

Preparation of a New Cd(II)-Imprinted Polymer and Its Application to Preconcentration and Determination of Cd(II) Ion from Aqueous Solution by SPE-FAAS

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Abstract: A novel ion Imprinted polymer (IIP) material with 8-hydroxyquinoline (8HQ) (the complexing ligands), methacrylic acid (MAA) (the monomers), ethylene glycol dimethacrylate (EGDMA) (the crosslinker agent), benzoyl peroxide (the initiator), and ethanol-acetonitrile (2:1) (the porogen) as adsorbent for the determination of cadmium by solid phase extraction-flame atomic absorption spectrometry (SPE-FAAS) has been synthesized. Synthesis of IIP was done by precipitation polymerization method. The imprinted Cd(II) ions were removed by leaching method using 1 mol L⁻¹ nitric acid. The IIP was characterized by Fourier transform infra-red (FT-IR) spectroscopy and scanning electron microscopy (SEM) to ensure successful synthesis of IIP. The experimental parameters for SPE extraction, such as pH of the sample, loading rate, and elution rate, have been optimized. The optimum pH for quantitative Cd(II) retention was 6, and the elution was completed with 2 mL of 1.0 mol L⁻¹ nitric acid. The optimum loading rate was 0.5 mL min⁻¹. Under optimum conditions, the proposed method with theoretical enrichment factor 50 times has a detection limit of 0.5 µg L⁻¹ and the recovery of 97.75%.

Keywords: cadmium(II); solid phase extraction; ion imprinted polymer; preconcentration

■ INTRODUCTION

Heavy metals are considered to be toxic to the environment. One of the toxic heavy metals is cadmium which is released into the environment by industrial wastewater. The most common industries releasing cadmium in their effluents are electroplating, smelting, alloy manufacturing, pigments, plastic, battery, mining, and refining process [1-2].

Cadmium is a naturally occurring metal and known as an extremely toxic. Cadmium causes nausea, salivation, muscular cramps, chronic pulmonary problems, skeletal deformity, hypertension, proteinuria, kidney stone formation, osteoporosis, non-hypertrophic emphysema,

anemia, eosinophilia, anosmia, chronic rhinitis, and Itai Itai disease. A high percent of accumulated cadmium is accumulated in the kidney and liver. Cadmium(II) ions may replace zinc(II) ions in some enzymes thereby affecting the enzyme activity in biological systems. Cadmium is also classified as a category 1 carcinogen (the human carcinogen) by the International Agency for Research on Cancer (IARC) [3-6].

The presence of cadmium in the waters is generally in a very low concentration, so an adequate analytical method is required. Several analytical techniques including cation exchange resin method [7], the competitive chelation method [8], stripping

voltammetry [9], dialysis [10], ion-selective electrodes [11], and permeation liquid membrane [12] have been used for the determination of cadmium in different matrixes. Most of these methods suffer from chemical interferences, competition effects, disturbance of solution equilibrium, long equilibrium time, poor on limits of detection, poor preconcentration, and poor selectivity [13-15]. Preconcentration techniques have been developed to improve sensitivity. One of them is the solid phase extraction (SPE).

Solid-phase extraction (SPE), also known as solid-liquid extraction is a separation technique that is used to separate the analyte from a mixture, using their physical and chemical properties. The basic principle of SPE can be described as the transfer of analytes from the liquid phase into the solid phase. The analyte after sorption on the solid phase is desorbed with a suitable solvent. The solid phase extraction method is an alternative to replace solvent extraction. Solid phase extraction is a chromatographic sample preparation technique that is used for the separation, purification, and preconcentration of chemical compounds from solution [16-17]. In SPE, the sample is detained or absorbed and later eluted adsorbent with elution in one stage or gradient. There are several advantages when compared with solvent extraction, including low cost, reducing the use of hazardous solvents, decreasing time consumption, absence of emulsion [18-19], easy to storage and transport [20], the desired component can be taken by using a selective solvent [21], and can be used for the separation and purification [22]. One of the new sorbents is an ionic-imprinted polymer (IIP). The ion-imprinting idea is based on the formation of a complex from the metal ion with a ligand and its subsequent immobilization in a cross-linked polymeric matrix. Upon metal ion extraction from the polymeric body spatially pre-arranged binding sites are left in the sorbent structure [23]. The selectivity of IIP is caused by the following factors such as the specificity of interaction of a ligand with the metal ion, the coordination geometry and the coordination number of metal ion, the charge of the metal ion, and the size of the metal ion [19].

Recently, IIP has been growing rapidly as an adsorbent for SPE because it has the advantage on the enrichment factor and high selectivity. Applications of IIP as adsorbent in the SPE has been widely reported to the extraction of arsenic [24], iron [25-26], cadmium [23,27-28], mercury [29], copper [30], lead [31-33], and zinc [34-35].

