

INTEGRATED MICROCONTROLLER MQ SENSORS FOR MONITORING BIOGAS: ADVANCEMENTS IN METHANE AND HYDROGEN SULFIDE DETECTION

SENSOR MQ MIKROKONTROLER TERINTEGRASI UNTUK MEMANTAU BIOGAS: KEMAJUAN DALAM DETEKSI METANA DAN HIDROGEN SULFIDA

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ABSTRAK

Terobosan teknologi mikrokontroler mampu memberikan keuntungan dalam sistem monitoring biogas. Penelitian ini bertujuan untuk memonitoring konsentrasi gas metana dan hidrogen sulfida pada biogas berbasis mikrokontroler. Bahan baku biogas yang digunakan di penelitian ini adalah 100% kotoran sapi dan 50:50 antara campuran kotoran sapi dengan limbah padat kota atau municipal solid waste (MSW). Instrumen penelitian terdiri dari: sensor MQ-4 diaplikasikan untuk mendeteksi konsentrasi metana dan sensor MQ-136 untuk mendeteksi hidrogen sulfida serta dilengkapi dengan termokopel tipe k untuk memonitoring temperatur anaerobik digester dan temperatur lingkungan. Semua sensor terintegrasi dengan mikrokontroler tipe ATmega 2560 yang diaplikasikan dalam penelitian ini. Teramati konsentrasi metana tertinggi pada biogas 100% kotoran sapi sebesar 3488 ppm pada waktu ke 21 hari. Hasil ini membuktikan biogas dari 100% kotoran sapi menghasilkan metana lebih baik dibandingkan biogas dari 50:50 campuran kotoran sapi dengan MSW. Konsentrasi hidrogen sulfida teramati mencapai 195 ppm pada 100% kotoran sapi dan 192 ppm untuk 50:50 campuran kotoran sapi dengan MSW. Temperatur teramati dalam kondisi mesofilik selama investigasi. Sistem monitoring biogas berbasis mikrokontroler merupakan teknologi yang menjanjikan karena mampu memberikan hasil secara real-time.

Kata kunci: Biogas; Microcontroller; Monitoring; Sensor MQ; Metana; Hidrogen sulfida.

ABSTRACT

Recent technological advances in microcontroller systems enable novel biogas monitoring capabilities. This study investigates microcontroller-based quantification of methane and hydrogen sulfide concentrations in biogas derived from anaerobic digestion. Anaerobic digesters were fed either 100% cow dung substrates or a 50:50 mixture of cow dung with municipal solid waste (MSW). Methane levels were monitored using an MQ-4 sensor, hydrogen sulfide via an MQ-136 sensor, and temperature with a K-type thermocouple, all integrated with an ATmega 2560 microcontroller system. The 100% cow dung digester produced biogas with maximum methane concentrations

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of 3488 ppm at 21 days, indicating improved methane production compared to the 50:50 mixture of cow dung with MSW. Hydrogen sulfide reached 195 ppm and 192 ppm for the 100% cow dung and mixed digesters. Mesophilic temperature conditions were maintained throughout the digestion process. Real-time quantification of biogas composition demonstrates the capabilities of microcontroller-based anaerobic digester monitoring to provide precise methane and hydrogen sulfide measurements.

Keywords: Biogas; Microcontroller; Monitoring; MQ sensors; Methane; Hydrogen sulfide.

INTRODUCTION

Fossil fuels, including oil, natural gas, and coal, currently provide most of the world's energy needs. However, increasing concerns about energy security and the environmental impacts of greenhouse gas emissions have prompted interest in renewable energy sources [1], [2]. Biogas, which is comprised primarily of methane (CH_4) 45-75% and carbon dioxide (CO_2) 25-45% [3], is one such renewable fuel that can be sustainably produced through the biological conversion of organic materials in the absence of oxygen, known as anaerobic digestion (AD) [4], [5].

Compared to other renewables like solar, wind, or hydropower, biogas offers unique advantages as an energy source that is continuously available, storable, and flexible for electricity, heating, or vehicle fuel [6], [7]. In addition, biogas production through anaerobic digestion provides an efficient waste management solution that helps mitigate potent greenhouse gas emissions from organic waste streams, including manure, crop residues, and food waste [8].

Cow dung is particularly suitable for biogas production because of its 55–65% methane content. The high cellulose and hemicellulose content in cow dung can be efficiently converted to methane by anaerobic digestion [9], [10]. The biogas production involves mixing cow dung with water and feeding it into a sealed underground anaerobic digester tank [11], [12].

