

Jurnal Perlindungan Tanaman Indonesia, Vol. 28, No. 2, 2024: 134–140 DOI: 10.22146/jpti.95262 Available online at http://jurnal.ugm.ac.id/jpti ISSN 1410-1637 (print), ISSN 2548-4788 (online)

Short Communication

An Update on the Fall Armyworm: Severity of Maize Damage and Susceptibility to Emamectine Benzoate and Chlorantraniliprole

Y. Andi Trisyono^{1)*}, Hadvina Nur Hendrayanti¹⁾, Arzaq P. Yuantomoputro¹⁾, Ary V. Setyaningrum¹⁾, Sriyanto Harjanto¹⁾, & Valentina E. F. Aryuwandari¹⁾

¹⁾Department of Plant Protection, Faculty of Agriculture, Universitas Gadjah Mada Jln. Flora No. 1, Bulaksumur, Sleman, Yogyakarta 55281 Indonesia

*Corresponding author. E-mail: anditrisyono@ugm.ac.id

Received April 4, 2024; revised April 19, 2024; accepted July 12, 2024

ABSTRACT

The damage area due to fall armyworm (FAW, *Spodoptera frugiperda*) in Indonesia decreased during the last three years (2021–2023) after the first outbreak in 2019. This insect continues to pose some risk for maize production with most reports documented the damage during the vegetative maize stage showing leave defoliation. This article provides an update on the high field population and the new type of damages caused by FAW by boring the stalks and feeding the cobs observed in the District of Grobogan, Central Java. The populations of egg masses, adults, and larvae were 0.4, 0.17, and 0.37 per plant of 27-day-old, respectively. The reproductive maize was heavily damaged by FAW with leave damage score of 9, 0.33 holes per stalk, and 75% of cobs damaged. The FAW larvae produced from the collected egg masses were still susceptible to emamectin benzoate and chlorantraniliprole. These findings prove that FAW can cause serious damage leading to almost total yield loss. As maize becomes more important in Indonesia, proper management in compliance with the Integrated Pest Management (IPM) principles is mandatory to keep the population low and prevent large-scale outbreaks.

Keywords: fall armyworm; Integrated Pest Management; stalk and cob damage; susceptibility

INTRODUCTION

Since the outbreak of Fall Armyworm (FAW), Spodoptera frugiperda J. E. Smith (Lepidoptera: Noctuidae), in African countries in 2016 that caused massive damage and economic losses (Goergen et al., 2016; IITA, 2016), this insect has spread and created problems in many Asian countries and Australia (Rane et al., 2023). In Indonesia, a report on the outbreak with noticeable damage was first published in 2019 (Trisyono et al., 2019). To the best of our knowledge, there is no document recording and showing the exact arrival time of this insect in Indonesia, and FAW has occupied 28 out of 33 provinces by 2023 (Directorate of Food Crop Protection MOA; personal communication in December 2023). Several researchers have published their research on FAW in different journals and proceedings (Mukkun et al., 2021; Sartiami et al., 2020; Vebryanti et al., 2023; Widhayasa et al.,

2022); however, research to determine the damage and economic losses due to this insect is still lacking.

Our communications with the farmers, agricultural officers, and pesticide companies showed that the FAW problems weakened after 2020 and this observation was in agreement with the data on damage areas at the national level. The damaged area in 2020 was 114.6 thousand ha and it was the highest up to the present time. Since then, the damaged area declined to 28.7 thousand ha in 2023 (Directorate of Food Crop Protection MOA; personal communication in December 2023). There are several plausible explanations for this declining trend, such as farmers and other stakeholders may have successfully found methods to manage FAW, local natural enemies have successfully adapted to this new host/prey resulting in increase natural control, or weather and climate have not been conducive for this insect to grow, develop, and an outbreak.

These factors may contribute individually or in combination to cause the decline. Maize plant resistance to FAW has been developed but it has not been commercialized in Indonesia until the date this paper was written. This control measure has the potential to be effective and easily adopted by the farmers once it is commercially available.