In this paper, we report the development methods of cadmium preconcentration in SPE. Cadmium concentration was analyzed using flame atomic absorption spectrophotometer (FAAS). The IIP was synthesized using 8-hydroxyquinoline (8HQ) as complexing ligands, methacrylic acid (MAA) as the functional monomer and ethylene glycol dimethacrylate (EGDMA) as agents of cross-linking. Characterization of the IIP was investigated using Fourier transform infrared spectrophotometry (FTIR) and scanning electron microscopy (SEM). The experiment parameter such as pH of the solution, flow rate, volume of HNO₃ for elution was investigated. This method was applied for the enrichment of trace amount of cadmium ion from aqueous solutions.

■ EXPERIMENTAL SECTION

Materials

The materials used were Cd(NO₃)₂·4H₂O (Merck, Germany), HNO₃ (Merck, Germany), 8-hydroxyquinoline (8HQ) (Sigma Aldrich, US), methacrylic acid (MAA) (Sigma Aldrich, US), ethylene glycol dimethacrylate (EGDMA) (Sigma Aldrich, US), benzoyl peroxide (Merck, Germany), ethanol (Merck, Germany), and acetonitrile (Merck, Germany).

Instrumentation

The characteristic of functional groups among adsorbent was confirmed by Fourier transform infrared spectroscopy (Perkin Elmer). The surface morphological image of adsorbent was obtained by scanning electron microscopy (FEL, Inspect-S50). The concentration of cadmium was analyzed by flame atomic absorption spectrophotometer Hitachi type Z-2000. A pH-meter (Mettler Toledo) was used for the pH adjustments.

Procedure

Preparation of IIP

The ionic imprinted polymer was synthesized following the method proposed by Barciela-Alonso et al. [36]. In the glass tube, 0.0309 g (0.1 mmol) $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ and 0.0363 g (0.25 mmol) 8-hydroxyquinoline was mixed in 60 mL of porogen (2:1 ethanol-acetonitrile). This solution was stirred for 30 min with a magnetic stirrer. Then, 0.34 mL (4 mmol) of methacrylic acid (monomer), 0.0485 g (0.2 mmol) of benzoyl peroxide (initiator), and 3.77 mL (20 mmol) of ethylene glycol dimethacrylate (cross-linker) were added, the glass tube purged with nitrogen for 5 min and immediately sealed. Precipitation polymerization was carried out by heating in a water bath at 60 °C for 8 h while stirring with a magnetic stirrer. The polymer obtained was filtered, washed with ethanol, and oven-dried for 2 h at 60 °C. The cadmium ions were leached from the polymer by using 100 mL of 1 mol L⁻¹ HNO_3 in magnetic stirrer for 30 min. The IIP was washed with distilled water, filtered, and dried at 100 °C for 24 h. The same procedure, except the presence of

$\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$, and leaching, was applied for preparing non-imprinted polymer (NIP). The polymer was synthesized by the same procedure, except the presence of 8HQ, $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ and leaching. The 8HQ, MAA, EGDMA, polymer, and IIP were characterized by FTIR. The IIP and NIP were characterized by SEM.

Effect of pH

The effect of solution pH was tested by shaking 0.01 g IIP and 20 mL of the Cd(II) solution at various pH for 10 min. The pH of the sample solutions was varied from pH 4 to 8. The IIP was separated by filtration, and the concentration of Cd(II) in the filtrate was determined by FAAS.

Effect of eluent volume

A cartridge with an i.d. of 5 mm was filled with 0.1 g of dry IIP. 50 mL of 2.5 mg L⁻¹ Cd(II) solution, adjusted to pH 6, was then passed through the column. Then the solution of 1 mol L⁻¹ HNO_3 was passed to desorb Cd(II) from the cartridge. The volume of HNO_3 was varied from 0.5 to 5.0 mL. The eluate that contains Cd(II) was determined by FAAS.