Municipal solid waste (MSW) refers to household trash and rubbish collected by local authorities from residential and commer-

cial areas. MSW mainly contains biodegradable organic components such as food waste, garden waste, and paper products, which have great potential for conversion to biogas through anaerobic digestion [13].

With rapid urbanization worldwide, volumes of MSW are rising sharply, posing a challenge for environmentally sustainable disposal. Landfilling and incineration also have limitations. Anaerobic digestion provides an alternative waste treatment method that produces clean energy as an end-product and digestate that can be used as fertilizer [14].

The anaerobic digestion process involves four phases: hydrolysis, acidogenesis, acetogenesis, and methanogenesis. In hydrolysis, extracellular enzymes released by hydrolytic bacteria convert complex insoluble organic polymers such as carbohydrates, proteins, and lipids into soluble monomeric units like sugars, amino acids, and fatty acids [15], [16].

Subsequently, in the acidogenesis phase, acidogenic bacteria ferment these monomers into intermediate products, including volatile fatty acids (VFAs), alcohols, hydrogen, and carbon dioxide. These intermediates are then converted into acetic acid, carbon dioxide, and hydrogen by acetogenic bacteria during acetogenesis [17]. The last phase is methanogenesis; methanogens utilize acetic acid, carbon dioxide, and hydrogen to generate methane gas [18].

Measurement of methane is imperative given its dual significance as a potent greenhouse gas and combustible biofuel. The methane concentration determines the calorific value of gases generated from renewable feedstocks, including biogas and landfill gas [19]. Conventional methane gas measurement has many disadvantages, such as relatively expensive costs and low measurement efficiency.

Additionally, the measurement results cannot be delivered in real time. The measurement of hydrogen sulfide (H_2S) present in biogas is imperative, as hydrogen sulfide is considered an impurity. Hydrogen sulfide is highly corrosive and can induce rapid cor-

rosion in metallic materials. This occurs because hydrogen sulfide gas dissociates into hydrogen (H+) and bisulfide (HS-) ions when dissolved in water [20], [21]. Therefore, measuring both gases CH₄ and H₂S is essential to determine the characteristics of biogas.

Laboratory analysis of methane composition usually employs gas chromatography with thermal conductivity detection. Alternative methane measurement instrumentation such as infrared Draeger 6811960 and GEM2000/5000 (Geotech et al.) sensors have been applied for in situ biogas and sewer line gas monitoring [22]. However, limitations exist with these analytical techniques, including high capital costs and the requirement for substantial sample volumes. The MQ-4 is an affordable semiconductor-based methane detector that measures CH₄ concentrations ranging from 200 to 10,000 ppm.

The sensor shows optimal functionality within an ambient temperature range of 10 to 50°C and relative humidity below 95% [23], [24]. The MQ-4's sensitivity to methane combined with adjustments for temperature/humidity enables real-time monitoring of biogas methane content under typical anaerobic digester operating conditions. This sensor can be integrated with microcontroller devices such as Arduino, Raspberry, and other microcontrollers [6].

This research investigates the concentrations of methane (CH₄) and hydrogen sulfide (H₂S) gases produced in small-scale anaerobic digestion biogas systems utilizing 100% cow dung and a 50:50 mixture of cow dung with municipal solid waste (MSW) as substrates. The biogas systems were integrated with microcontroller technology utilizing an ATmega 2560 microcontroller for real-time monitoring and data acquisition.

In addition to gas concentrations, the ambient and digester temperatures were observed throughout the anaerobic digestion process. The acquired real-time data on temperature profiles and biogas composition from the integrated monitoring systems may facilitate the identification of optimal temperature ranges and organic loading rates

to maximize methane yields in these small-scale biogas digesters.

METHOD

Materials

The material tested in this paper is biogas generated from 100% cow dung and a 50:50 mixture of cow dung with MSW (Municipal et al.). The use of 100% cow dung is considered due to its abundant availability and specific characteristics. Cow dung contains approximately 18–20% volatile solids on a dry weight basis, including carbohydrates, proteins, fats, cellulose, hemicellulose, and lignin [25], [26].

The carbon-to-nitrogen ratio in fresh cow dung averages around 20:1, which is optimal for methanogenic bacteria to carry out anaerobic digestion. Cow dung also possesses a natural population of hydrolytic, fermentative, acetogenic, and methanogenic microbes required to catalyze the four stages of anaerobic digestion. Globally, biogas derived from cow dung is composed of around 55–70% methane, 30–50% carbon dioxide, and trace amounts of other gases [27]. In addition, biogas can also be produced through landfills, as shown in Table 1.