FAW poses a continuous threat to maize farmers although the 2021-2023 situation was better than the first two years of the outbreak (2019-2020). In the origin countries, FAW has continuously become a main pest because of its biotic potential and its ability to adapt to the control technologies being applied in the form of resistance development to chemical insecticides (Carvalho et al., 2013; Muraro et al., 2022; Nascimento et al., 2016; Yu, 1991) and transgenic maize expressing Bacillus thuringiensis toxins (Boaventura et al., 2021; Gutierrez-Moreno et al., 2020; Yang et al., 2021). Monitoring becomes a priority to determine the resistance shift of the population, and early detection would provide early warning and time for mitigation. In the same sense, this article documents and reports the reality of heavy damage and the high population of FAW in maize (potential field damage) in a couple of locations in the District of Grobogan, Central Java. This finding provides a precautionary understanding for all stakeholders on the potential damage and vield reduction caused by FAW, and insightful efforts in managing this pest to prevent further losses.

MATERIALS AND METHODS

Selected Locations

Field observation was carried out in the District of Grobogan, one of the maize production centers in the Province of Central Java. The sites of field observation were selected purposively with the criteria of maize heavily damaged and infested by FAW. Before the site selection, information regarding heavily damaged maize and high infestation was received from farmers and agricultural officers followed by a field survey to confirm the damage in several sites in the District of Grobogan, Central Java. The two selected sites for field observations were Sugihmanik, Sub-district of Tanggungharjo; 7°05'20.2"S 110°36'46.7"E, and Nampu, Sub-district of Karangrayung; 7°14'22.9"S 110°47'07.5"E. Sugihmanik represented lowland area with vegetative maize (27 days after planting [DAP]) infested by high population of FAW. Nampu represented a high land area with a reproductive stage (> 60 DAP) and was heavily damaged by FAW.

Sampling Method for Damage Assessment

The field assessment was conducted on January 27, 2024, for both selected locations. The site for sampling in Sugihmanik was a plot of 0.24 ha maize of 27 DAP. This plot was surrounded by >100 ha of reproductive maize (Figure 1A). Four sampling units with each unit consisting of 10 plants were selected randomly. The 10 maize plants were distributed in two adjacent rows, each consisting of five plants consecutively. The distance between each sampling unit was >10 m apart. In each plant sample, observation was made to determine the defoliation using Davis's scale (Davis et al., 1992), population of larvae, adults, and egg masses. More than 100 egg masses were collected and brought to the laboratory for further assessment of the susceptibility of newly hatched larvae against two insecticides (emamectin benzoate and chlorantraniliprole). Based on the information from the owner, this plot has been sprayed twice with emamectin benzoate and chlorantraniliprole and applied once with carbofuran by pouring the granule formulation into the whorl.

Different from the Sugihmanik site, Nampu was the maize center production in the upland area surrounded by teak plantations. The sampling procedure for Nampu was similar to that of Sugihmanik. Two different plots were observed in Nampu for different purposes. These two plots were side by side separated by a village road. The first plot was used to observe the defoliation and stalk borer; while the second plot was used to observe the cob damage. Because of heavy defoliation and stalk damage (Figure 1B), this maize did not produce sufficient cobs for observations. Defoliation was score using Davis's scale (Davis et al., 1992), and stalk damage was assessed by counting the number of plants showing one or more holes caused by FAW. The holes made by larvae of FAW (Figure 2A) were different from those made by Asian corn borer (ACB), Ostrinia furnacalis (Figure 2B). Similar to stalk damage, the number of cobs damaged was counted

in 10 plants per unit with a total of four sampling units (Figure 2C). This symptom was unique and different from cob damage due to three other Lepidopteran species often attacking the cobs: ACB, corn earworm *Helicoverpa armigera* (CEW), tobacco cutworm *Spodoptera litura* (TC).

Susceptibility of FAW Larvae to Insecticides

Newly hatched larvae from field-collected egg masses were tested for their susceptibility against the two commonly used insecticides for controlling FAW, emamectin benzoate (Emacel® 30 EC, Indonesia) and chlorantraniliprole (Prevathon® 50 SC, Indonesia) under laboratory conditions. Bioassays were carried out by dipping artificial diet in insecticide solutions for 10 seconds. Three concentrations were examined for each insecticide: 3.75, 15, and 60 ppm for emamectin benzoate and 6.25, 25, and 100 ppm for chlorantraniliprole. These concentrations ranged from the possible lowest to the highest field reccomendation rates. After dipping, the cube diets $(1 \times 1 \times 1 \text{ cm})$ were air dried before placing them in the plastic cups (3.3 cm in diameter and 4.3 cm in height, one diet per cup). Ten newly hatched larvae were then transferred into a treated or control diet, and each treatment was replicated four times. Mortality in the control and treated diet was observed 4 days after treatment.