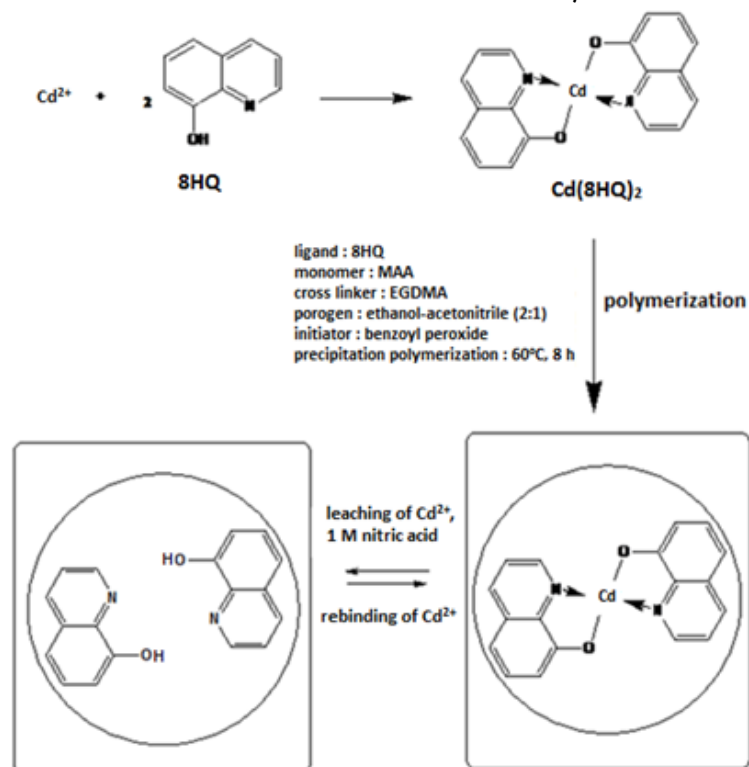


Fig 1. Illustration of IIP formation

Effect of flow rate

A cartridge with an i.d. of 5 mm was filled with 0.1 g of IIP. 50 mL of 2.5 mg L⁻¹ Cd(II) solution, adjusted to pH 6, was passed through the column at various of flow rate. The flow rate was varied from 0.18 to 2.17 mL min⁻¹. Then, Cd(II) was eluted from the column with 2 mL of 1 mol L⁻¹ HNO₃, and concentration of Cd(II) was then determined by FAAS.

Solid phase extraction

A cartridge with an i.d. of 5 mm was filled with 0.1 g of IIP. The sorbent was conditioned by passing 3 mL of buffer pH 6. For the loading of the analyte, the pH of Cd(II) standard solution was adjusted to pH 6 and passed through the column at a flow rate of 0.5 mL min⁻¹. Then, Cd(II) was eluted from the cartridge with 2 mL of 1 mol L⁻¹ HNO₃, and concentration of Cd(II) was determined by FAAS.

RESULTS AND DISCUSSION

The new material was prepared by using 8-hydroxyquinoline (8HQ) as complexing ligands, methacrylic acid (MAA) as the monomer and ethylene glycol dimethacrylate (EGDMA) as agents of cross-linking. The polymer was synthesized by precipitation polymerization technique. The synthesis of IIP was carried out in three step (Fig. 1), i.e., (1) The complexation Cd(II) with 8HQ (2) the copolymerization of monomer (MAA) in the presence of the template (Cd(II)), and (3) the removal of the template after copolymerization from the polymer.

Characterization of IIP

The FTIR spectra of complexing ligand (8HQ), monomer (MAA), crosslinker (EGDMA), polymer, and IIP are presented in Fig. 2. The intense absorption centered at 1724 cm⁻¹ in spectra polymer (Fig. 2(b)) was assigned to carbonyl groups from EGDMA and MAA. The polymer formed by the addition reaction of the C=C double bond to a single C-C bond. It is shown in the FTIR spectra of polymer that is absent a peak at 1524 cm⁻¹ when compared to spectra of MAA (Fig. 2(d)). This confirmed the successful polymerization of the MAA and EGDMA. The intense absorption centered at 1492 cm⁻¹ in spectra

IIP (Fig. 2(a)) was assigned to C=C aromatic groups from 8HQ (Fig. 2(e)). The presence of a C=C aromatic group on the spectra FTIR of IIP when compared with the FTIR spectra of polymer, it shows the successful synthesis of IIP.

Scanning electron microscopy (SEM) images were presented in Fig. 3. The textural characteristics were closely examined at 10,000× magnification. The particles of both NIP and IIP were present in a spherical shape. The SEM images of IIP (Fig. 3(b)) physically look more cavities than NIP (Fig. 3(a)).

Influence of Initial pH on Cd(II) Adsorption

The influence of the initial pH on adsorption of Cd(II) on the IIP was studied to determine the optimum pH. The pH of the solution has been found to be the most