Table 1.
Anaerobic digestion and landfill biogas compositions

Component	AD biogas	Landfill biogas	Units
CH ₄	53–70	30–65	vol%
CO ₂	30–50	25–47	vol%
N ₂	2–6	<1–17	vol%
O ₂	0–5	<3–1	vol%
H ₂	NA	0–3	vol%
C _x H _y	NA	NA	vol%
H ₂ S	0–2000	30–500	ppm
NH ₃	<100	0–5	ppm
Chlorines	<0.25	0.3–225	mg Nm ³

Source: N. de Nooijer et.al (2018) [27]

The MSW used in this research included banana, tomato, and carrot peels. Banana

peels are a promising feedstock for biogas digesters due to their high carbohydrate and nutrient content. The main components are cellulose, hemicellulose, lignin, starch, and sugars. The biogas yield from banana peels with a cow manure content of 10% at 18 and 22 g of volatile solids (gvs) per liter was 50.20 and 40.49 gvs per day, respectively [28].

The actual yield can vary based on digester conditions and retention time. Tomato peels are rich in sugars and nutrients like nitrogen. The high moisture content and soft texture make them easily degradable [29]. Carrot peels also have high carbohydrate and nitrogen content. Their lignin content is lower than other vegetable wastes, making them more readily degraded by anaerobic bacteria [30].

Instrumentation Details for Experimental Setup

The anaerobic digestion process was conducted in a plastic drum digester under batch conditions, with 100% cow dung and

a 50:50 mixture of cow dung with MSW as the feedstock. The biogas plant operated at mesophilic temperature, and a highly active methanogenic community was present to facilitate the AD process. The experimental schematic is shown in Figure 1. Anaerobic digestion experiments were performed using an instrumented lab-scale digester continuously monitored by methane (CH_4), hydrogen sulfide (H_2S), and temperature sensors interfaced to an ATmega 2560 microcontroller system.

Methane and hydrogen sulfide gas were detected using MQ-4 and MQ-136 metal oxide semiconductor sensors in the digester headspace. The sensors operate on a resistance change principle when target gases are absorbed onto the heated sensor surface. Analog voltage signals proportional to gas concentrations are produced based on sensor resistances calculated through a Wheatstone bridge circuit with an output range of 0–5V corresponding to 0–10,000 ppm [31].

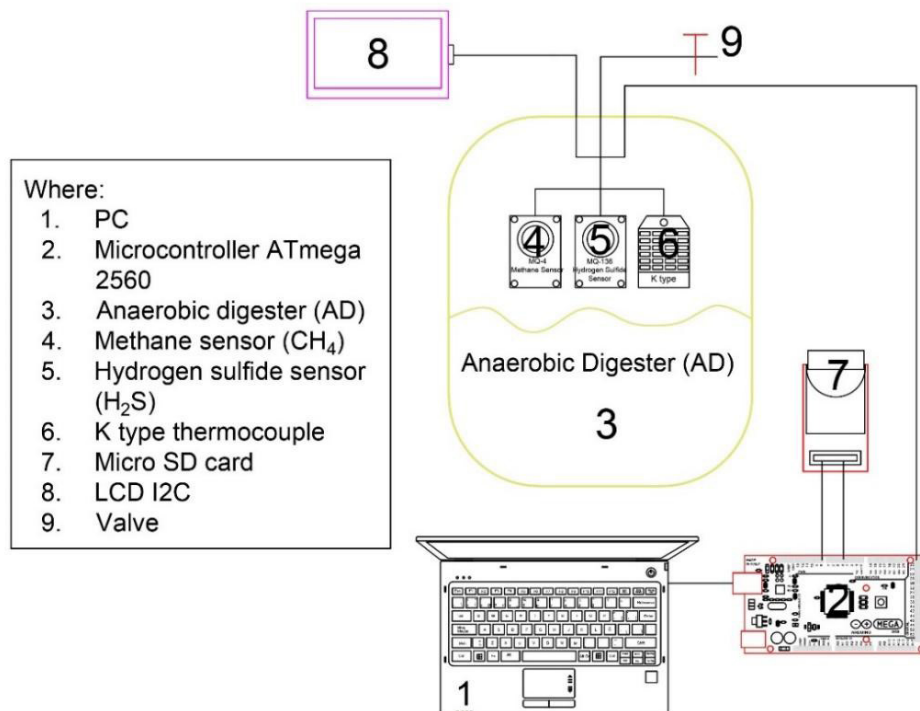


Figure 1.
 Experimental schematic of a microcontroller-integrated biogas system
 Source: Researchers' analysis (2023)