Figure 1. Vegetative maize (27-day-old plants) surrounded by reproductive maize and heavily populated by the Fall Armyworm (FAW), *Spodoptera frugiperda*, in Sugihmanik, Grobogan, Jawa Tengah (A) and reproductive maize heavily damaged by FAW in Nampu, Karangrayung, Grobogan (B)



Figure 2. A hole in the maize stalk bored by a larva of the Fall Armyworm (FAW), *Spodoptera frugiperda* (A), the Asian corn borer, *Ostrinia furnacalis* (B), and a cob damaged by FAW (C)

In addition, a mass susceptibility test was done by exposing >1000 newly hatched larvae from the field-collected egg masses into the diet ($8 \times 8 \times 1$ cm) treated with chlorantraniliprole (25 ppm) in plastic jars (15 cm in diameter and 5.5 cm in height). Larval mortality was observed on the fourth day after treatment.

RESULTS AND DISCUSSION

Maize Damage

The maize in Sugihmanik received light defoliation by FAW with the score ranging from 2–3 on Davis's scale. However, the crops were heavily infested by egg masses with different stages starting from newly laid (Figure 3A) to early hatching (Figure 3B). In addition, adults (Figure 3C) were also found. The average populations for egg masses, adults, and larvae were 0.4, 0.17, and 0.37 per maize crop, respectively.

Very low damage on leaves was the result of intensive insecticide applications (three times within the span of 27-day-old-maize). The observed larvae were the survivors of these treatments. Their survival may be due to insufficient exposure to insecticide applications or their ability to resist applied insecticides. Early reports showed that the seven population of FAW. in Central Java were still susceptible to emamectin benzoate, chlorantraniliprole, and spinetoram (Suryani *et al.*, 2022). However, intensive application of insecticides in Grobogan may have been selected for resistance development. Further experiments are needed to determine whether the resistant population of FAW exists.

The high population of adults and egg masses indicates that there had been adult migration coming from the surrounding areas which was dominated by reproductive maize. Our field observations in many provinces, including in Lampung in 2019 (Trisyono *et al.*, 2019), showed that females preferred to lay eggs on the vegetative stage of maize approximately 1–2 weeks after planting and the most damage also occurred during the vegetative stage. This finding provides some insights related to FAW management, particularly in designing the planting seasons and crop rotations.

In Nampu, the reproductive maize was heavily damaged by FAW with 100% of the plants defoliated with an average damage score of 9 (the maximum score) (Figure 1B) and an average of holes of



Figure 3. Newly laid egg mass (A), egg hatching (B), and adult (C) of the Fall Armyworm (FAW), *Spodoptera frugiperda* observed in maize on the same day (January 27, 2024) in Sugihmanik, Grobogan, Central Java

0.33 holes per stalk (Figure 2B). In this plot, plants mostly did not produce cobs because of highly damaged maize before the cob formation. In another plot, the defoliation and stalk damage were less and cobs were formed. Unfortunately, these cobs were also heavily damaged with an average of 75%. The damaged cobs were marked with one or more holes (Figure 4A), and these damaged cobs almost had no kernels inside (Figures 2C and 4B). The heavy damage in the reproductive stage was

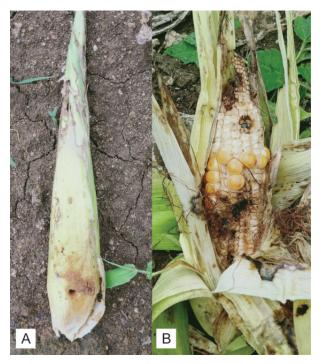


Figure 4. A cob with holes made the Fall Armyworm (FAW), *Spodoptera frugiperda* (A), and the more developed and heavily damaged cob FAW (B)

first publicly reported in Indonesia but these types of damage have already been reported in other countries (Rwomushana, 2019). This may suggest that heavy damage could occur anytime in any place if proper management is not in place.