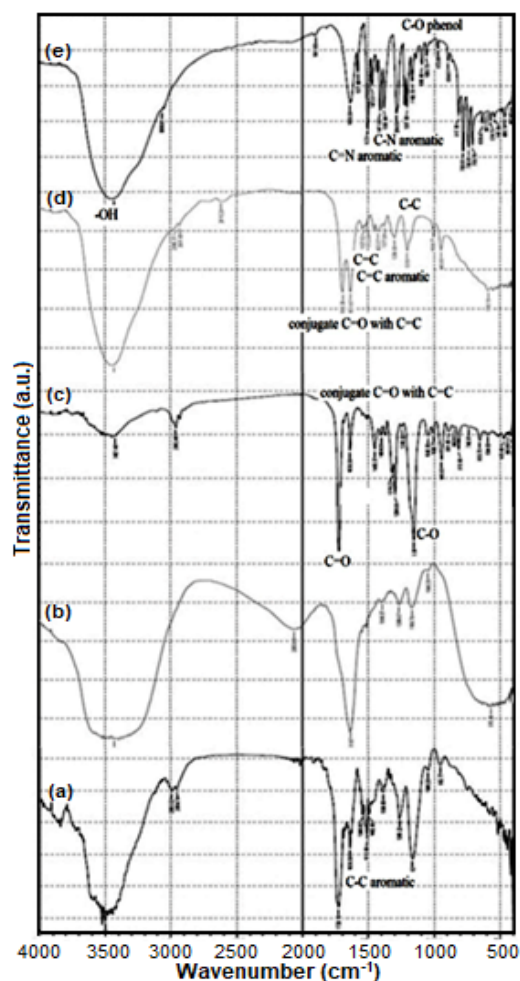


Fig 2. FTIR spectra of (a) IIP, (b) polymer, (c) EGDMA, (d) MAA, and (e) 8HQ

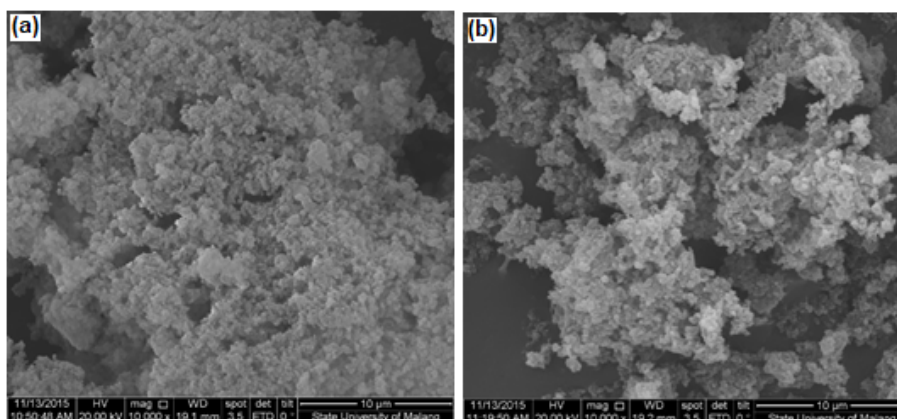


Fig 3. SEM images of (a) NIP and (b) IIP

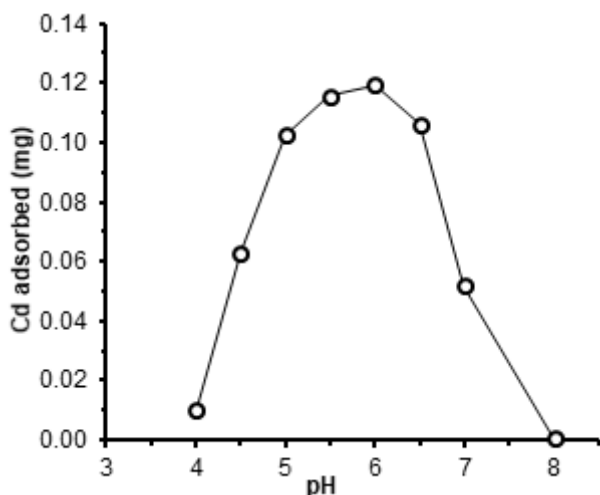


Fig 4. Influence of pH on the adsorption of Cd(II) on IIP

important for adsorption heavy metals onto the adsorbent. It influences the speciation of metal ions and the ionization of functional groups on the adsorbent surface active sites [37-38].

The adsorption of Cd(II) as a function of the pH is shown in Fig. 4. The adsorption of Cd(II) increased greatly between pH 4 to 5.5 and then almost constant from 5.5 to 6, and then gradually decreased. At low pH, there are many H^+ in the solution, and the higher concentration of H^+ competes with Cd^{2+} for binding to the negatively charged active sites on the adsorbent active sites [39]. The optimum of pH for adsorption Cd(II) on IIP is at pH 6. At pH above 7, adsorption decreases because there are a large amount of OH^- ligands in the solution [40]. At a pH of above 7, only $Cd(OH)^+$ and $Cd(OH)_2$ significantly contribute to the total cadmium in