Temperature profiling utilized a type K thermocouple probe with chromel and alumel conductors to generate a temperature-dependent voltage via the thermoelectric effect. The millivolt-level output spans the -270°C to 1300°C measurement range [32]. The sensors were connected via jumper cables to analog inputs of the ATmega 2560, an 8-bit AVR RISC microcontroller clocked at 16MHz with 256KB flash and 8KB SRAM memory. ATmega 2560 specification can be seen in Table 2. Analog voltages from the sensors were digitized by the 10-bit analog-to-digital converter at a sampling rate of 1 kHz for high-resolution real-time data. Serial I2C communication enabled a liquid crystal display interfacing to visualize the measured parameters. An Arduino IDE programming environment facilitated custom firmware development for sensor data acquisition, processing, logging, and control. Real-time sensor measurements were transmitted over USB to a PC for 21 days. All measurement results are stored on the microSD and integrated with the microcontroller.

Table 2.
ATmega 2560 microcontroller technical specifications

Type	Description
Microcontroller	ATmega 2560
Operating voltage	5V
Input voltage	6-20V
Digital I/O pins	54 (of which provide PWM output)
Analog input pins	16
DC current per I/O pins	20 mA
Flash memory	256 kB of which 8 kB used by bootloader
SRAM	8 kB
EEPROM	4 kB
Clock speed	16 MHz

Source: A. S. Ismailov (2022) and Researchers' analysis

Setup of Microcontroller Configuration with MQ Gas Sensors

The analog input pin A0 on the ATmega 2560 microcontroller is connected to the analog output pin A0 on the MQ-4 gas sensor.

The analog input pin A1 on the ATmega 2560 is connected to the analog output pin A0 on the MQ-136 gas sensor. The VCC (power) pins on both gas sensors are connected to the 5V power rail on the ATmega 2560, while the GND (ground) pins on the sensors are wired to the ground rail on the ATmega 2560.

The positive and negative thermocouple wires from the K-type thermocouple are connected to the input terminals on the MAX6675 thermocouple amplifier, whose SCK (serial clock) pin is connected to digital I/O pin 12, CS (chip select) pin to digital I/O pin 11, and SO (serial data out) pin to digital I/O pin ten on the ATmega 2560.

The I2C interface pins SCL (serial clock) and SDA (serial data) on the 16x2 LCD are connected to the SCL and SDA pins on the ATmega 2560 at digital I/O pins 21 and 20, respectively. Power and ground for the LCD come from the 5V and GND rails on the ATmega 2560. The SPI interface pins on the microSD card - MISO (master in, an enslaved person out), MOSI (master out, an enslaved person in), SCK (serial clock), and CS (chip select) - are connected to digital I/O pins 50, 51, 52 and 53 respectively on the ATmega 2560.

Finally, the ATmega 2560 interfaces with the PC via a USB connection, as shown in Figure 2. The actual implementation of the microcontroller system with integrated sensors can be observed in Figure 3. The programming code is shown in Figure 4.

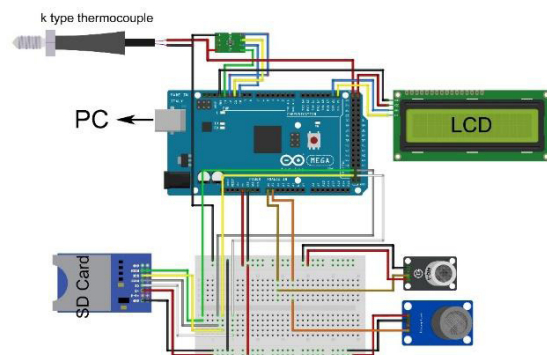


Figure 2.
Wiring and Interfacing Sensors to an ATmega 2560 Microcontroller
Source: Researchers' analysis (2023)

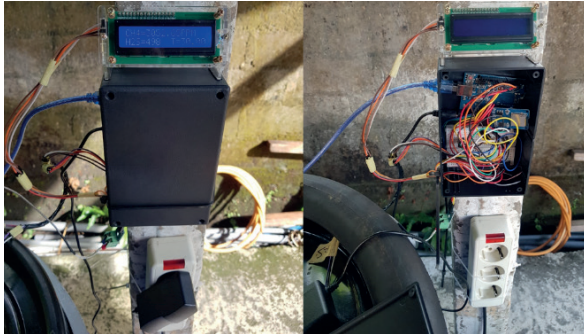


Figure 3.
 Actual implementation of the microcontroller system with integrated sensors
 Source: Researchers' analysis (2023)

