> and B<sub>12</sub> ( $p > 0.001$ ;  $r > 0.94$ ) for GSTs from both uninduced (GST-N) and phenobarbital-induced (GST-PB) rat liver cytosol. For B<sub>11</sub>, there was no strong correlation between concentration and inhibitory effect. However, there was some inhibitory activity of B<sub>11</sub> on both GSTs from uninduced and phenobarbital-induced rat liver. At an inhibitor concentration of 10  $\mu$ M, the relative inhibitory effects were 53.6% (for curcumin), 79.9% (for B<sub>1</sub>), 25.6% (for B<sub>11</sub>), and 26.9% (for B<sub>12</sub>) relative to controls.

The IC<sub>50</sub> values of curcumin, B<sub>1</sub> and B<sub>12</sub> were 9.6, 2.7 and 43.5  $\mu$ M (for uninduced GST), while

corresponding IC<sub>50</sub> values for phenobarbital-induced rat liver were 12.2, 2.5 and 54.7  $\mu$ M respectively. These results showed that compound B<sub>1</sub> was the most potent inhibitor of GST activity.

In the presence of curcumin and B<sub>1</sub>, all V<sub>max</sub> values decreased compared to controls, while K<sub>m</sub> values increased (Tables I and II). For compound B<sub>12</sub>, the V<sub>max</sub> values also decreased in the presence of inhibitor, but the K<sub>m</sub> value decreased only slightly upon the addition of a low concentration of B<sub>12</sub>. The K<sub>m</sub> value increased when a higher concentration of B<sub>12</sub> (Table III) was added.

Inhibition of GSTs by curcumin and analogues B<sub>1</sub> and B<sub>12</sub> have been shown here for the first time. Other plant phenols [25] and polyphenols [26] have been shown to be inhibitors of GSTs and they share a phenolic structure with the three compounds showing inhibitory activity here. From a therapeutic point of view, inhibition of GST activity is important as these enzymes are involved in drug resistance and in the biosynthesis of prostaglandin and

leucotriene from arachidonic acid [8]. Modulation of GST-activity can be used for controlling such compounds [27]. The use of a GST inhibitor, together with a cytostatic drug, is a strategy for increasing therapeutic efficiency of the latter, as some tumors show increasing GSH concentration and GST activity [28]. A role of GST in the resistance to alkylating agents has also been shown by the use of GST inhibitors. Inhibition of GST would thus be potentially beneficial in the treatment of tumors [29]. Tew *et al.* [30] used ethacrynic acid and piroprost as GST inhibitors to enhance the cytotoxic action of alkylating agents in drug resistance and sensitive cell lines. Further, ethacrynic acid treatment of patients with advanced cancer inhibited cellular GST-activity and increased the plasma levels of the alkylating agent thiopeta [31]. Another inhibitor of GST, indomethacin, also acts as an anti-inflammatory drug and potentiates the cytotoxicity of chlorambucil in CHO cells resistance to nitrogen mustards [11].

**Table 1** V<sub>max</sub>, K<sub>m</sub> values in DCNB-GSH conjugation catalyzed by GST-N and GST-PB in the presence of curcumin

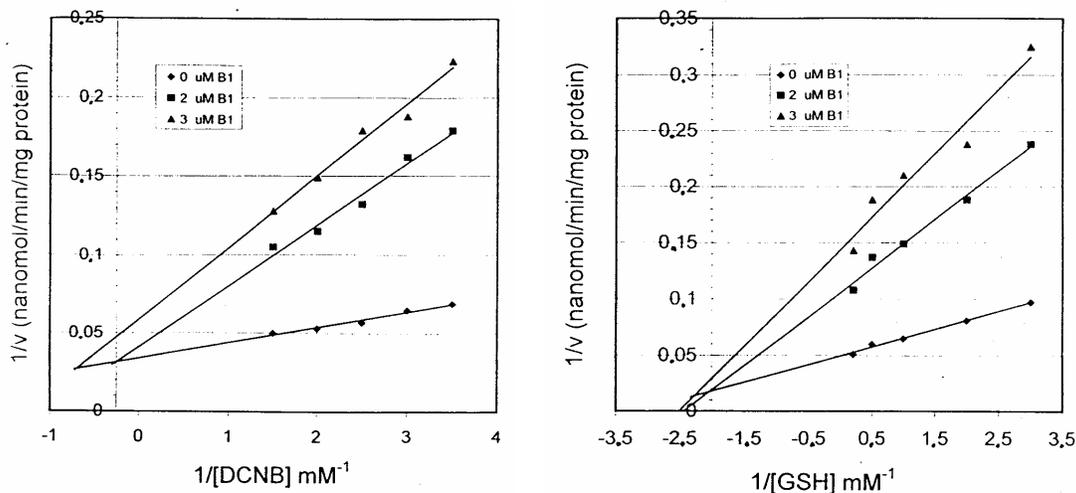
GST	Curcumin ( $\mu$ M)	GSH constant at 5 mM; DCNB concentration range 0.29-0.67 mM		Curcumin ( $\mu$ M)	DCNB constant at 0.67 mM; GSH concentration range 0.2 – 1.0 mM	
		V <sub>max</sub> (nanomol/min/mg)	K <sub>m</sub> (mM)		V <sub>max</sub> (nanomol/min/mg)	K <sub>m</sub> (mM)
GST-N	0	41.75	0.65	0	20.15	0.31
	5	19.10	0.64	5	10.04	0.54
	10	18.90	0.99	10	7.81	0.69
GST-PB	0	72.94	0.43	0	42.25	0.29
	5	56.69	1.36	5	21.70	0.60
	10	68.48	2.38	10	16.50	0.50