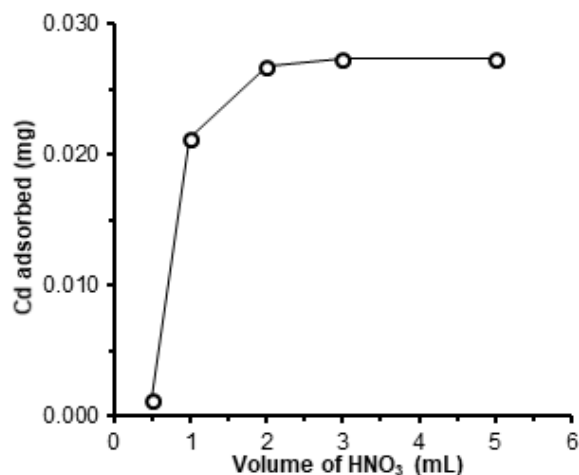
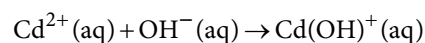


Fig 5. Influence of eluent volume on the eluted Cd(II) from IIP

a solution as shown in the following possible reactions [41]:



Influence of Eluent Volume

The influence of the eluent volume on adsorption of Cd(II) on the IIP was studied. The HNO_3 was used as an eluent. It was used to dissociate the bond between the analyte and the adsorbent. After sample loading, the adsorbed Cd(II) was eluted by passing 1 mol L^{-1} HNO_3 at different volume (from 0.5 to 5 mL), and the result was presented in Fig. 5. The recovery of Cd(II) was increased greatly from 0.5 to 1 mL and then increased slightly from 1 to 2 mL. The volume above 2 mL had a constant

recovery of Cd(II), and volume 2 mL was then chosen as an optimum volume. This result was similar to the study by Ebrahimzadeh et al. [42] which obtained that increase in volume and concentration of eluent caused higher recovery.

Influence of Flow Rate

The flow rate of the sample is related to the time of interaction between the analyte and the adsorbent. The higher flow rate will be more advantageous regarding obtaining a high enrichment factor because the higher sample volume is loaded into the column. But this is limited by the adsorption kinetics [23]. The sample flow rate through the sorbent should be low enough to enable efficient retention of the analytes, and high enough to avoid excessive duration analysis [16]. Different experiments were performed to find the optimum flow rates for the Cd(II) standard solution loading. The solution of Cd(II) standard was passed at a different flow rate (from 0.18 to 2.17 mL min⁻¹). As shown in Fig. 6, concentration of Cd(II) decrease for flow rates larger than 0.5 mL min⁻¹. The flow rate of 0.5 mL min⁻¹ was chosen as the optimum flow rate to decrease the time needed for analysis.

Analytical Characterization

The limit of detection (LoD) and limit of quantification (LoQ) were calculated according to $LoD = (3SD)/m$ and $LoQ = (10SD)/m$, where SD is the standard deviation and m is the slope of the calibration graph. The

calibration graph of Cd(II) without preconcentration and with 50-fold theoretical preconcentration was presented

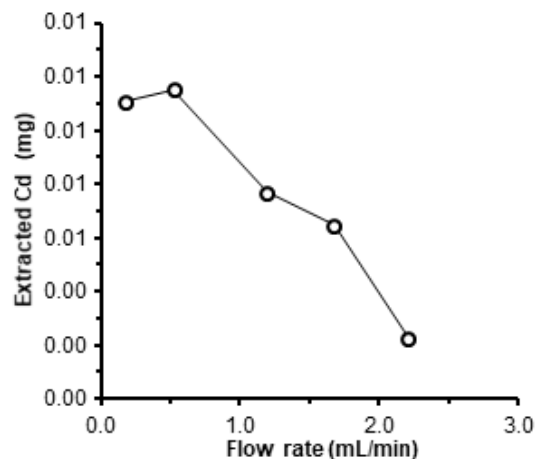


Fig 6. Influence of eluent flow rate on the extraction of Cd(II) on IIP

Table 1. Recovery of Cd(II) from solution samples after its SPE extraction on IIP (sample pH: 6; flow rate: 0.5 mL min⁻¹; elution: 2 mL of 1 mol L⁻¹ HNO₃)

No.	Added Cd (mg L ⁻¹)	Found Cd (mg L ⁻¹)	Recovery (%)
1	0.010	0.00966	96.60
2	0.020	0.01912	95.60
3	0.030	0.02882	96.07
4	0.040	0.04040	101.00
5	0.050	0.04964	99.28
Average			97.75
RSD (%)			2.32