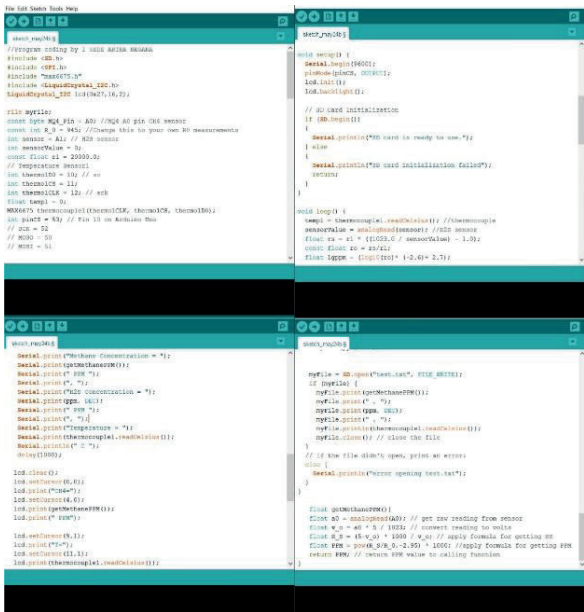


Figure 4.
 Programmed code utilizing the Arduino IDE software
 Source: Researchers' analysis (2023)

RESULTS AND DISCUSSION

Response of MQ-4 of Samples of Biogas

Figure 5 shows the response of the MQ-4 gas sensor to biogas over three trial repetitions, each spanning 2 minutes, with measurements recorded every second. The MQ-4 sensor being utilized in this experiment is designed to detect and measure methane concentration levels in biogas mixtures. The

biogas sample analyzed contains an approximate methane concentration of 3000 ppm.

As evident in Figure 5, the sensor measurements demonstrate the presence of small-scale random fluctuations and variability throughout the sampling duration.

Based on the three trials, this measured response exhibits a mean methane concentration of 3578 ppm. To quantify the variability, the standard deviation was 55.1 ppm, corresponding to 1.540% of the mean value. This relatively low standard deviation expresses little dispersion around the mean. In other words, the replicated trials aligned well, without substantial deviations between them under consistent test conditions.

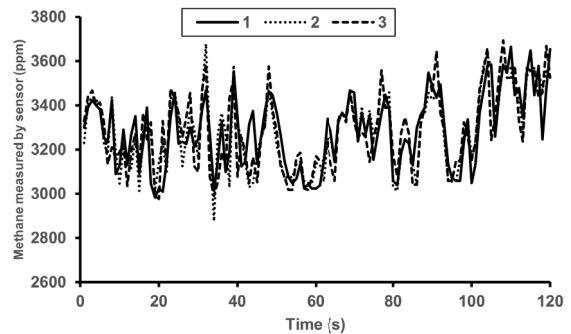


Figure 5.
 Response of the MQ-4 sensor to biogas
 Source: Researchers' analysis (2023)

Response of MQ-136 of Samples of Biogas

Figure 6 shows the response of the MQ-136 sensor to biogas over three repeated trials. The MQ-136 sensor was used to monitor hydrogen sulfide concentration levels every second for two minutes during each trial repetition. The hydrogen sulfide concentration exhibited fluctuations throughout the three trials, ranging from a minimum of 171.04 ppm to a maximum of 191.36 ppm. The observed fluctuations in biogas levels during the repeated trials highlight the importance of conducting multiple measurements to adequately characterize sensor response and account for inherent variability. The three repeats' mean, and standard deviation (expressed as % of mean) were $189.297 \pm 0.057\%$.