**Table 2** V<sub>max</sub>, K<sub>m</sub> values in DCNB-GSH conjugation catalyzed by GST-N and GST-PB in the presence of analogue B<sub>1</sub>

GST	B <sub>1</sub> ( $\mu$ M)	GSH constant at 5 mM; DCNB concentration range 0.29-0.67 mM		B <sub>1</sub> ( $\mu$ M)	DCNB constant at 0.67 mM; GSH concentration range 0.2 – 1.0 mM	
		V <sub>max</sub> (nanomol/min/mg)	K <sub>m</sub> (mM)		V <sub>max</sub> (nanomol/min/mg)	K <sub>m</sub> (mM)
GST-N	0	41.75	0.65	0	20.15	0.31
	2	23.54	0.90	2	9.40	0.40
	3	16.85	0.77	3	6.93	0.39
GST-PB	0	72.94	0.43	0	42.25	0.29
	1	86.09	1.38	1	28,32	0.78
	2	50.16	1.26	2	13,74	0.37

**Table 3**  $V_{max}$ ,  $K_m$  values in DCNB-GSH conjugation catalyzed by GST-N and GST-PB in the presence of analogue  $B_{12}$ 

GST	$B_{12}$ ( $\mu$ M)	GSH constant at 5 mM; DCNB concentration range 0.29-0.67 mM		$B_{12}$ ( $\mu$ M)	DCNB constant at 0.67 mM; GSH concentration range 0.2 – 1.0 mM	
		$V_{max}$ (nanomol/min/mg)	$K_m$ (mM)		$V_{max}$ (nanomol/min/mg)	$K_m$ (mM)
GST-N	0	41.75	0.65	0	20.15	0.31
	20	23.27	0.41	20	16.39	0.31
	40	26.84	0.85	40	11.98	0.52
<b>GST-PB</b>	Experiments for $K_1$ were not performed because the $IC_{50}$ could not be obtained					

**Fig 3** Lineweaver-Burk plot Showing mixed-type inhibition of uninduced rat liver GST's toward DCNB by 2 (■) or 3  $\mu$ M (▲) 2,5-bis-(4-hydroxy-3-methoxy benzylidene) cyclopentanone (left) of non-competitive inhibition towards glutathione (right). Control (◆). The values are the average of four incubations.

In the results presented here (Fig 3), curcumin,  $B_1$  and  $B_{12}$  showed a mixed type inhibition [23] with respect to DCNB. Curcumin at low concentrations, showed non-competitive inhibition of uninduced GSTs, while at low concentrations of  $B_1$ , competitive inhibition was observed for phenobarbital-induced GSTs. In comparison, with respect to GSH, curcumin showed a mixed type inhibition both on uninduced and phenobarbital-induced GSTs. Both  $B_1$  and  $B_{12}$  showed non-competitive inhibition on uninduced GSTs, while mixed type inhibition was observed for phenobarbital-induced GSTs.

## CONCLUSION

From the results, it can be concluded that  $B_1$  is the most potent inhibitor of GSTs both from uninduced and phenobarbital-induced rat liver cytosol, with DCNB representing a substrate for the

$\mu$  class GSTs. These inhibitory properties might be part of the anti-inflammatory activity of curcumin and benzylidene cyclopentanone derivatives ( $B_1$  dan  $B_{12}$ ).

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## REFERENCES

1. Srimal, R.C. and Dhawan, B.N., 1973, *J. Pharm. Pharmacol.*, 25, 447-452.
2. Masuda, T., Jitoe, A., Isobe, J., Nakatani, N., and Yonemori, S., 1993, *Phytochemistry*, 32 (6), 1557-1560.