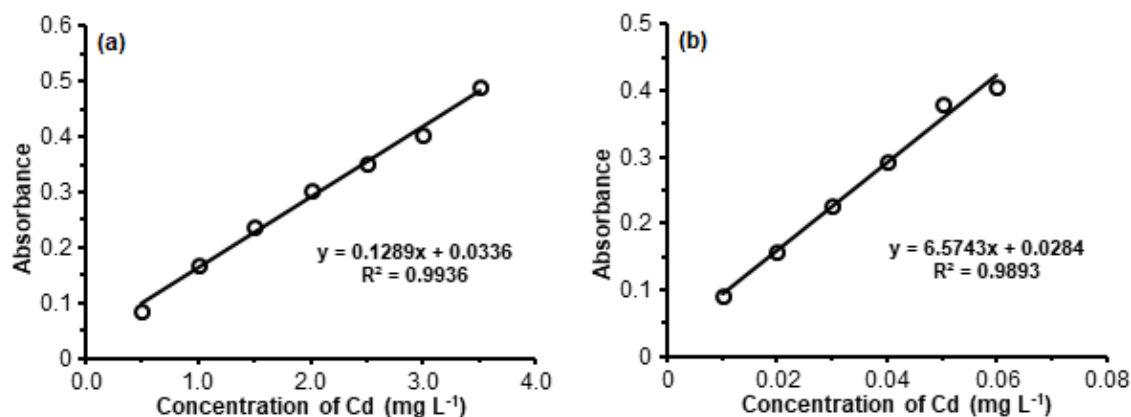


Fig 7. Calibration graph of Cd(II) solution (a) without preconcentration and (b) with 50-fold theoretical preconcentration

in Fig. 7. The LOD and LOQ obtained were 4.5 and 15 $\mu\text{g L}^{-1}$ for without preconcentration, 0.5 and 1.67 $\mu\text{g L}^{-1}$ for with 50-fold theoretical preconcentration. Application of the proposed method on the analysis of Cd(II) in an aqueous solution is presented in Table 1. Analytical recovery was assessed by analyzing fifth replicates of cadmium standard solution 0.01–0.05 mg L^{-1} and preconcentration by SPE. The analytical recovery obtained was 97.75%.

■ CONCLUSION

The new adsorbent IIP for the analysis of Cd(II) by SPE-FAAS has been synthesized through a precipitation polymerization process using 8-hydroxyquinoline as a complexing ligand, methacrylic acid as a monomer and ethylene glycol dimethacrylate as a crosslinker. Ethanol-acetonitrile (2:1) was used as a porogen, and benzoyl peroxide was used as an initiator. The optimized procedure consists of a sample loading (sample pH of 6) through IIP-SPE columns containing 100 mg of the IIP at a flow rate of 0.5 mL min^{-1} . Elution of Cd(II) from IIP was performed by passing 2 mL of 1 mol L^{-1} HNO_3 . The LoD and LoQ obtained were 0.5 and 1.67 $\mu\text{g L}^{-1}$ for with 50-fold theoretical preconcentration. The accuracy of the method of SPE-FAAS was 97.75%.

■ REFERENCES

- [1] Veneu, D.M., Pino, G.A.H., Torem, M.L., and Saint'Pierre, T.D., 2012, Biosorptive removal of cadmium from aqueous solutions using a *Streptomyces lunalinharesii* strain, *Miner. Eng.*, 29, 112–120.
- [2] Wang, K., Zhao, J., Li, H., Zhang, X., and Shi, H., 2016, Removal of cadmium(II) from aqueous solution by granular activated carbon supported magnesium hydroxide, *J. Taiwan Inst. Chem. Eng.*, 61, 287–297.
- [3] Salah, T.A., Mohammad, A.M., Hassan, M.A., and El-Anadouli, B.E., 2014, Development of nano-hydroxyapatite/chitosan composite for cadmium ions removal in wastewater treatment, *J. Taiwan Inst. Chem. Eng.*, 45 (4), 1571–1577.
- [4] Mahmoud, M.E., and Haggag, S.M.S., 2011, Static removal of cadmium from aqueous and nonaqueous matrices by application of layer-by-layer chemical deposition technique, *Chem. Eng. J.*, 166 (3), 916–922.
- [5] Kula, I., Uğurlu, M., Karaoğlu, H., and Çelik, A., 2008, Adsorption of Cd(II) ions from aqueous solutions using activated carbon prepared from olive stone by ZnCl_2 activation, *Bioresour. Technol.*, 99 (3), 492–501.
- [6] Waisberg, M., Joseph, P., Hale, B., and Beyersmann, D., 2003, Molecular and cellular mechanisms of cadmium carcinogenesis, *Toxicology*, 192 (2-3), 95–117.
- [7] Christensen, T.H., and Lun, X.Z., 1989, A method for determination of cadmium species in solid waste leachates, *Water Res.*, 23 (1), 73–80.
- [8] Workman, S.M., and Lindsay, W.I., 1990, Estimating divalent cadmium activities measured in arid-zone soils using competitive chelation, *Soil Sci. Soc. Am. J.*, 54 (4), 987–993.
- [9] Pinto, L., and Lemos, S.G., 2013, Multivariate optimization of the voltammetric determination of Cd, Cu, Pb and Zn at bismuth film. Application to analysis of biodiesel, *Microchem. J.*, 110, 417–424.
- [10] Holm, P.E., Andersen, S., and Christensen, T.H., 1995, Speciation of dissolved cadmium: Interpretation of analysis, ion exchange, and computer (GEOCHEM) methods, *Water Res.*, 29 (3), 803–809.
- [11] Gupta, K.C., and D'Arc, M.J., 2000, Effect of concentration of ion exchanger, plasticizer and molecular weight of cyanocopolymers on selectivity and sensitivity of Cd(II) ion selective electrode, *Talanta*, 52 (6), 1087–1103.
- [12] Bayen, S., Worms, I., Parthasarathy, N., Wilkinson, K., and Buffle, J., 2015, Cadmium bioavailability and speciation using the permeation liquid membrane, *Anal. Chim. Acta*, 575 (2), 267–273.
- [13] Fan, H.T., Lu, Y., Liu, A.J., Jiang, B., Shen, H., Huang, C.C., and Li, W.X., 2015, A method for measurement of free cadmium species in waters