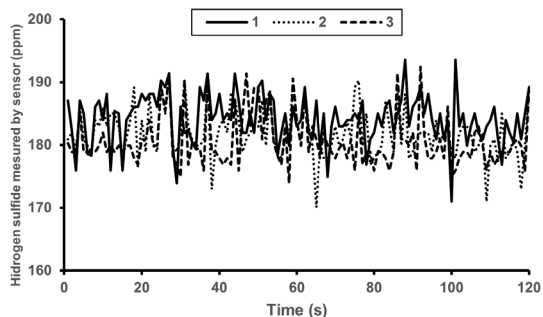


Figure 6.
Response of the MQ-136 sensor to biogas
Source: Researchers' analysis (2023)

Monitoring Methane and Hydrogen Sulfide Concentration in Anaerobic Digester (AD)

Figure 7 shows the variation in methane concentration between 100% cow dung and a 50:50 mixture of cow dung with MSW. There was no significant difference. Both samples were similar after six days of investigation. Methane concentration increased sharply in 100% cow dung, reaching 1224 ppm at approximately 12 days of incubation under mesophilic conditions at 35°C. This correlates to the exponential CH₄ production phase as methanogenic archaea generate biogas from intermediates like volatile fatty acids formed during initial hydrolysis and acetogenesis steps [23].

In contrast, the 50:50 mixture of cow dung with MSW co-digestion feedstock showed lower CH₄, reaching just 562 ppm by day 12. The delayed and reduced CH₄ production is likely due to the increased proportion of complex particulate organics in MSW, requiring longer hydrolysis than readily biodegradable cow dung [3]. Moreover, at 18 days, the methane concentration in 100% cow dung increased to 3046 ppm, while it reached 1284 ppm for the 50:50 mixture of cow dung with MSW.

The significantly higher CH₄ levels in mono-digestion of cow dung can be attributed to the fiber-rich composition, which provides ideal substrates for acetoclastic methanogenesis [33]. Cow dung contains

a significant amount of methane produced during the normal digestive process.

Figure 7 also shows that the methane concentration in both samples increases as the investigation time increases. In addition, the highest methane concentration was observed in 100% cow dung at 3488 ppm, while it was 1624 ppm for the 50:50 mixture of cow dung with MSW. As expected, cow dung's high cellulosic and hemicellulosic content promotes maximal methane generation by the endogenous gut archaea. In contrast, slowly biodegradable and inert fractions in MSW diluted the CH₄-producing potential in the co-digestion.

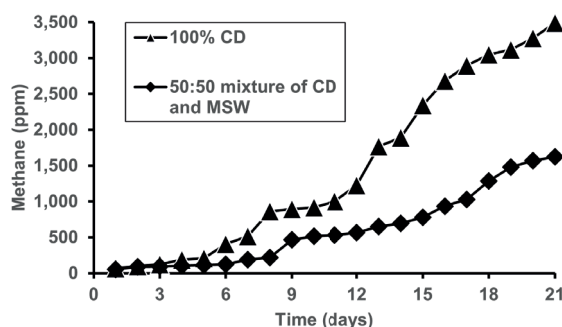


Figure 7.
Varied methane concentrations between 100% cow dung and a 50:50 mixture of cow dung with MSW
Source: Researchers' analysis (2023)

Variations in hydrogen sulfide concentration between 100% cow dung and a 50:50 mixture of cow dung with MSW are shown in Figure 8. From the figure, the hydrogen sulfide concentration increased significantly in both samples. At five days, the hydrogen sulfide concentration was 117 ppm in 100% cow dung, while 102 ppm for the 50:50 mixture of cow dung with MSW.

Cow dung contains sulfur-bearing organic compounds which serve as precursors for H₂S production. These include proteins like keratin and enzymes, amino acids such as methionine and cysteine, and other sulfur organics excreted in the manure [34]. The sulfur compounds get converted to H₂S gas during the anaerobic digestion process. Moreover, at around 15 days, both samples

showed an increase of 171 ppm in 100% cow dung and 157 ppm in the 50:50 mixture of cow dung with MSW.

The hydrogen sulfide concentration in the 50:50 mixture of cow dung with MSW tends to be lower than 100% cow dung. This is due to MSW containing lower sulfur content than cow dung [35]. MSW provides more balanced nutrition for methane-forming archaea, reducing H_2S formation. The maximum hydrogen sulfide observed in 100% cow dung was 195 ppm at 21 days of investigation. Inside a digester, anaerobic bacteria convert and ferment the organic matter in cow dung into biogas.

Sulfur compounds are metabolized into hydrogen sulfide and other sulfur byproducts like carbonyl sulfide. Factors including pH, temperature, and organic loading rate impact H_2S production. More free hydrogen ions can react with sulfur species at a neutral pH level to form H_2S . Higher temperatures speed up reaction kinetics. Overloading digesters can inhibit methane-forming archaea, leading to increased H_2S formation. Longer retention times also allow more sulfate reduction to H_2S [36].