3. Tonnesen, H.H. and Karlsen, J., 1985., *Z. Lebensm. Unters. Forsch.*, 180, 132-134.
4. Oetari, S., Sudibyo, M., Commandeur J.N.M., Samhoedi, R., and Vermeulen N.P.E., 1996, *Biochem. Pharmacol.*, 51, 39-45.
5. Tonnesen, H.H., Karlsen, J., and van Henegouwen, G.B., 1986, *Z. Lebensm. Unters. Forsch.*, 183, 116-122.
6. Dahl, T.A., Bilski, P., Reszka, K.J., and Chignell, C.F., 1994, *Photochem. Photobiol.*, 59 (3), 290-294.
7. Hsieh, C.H., Liu, L.F., Tsai, S.P., and Tam, M.F., 1999, *Biochem. J.*, 343, 87-93.
8. Timmerman, H., 1997, *New Perspectives for Anti-Inflammatory Drugs*, in : Suwijiyono Pramono (Ed.), Recent development in curcumin pharmacology, *Proceedings of the International Symposium on Curcumin Pharmacology-chemistry* (ISCP), August 29-31, 1995, Yogyakarta, Indonesia, 1-12.
9. Bach, M.K., Brashier, J.R., and Johnson, M.A., 1985, *Biochem. Pharmacol.*, 34, 2695-2704.
10. Hall, A., Robson, C.N., Hickson, I.D., Harris, A.L., Proctor, S.J., and Cattar, A.R., 1989, *Cancer Res.*, 49, 6265-6268.
11. Flatgaard, J.E., Bauer, K.E., and Kaufar, L.M., 1993, *Cancer Chemother. Pharmacol.*, 33, 63-70.
12. Ammon, H.P.T., Anazodo, M.I., Safayhi, H., Dhawan, B.N., and Srimal, R.C., 1992, *Planta Med.*, 58, 226-231.
13. Huang, M.T., Lysz T., Ferraro, T., Abidi, T.F., Laskin, F.A., and Conney A.H., 1991, *Cancer Res.*, 51, 813-819.
14. Sudibyo, M., 1996, *Indon. J. Pharm.*, 7 (1), 39-51.
15. Sudibyo, M., Hakim, L., Samhoedi, M., Commandeur, J.N.M., and Vermeulen, N.P.E., 1997, *Effect of Curcumin and Its Derivatives on Glutathione S-Transferase Activity in Liver Cytosolic of  $\beta$ -Naphthoflavone-Treated Rats*, in: Suwijiyono Pramono (Eds.), Recent Development in Curcumin Pharmacology, *Proceedings of the International Symposium on Curcumin Pharmacology-chemistry* (ISCP), August 29-31, 1995, Yogyakarta, Indonesia, 126-135.
16. Ujihara M, Tsuchida, S, Sato, H and Urade, Y, 1988, *Archs. Biochem. Biophys.*, 264: 428-437.
17. Sardjiman, 2000, *Synthesis of Some New Series of Curcumin Analogues, Antioxidative, Antiinflammatory, Antibacterial Activities and Qualitative Structure-Activity Relationship*, Dissertation, Gadjah Mada University, Yogyakarta, Indonesia.
18. Sardjiman, 1993, *Sintesis 2,6-bis-(3,5-dimetil-4-hidroksibenzilidin) sikloheksanon, 2,5-bis (3,5-dimetil-4-hidroksibenzilidin) siklopentanon dan 1,5-bis (3,5-dimetil-4-hidroksifenil)-1,3-penta-diene-3-on dan daya antioksidannya*, Laporan Penelitian DPP-SPP, Fakultas Farmasi UGM, 1992/1993.
19. Pabon, H.J.J., 1964, *Rec. Trav. Chim. Pays Bas*, 83, 379-386.
20. Bradford, M.M., 1976, *Anal. Biochem.*, 72, 248-254.
21. Habig, W.H., Pabst, M.J., and Jakoby, W.B., 1974, *J. Biol. Chem.*, 249 (22), 7130-7139.
22. Cai, P., Bennet, D., Nair, R.V., Ceska, O., Smith, M.J.A., and Giovanni, J.D., 1993, *Chem. Res. Toxicol.*, 6, 872-879.
23. Metzler, D.E., 1977, *Biochemistry. The Chemical Reactions of Living Cells*, p. 301-318 Academic Press, Inc., New York.
24. Das, M., Bickers, D.R., and Mukthar, H., 1984, *Biochem. Biophys. Res. Commun.*, 120, 427-433.
25. Iio, M., Kawaguchi, H., Sakota, Y., Otonari, J., and Nitahara, H., 1993, *Biosci. Biotech. Biochem.*, 57 (10), 1678-1680.
26. Van Bladeren P.J. and Van Ommen, B., 1991, *Pharmacol. Ther.*, 51, 35-46.
27. Commandeur, J.N.M., Stijntjes G., and Vermeulen N.P.E., 1995, *Pharmacol. Rev.*, 47 (2), 271-330.
28. Ploemen, J.H.T.M., Van Ommen, B., and Van Bladeren, P.J., 1990, *Biochem. Pharmacol.*, 40 (7), 1631-1635.
29. Tew, K.D., Bomber, A.M., and Hoffman, S.J., 1988, *Cancer Res.*, 48, 3622-3625.
30. O'Dwyer, P.J., La Creta, F., Nash, S., Tinsley, P.W., Schilder, R., Clapper, M.L., Tew, K.D., Panting, L., Litwin, S., and Comis, R.L., 1991, *Cancer Res.*, 51, 6059-6065.