- using diffusive gradients in thin films technique with an ion-imprinted sorbent, *Anal. Chim. Acta*, 897, 24–33.
- [14] Nolan, A.L., Mclaughlin, M.J., and Mason, S.D., 2003, Chemical speciation of Zn, Cd, Cu, and Pb in pore waters of agricultural and contaminated soils using Donnan dialysis, *Environ. Sci. Technol.*, 37 (1), 90–98.
- [15] Temminghoff, E.J.M., Plette, A.C.C., Eck, R.V., and Van Riemsdijk, W.H., 2000, Determination of the chemical speciation of trace metals in aqueous systems by the Wageningen Donnan Membrane Technique, *Anal. Chim. Acta*, 417 (2), 149–157.
- [16] Camel, V., 2003, Solid phase extraction of trace elements, *Spectrochim. Acta, Part B*, 58, 1177–233.
- [17] Wells, M.J.M., Riemer, D.D., and Wells-Knecht, M.C., 1994, Development and optimization of a solid-phase extraction scheme for determination of the pesticide metribuzin, atrazine, metalochlor, and esfenvalerate in agricultural run off water, *J. Chromatogr. A*, 659 (2), 337–348.
- [18] Johnson, W.E., Fendinger, N.J., and Plimmer, J.R., 1991, Solid phase extraction of pesticide from water: possible interferences from dissolved organic material, *Anal. Chem.*, 63 (15), 1510–1513.
- [19] Rao, T.P., Daniel, S., and Gladis, J.M., 2004, Tailored materials for preconcentration or separation of metals by ion-imprinted polymers for solid-phase extraction (IIP-SPE), *TrAC, Trends Anal. Chem.*, 23 (1), 28–35.
- [20] Hennion, M.C., and Coquart, V., 1993, Comparison of reversed-phase extraction sorbents for on-line trace enrichment of polar organic compounds in environmental aqueous samples, *J. Chromatogr. A*, 642 (1-2), 211–224.
- [21] Font, G., Manes, J., Molto, J.C., and Pico, Y., 1993, Solid-phase extraction in multi-residue pesticide analysis of water, *J. Chromatogr. A*, 642 (1-2), 135–161.
- [22] Snyder, L.R., Kirkland, J.J., and Glajch, J.L., 1997, *Practical HPLC Method Development*, 2nd ed., John Wiley & Sons, Inc., New York.
- [23] Gawin, M., Konefał, J., Trzewik, B., Walas, S., Tobiasz, A., Mrowiec, H., and Witek, E., 2010, Preparation of a new Cd(II)-imprinted polymer and its application to determination of cadmium(II) via flow-injection-flame atomic absorption spectrometry, *Talanta*, 80 (3), 1305–1310.
- [24] Tsoi, Y.K., Ho, Y.M., and Leung, K.S., 2012, Selective recognition of arsenic by tailoring ion-imprinted polymer for ICP-MS quantification, *Talanta*, 89, 162–168.
- [25] Özkütük, E.B., and Karabörk, M., 2007, Fe³⁺-Imprinted polymeric systems, *Hacettepe J. Biol. Chem.*, 35 (3), 195–202.
- [26] Utku, S., Yılmaz, E., Türkmen, D., Uzun, L., Garipcan, B., Say, R., and Denizli, A., 2008, Ion-imprinted thermosensitive polymers for Fe³⁺ removal from human plasma, *Hacettepe J. Biol. Chem.*, 36 (4), 291–304.
- [27] Buhani, Narsito, Nuryono, and Kunarti, E.S., 2010, Production of metal ion imprinted polymer from mercapto-silica through sol-gel process as selective adsorbent of cadmium, *Desalination*, 251 (1-3), 83–89.
- [28] Singh, D.K., and Mishra, S., 2009, Synthesis, characterization and removal of Cd(II) using Cd(II)-ion imprinted polymer, *J. Hazard. Mater.*, 164 (2-3), 1547–1551.
- [29] Singh, D.K., and Mishra, S., 2010, Synthesis and characterization of Hg(II)-ion-imprinted polymer: Kinetic and isotherm studies, *Desalination*, 257 (1-3), 177–183.
- [30] Dakova, I., Karadjova, I., Ivanov, I., Georgieva, V., Evtimora, B., and Georgiev, G., 2007, Solid phase selective separation and preconcentration of Cu(II) by Cu(II)-imprinted polymethacrylic microbeads, *Anal. Chim. Acta*, 584 (1), 196–203.
- [31] Cai, X., Li, J., Zhang, Z., Yang, F., Dong, R., and Chen, L., 2014, Novel Pb²⁺ ion imprinted polymers based on ionic interaction via synergy of dual functional monomers for selective solid-phase extraction of Pb²⁺ in water samples, *Appl. Mater. Interfaces*, 6 (1), 305–313