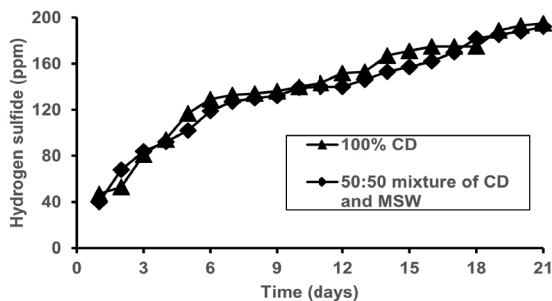


Figure 8.

Varied hydrogen sulfide concentrations between 100% cow dung and a 50:50 mixture of cow dung with MSW
 Source: Researchers' analysis (2023)

Monitoring Environmental and Anaerobic Digester (AD) Temperatures

Figure 9 shows environmental variations and anaerobic digester (AD) temperatures. From the observation, the digester temperature is relatively higher than the sur-

rounding environment temperature throughout the study period. This phenomenon can be attributed to the closed, insulated nature of anaerobic digesters, which retain the heat produced during the bacterial breakdown of organic matter. In contrast, the temperature of the surrounding environment is lower as external weather conditions influence it. At three days, the digester temperature was observed to be 43°C, while the surrounding environment was 35°C.

On the other hand, at nine days, the temperature of both samples decreased by 35°C in the digester and 28°C in the surrounding environment temperature. This temperature decline may have been caused by lower microbial activity or feedstock input rates during this period. The highest temperature observed was 44°C in the digester around 3 days of investigation.

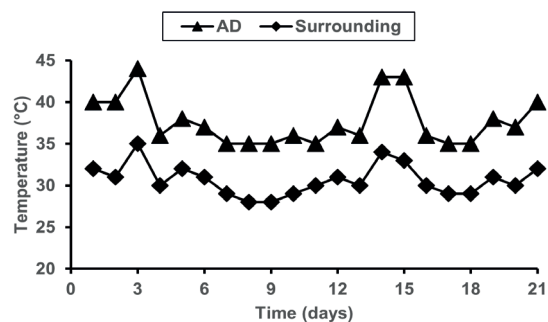


Figure 9.

Temperature variance between the anaerobic digester (AD) and the surrounding environment
 Source: Researchers' analysis (2023)

Microcontroller Role for Anaerobic Digestion Monitoring System

Due to its technical specifications and connectivity, the ATmega 2560 microcontroller has significant potential for real-time monitoring and control applications in biogas systems. The 8-bit AVR RISC-based processor operating at 16MHz provides sufficient computational performance for anaerobic digestion process control algorithms. Substantial compiled code and sensor data can be stored with 256KB of program memory and 8KB RAM. Sixteen 10-bit ADC channels allow interfacing with analog biogas sensors

to measure methane, hydrogen sulfide, and temperature quantitatively.

Digital I/O enables control of valves, pumps, and heating elements for automation. UART, I2C, and SPI buses support adding wireless modules for remote monitoring. The omega 2560's proven reliability in industrial environments, cost-effectiveness, and availability of programming libraries like Arduino make it highly adaptable for continuous sensing, logging, and real-time control in biogas plants. With proper integration of modern sensors and prudent firmware design, the ATmega 2560 has considerable scientific merit for increasing biogas yields through fine-grained monitoring of anaerobic digestion and automated system optimization.

CONCLUSION

This study demonstrates the effectiveness of microcontroller-implemented sensors for monitoring the quantification of critical process parameters in small-scale anaerobic digesters. Experimental data were collected on methane (CH₄), hydrogen sulfide (H₂S), and temperature levels during a 21-day mesophilic digestion investigation utilizing substrate formulations of 100% cow dung and a 50:50 mixture of cow dung with municipal solid waste (MSW).

Results indicate higher CH₄ production for the mono-digestion of cow dung compared to co-digestion, with maximum concentrations reaching 3488 ppm at day 21. Negligible differences in H₂S evolution were observed between the two feedstock conditions, approaching 195 ppm and 192 ppm for the cow dung and co-digestate, respectively. Operating temperatures were maintained below the mesophilic threshold throughout the investigation. The microcontroller-enabled monitoring system provided continuous, high-accuracy measurements of biogas composition, facilitating data analysis. Overall, this research validates the promise of automated on-site sensors for evaluating and optimizing small-scale biogas digesters.