Since the first outbreak in Sumatra in 2019, damaged area spread dan increase in 2020, the damage areas tended to decrease at the national level during the last two years (2021 and 2022) (Directorate of Food Crop Protection MOA, 2023; personal communication). This trend was similar in other regions, including in African and Southeast Asian countries. This finding provides an early warning of how bad FAW could cause damage and significantly reduce the yield. Continuous monitoring of this invasive species is a must and the development of Integrated Pest Management (IPM) for FAW could protect and prevent the damage and yield loss due to this insect reaching its maximum capacity.

Larval Susceptibility to Insecticides

All tested concentrations for both insecticides (emamectin benzoate and chlorantraniliprole) resulted in 100% mortality by the fourth day after treatment while no control mortality was observed. In addition, mass treatment of 25 ppm chlorantraniliprole yielded the same result. These suggest that the larvae hatching from the field-collected egg masses were still susceptible to these two insecticides. However, we also observed some larval survivors in the same field which had been sprayed with the same insecticides. At this point, we cannot

conclude if these larvae were resistant to the insecticides and further experiments are needed to determine their status. The results from these labora- tory bioassays should not be directly connected with the field situation because the females FAW that laid egg masses and used for these laboratory tests were different from those that laid egg masses earlier and produced the larvae even after insecticide application. In other words, the female populations may be two different genetic pools. Another possible explanation for the surviving larvae in the field was due to sublethal exposure because of misuse of insecticide application, such as applying at a lower rate, improper use of nozzle causing uneven coverage of leaves surface, missing the proper timing, or simply because the larvae escape temporarily from exposure.

This update should be treated as an awakening call that FAW continues to pose a threat to maize growers exacerbated with the ACB and other pest and diseases. Improper use of insecticides may contribute to unintended impacts, such as reducing the role of local natural enemies and the development of resistance that will lead to control failure. As maize becomes more important from year to year in Indonesia, proper management in compliance with the IPM principles including the use of maize plant resistance, is mandatory to keep the population low and prevent large-scale outbreaks.

ACKNOWLEDGEMENT

We would like to thank the farmers for allowing us to make observations on their maize crops and giving some information regarding their agronomic practices.

LITERATURE CITED

- Boaventura, D., Buer, B., Hamaekers, N., Maiwald, F., & Nauen, R. (2021). Toxicological and Molecular Profiling of Insecticide Resistance in a Brazilian Strain of Fall Armyworm Resistant to Bt Cry1 Proteins. *Pest Management Science*, 77(8), 3713–3726. https://doi.org/10.1002/ps.6061
- Carvalho, R.A., Omoto, C., Field, L.M., Williamson, M.S., & Bass, C. (2013). Investigating the Mo-

lecular Mechanisms of Organophosphate and Pyrethroid Resistance in the Fall Armyworm *Spodoptera frugiperda*. *PLoS ONE*, *8*(4), e62268. https://doi.org/10.1371/journal.pone.0062268

- Davis, F.M., Ng, S.S., & Williams, W.P. (1992). Visual Rating Scales for Screening Whorl-stage Corn for Resistance to Fall Armyworm (Technical Bulletin 186 MS39762; pp. 1–9). Mississippi State University.
- Goergen, G., Kumar, P.L., Sankung, S.B., Togola, A., & Tamò, M. (2016). First Report of Outbreaks of the Fall Armyworm *Spodoptera frugiperda* (J E Smith) (Lepidoptera, Noctuidae), a New Alien Invasive Pest in West and Central Africa. *PLoS ONE*, *11*(10), e0165632. https://doi.org/10. 1371/journal.pone.0165632
- Gutierrez-Moreno, R., Mota-Sanchez, D., Blanco, C.A., Chandrasena, D., Difonzo, C., Conner, J., Head, G., Berman, K., & Wise, J. (2020). Susceptibility of Fall Armyworms (*Spodoptera frugiperda* J.E.) from Mexico and Puerto Rico to Bt Proteins. *Insects*, 11(12), 831. https://doi.org/ 10.3390/insects11120831
- International Institute of Tropical Agriculture [IITA]. (2016, June 18). First Report of Outbreaks of the "Fall Armyworm" on the African Continent. IITA News. http://bulletin.iita.org/first-report-ofoutbreaks-of-the-fall-armyworm-on-the-africancontinent/
- Mukkun, L., Kleden, Y.L., & Simamora, A.V. (2021). Detection of *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) in Maize Field in East Flores District, East Nusa Tenggara Province, Indonesia. *International Journal of Tropical Drylands*, 5(1), 20–26. https://doi.org/10.13057/tropdrylands/t050104
- Muraro, D.S., Salmeron, E., Cruz, J.V.S., Amaral, F.S.A., Guidolin, A.S., Nascimento, A.R.B., Malaquias, J.B., Bernardi, O., & Omoto, C. (2022). Evidence of Field-Evolved Resistance in *Spodoptera frugiperda* (Lepidoptera: Noctuidae) to Emamectin Benzoate in Brazil. *Crop Protection*, 162, 106071. https://doi.org/10.1016/j.cropro. 2022.106071