- [32] Liu, Y., Liu, Z., Gao, J., Dai, J., Han, J., Wang, Y., Xie, J., and Yan, Y., 2011, Selective adsorption behavior of Pb(II) by mesoporous silica SBA-15-supported Pb(II)-imprinted polymer based on surface molecularly imprinting technique, *J. Hazard. Mater.*, 186 (1), 197–205.
- [33] Chunxiang, L., Gao, J., Pan, J., Zhang, Z., and Yan, Y., 2009, Synthesis, characterization, and adsorption performance of Pb(II)-imprinted polymer in nano-TiO₂ matrix, *J. Environ. Sci.*, 21 (12), 1722–1729.
- [34] Shamsipur, M., Rajabi, H.R., Pourmortazavi, S.M., and Roushani, M., 2014, Ion imprinted polymeric nanoparticles for selective separation and sensitive determination of zinc ions in different matrices, *Spectrochim. Acta, Part A*, 117, 24–33.
- [35] Kim, M., Jiang, Y., and Kim, D., 2013, Zn²⁺-imprinted porous polymer beads: Synthesis, structure, and selective adsorption behavior for template ion, *React. Funct. Polym.*, 73 (6), 821–827.
- [36] Barciela-Alonso, M.C., Plata-García, V., Rouco-López, A., Moreda-Piñeiro, A., and Bermejo-Barrera, P., 2014, Ionic imprinted polymer based solid phase extraction for cadmium and lead pre-concentration/determination in seafood, *Microchim. J.*, 114, 106–110.
- [37] Lee, S.H., Jung, C.H., Chung, H., Lee, M.Y., and Yang, T.W., 1998, Removal of heavy metals from aqueous solution by apple residues, *Process Biochem.*, 33 (2), 205–211.
- [38] Doğan, M., Özdemir, Y., and Alkan, M., 2007, Adsorption kinetics and mechanism of cationic methyl violet and methylene blue dyes onto sepiolite, *Dyes and Pigments*, 75, 701–713.
- [39] Al-Homaidan, A.A., Alabdullatif, J.A., Al-Hazzani, A.A., Al-Ghanayem, A.A., and Alabbad, A.F., 2015, Adsorptive removal of cadmium ions by *Spirulina platensis* dry biomass, *Saudi J. Biol. Sci.*, 22 (6), 795–800.
- [40] Shofiyani, A., Narsito, Santosa, S.J., Noegrohati, S., Zahara, T.A., and Sayekti, E., 2015, Cadmium adsorption on chitosan/chlorella biomass sorbent prepared by ionic imprinting technique, *Indones. J. Chem.*, 15 (2), 163–171.
- [41] Luo, C., Wei, R., Guo, D., Zhang, S., and Yan, S., 2013, Adsorption behavior of MnO₂ functionalized multi-walled carbon nanotubes for the removal of cadmium from aqueous solutions, *Chem. Eng. J.*, 225, 406–415.
- [42] Ebrahimzadeh, H., Behbahani, M., Yamini, Y., Adlnasab, L., and Asgharinezhad, A.K., 2013, Optimization Cu(II)-ion imprinted nanoparticles for trace monitoring of copper in water and fish samples using a Box-Behnken design, *React. Funct. Polym.*, 73 (1), 23–29.