- Nascimento, A.R.B.D., Farias, J.R., Bernardi, D., Horikoshi, R.J., & Omoto, C. (2016). Genetic Basis of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) Resistance to the Chitin Synthesis Inhibitor Lufenuron. *Pest Management Science*, 72(4), 810–815. https://doi.org/10.1002/ps.4057
- Rane, R., Walsh, T.K., Lenancker, P., Gock, A., Dao, T.H., Nguyen, V.L., Khin, T.N., Amalin, D., Chittarath, K., Faheem, M., Annamalai, S., Thanarajoo, S.S., Trisyono, Y.A., Khay, S., Kim, J., Kuniata, L., Powell, K., Kalyebi, A., Otim, M. H., ... Tay, W.T. (2023). Complex Multiple Introductions Drive Fall Armyworm Invasions into Asia and Australia. *Scientific Reports*, *13*(1), 660. https://doi.org/10.1038/s41598-023-27501-x
- Rwomushana, I. (2019). Spodoptera frugiperda (Fall Armyworm). CABI Compendium, 29810. https:// doi.org/10.1079/cabicompendium.29810
- Sartiami, D., Dadang, Harahap, I.S., Kusumah, Y. M., & Anwar, R. (2020). First Record of Fall Armyworm (*Spodoptera frugiperda*) in Indonesia and Its Occurence in Three Provinces. *IOP Conference Series: Earth and Environmental Science*, 468(1), 012021. https://doi.org/10.1088/1755-1315/468/1/012021
- Suryani, J.N., Trisyono, Y.A., & Martono, E. (2022). Susceptibility of *Spodoptera frugiperda* J. E. Smith (Lepidoptera: Noctuidae) Collected from Central Java Province to Emamectin Benzoate, Chlorantraniliprole, and Spinetoram. *Jurnal Perlindungan Tanaman Indonesia*, 26(2), 159–166. https://doi. org/10.22146/jpti.76195
- Trisyono, Y.A., Suputa., Aryuwandari, V.E.F., Hartaman, M., & Jumari. (2019). Occurrence of Heavy Infestation by the Fall Armyworm *Spodoptera frugiperda*, a New Alien Invasive Pest, in Corn in Lampung Indonesia. *Jurnal Perlindungan Tanaman Indonesia*, 23(1), 156–160. https://doi. org/10.22146/jpti.46455
- Vebryanti, A., Daud, I.D., & Dungga, N.E. (2023). Damage Caused by Spodoptera frugiperda J.E Smith on Corn in Climate Zones in South Sulawesi, Indonesia. IOP Conference Series: Earth and Environmental Science, 1255(1), 012011. https://doi.org/10.1088/1755-1315/1255/1/ 012011

- Widhayasa, B., Darma, E.S., Gendroyono, H., & Prasetyani, E.D. (2022). Detection of the Fall Armyworm Spodoptera frugiperda and Its Damage Symptoms to Maize in East Kalimantan, Indonesia. IOP Conference Series: Earth and Environmental Science, 1083(1), 012094. https://doi. org/10.1088/1755-1315/1083/1/012094
- Yang, F., Williams, J., Huang, F., & Kerns, D.L. (2021). Genetic Basis and Cross-Resistance of Vip3Aa Resistance in *Spodoptera frugiperda* (Lepidoptera: Noctuidae) Derived from Texas, USA. *Crop Protection*, 147, 105702. https://doi.org/ 10.1016/j.cropro.2021.105702
- Yu, S.J. (1991). Insecticide Resistance in the Fall Armyworm, Spodoptera frugiperda (J. E. Smith). Pesticide Biochemistry and Physiology, 39(1), 84–91. https://doi.org/10.1016/0048-3575(91)90216-9