BIBLIOGRAPHY

- [1] Zeng, X., Chen, G., Luo, S., Teng, Y., Zhang, Z. and Zhu, T. 2022. Renewable transition in the power and transport sectors under the goal of carbon-neutral in Sichuan, China. *Energy Reports*. Vol. 8, pp. 738-748, doi: 10.1016/j.egyr.2022.02.213.
- [2] Utami, I., Riski, M. A. and Hartanto, D. R. 2022. Nuclear Power Plants Technology to Realize Net Zero Emission 2060. *International Journal of Business and Management Technology*. Vol. 6, no. 1, pp. 158-162, [Online]. Available: www.theijbmt.com
- [3] Negara, I. G. A., Mulawarman, A. A. N. B., Santosa, I. G. and Midiani, L. P. I. 2023. Studi Eksperimental Generator Elektrik Berbahan Bakar Biogas Guna Mendukung Net Zero Emission. *Jurnal Rekayasa Mesin*. Vol. 14, no. 2, pp. 689-700, 2023, doi: 10.21776/jrm.v14i2.1431.
- [4] Kabeyi, M. J. B. and Olanrewaju, O. A. 2022. Technologies for biogas to electricity conversion. *Energy Reports*. Vol. 8, pp. 774-786, doi: 10.1016/j.egyr.2022.11.007.
- [5] Nindhia, T. G. T., McDonald, M. and Styles, D. 2021. Greenhouse gas mitigation and rural electricity generation by a novel two-stroke biogas engine. *Journal of Cleaner Production*. Vol. 280, p. 124473, doi: 10.1016/j.jclepro.2020.124473.
- [6] Zabed, H. M., Akter, S., Yun, J., Zhang, G., Zhang, Y. and Qi, X. 2020. Biogas from microalgae: Technologies, challenges and opportunities. *Renewable and Sustainable Energy Reviews*. Vol. 117, no. January, 2020, doi: 10.1016/j.rser.2019.109503.
- [7] Feiz, R., Johansson, M., Lindkvist, E., Moestedt, J., Pålédal, S. N. and Ometto, F. 2022. The biogas yield, climate impact, energy balance, nutrient recovery, and resource cost

- of biogas production from household food waste—A comparison of multiple cases from Sweden. *Journal of Cleaner Production*. Vol. 378, no. September, 2022, doi: 10.1016/j.jclepro.2022.134536.
- [8] Lindfors, A., Hagman, L. and Eklund, M. 2022. The Nordic biogas model: Conceptualization, societal effects, and policy recommendations. *City and Environment Interactions*. Vol. 15, no. June, p. 100083, 2022, doi: 10.1016/j.cacint.2022.100083.
- [9] Haryanto, A., Triyono, S. and Wicaksono, N. H. 2018. Effect of hydraulic retention time on biogas production from cow dung in a semi continuous anaerobic digester. *International Journal of Renewable Energy Development*. Vol. 7, no. 2, pp. 93–100, 2018, doi: 10.14710/ijred.7.2.93-100.
- [10] Obileke, K., Mamphweli, S., Meyer, E. L., Makaka, G., and Nwokolo, N. 2020. Design and Fabrication of a Plastic Biogas Digester for the Production of Biogas from Cow Dung. *Journal of Engineering*. Vol. 2020, pp. 1–11, 2020, doi: 10.1155/2020/1848714.
- [11] Singh, S., Hariteja, N., Sharma, S., Raju, N. J. and Prasad, T. J. R. 2021. Production of biogas from human faeces mixed with the co-substrate poultry litter & cow dung. *Environmental Technology & Innovation*. Vol. 23, no. April, 2021, doi: 10.1016/j.eti.2021.101551.
- [12] Negara, I. G. A., Anakottapary, D. S., Midiani, L. P. I., Temaja, I. W. and Santosa, I. D. M. C.. 2023. Experimental Study of Cooling Performance and Electrical Parameters in a Microcontroller-Driven Inverter AC System. Vol. 23, no. 2, pp. 81–90.
- [13] Llano, T., Arce, C. and Finger, D. C. 2021. Optimization of biogas production through anaerobic digestion of municipal solid waste: a case study in the capital area of Reykjavik, Iceland. *Journal of Chemical Technology and Biotechnology*. Vol. 96, no. 5, pp. 1333–1344, 2021, doi: 10.1002/jctb.6654.
- [14] Norouzi, O. and Dutta, A. 2022. The Current Status and Future Potential of Biogas Production from Canada’s Organic Fraction Municipal Solid Waste. *Energies*. Vol. 15, no. 2, doi: 10.3390/en15020475.
- [15] Anukam, A., Mohammadi, A., Naqvi, M. and Granström, K. 2019. A review of the chemistry of anaerobic digestion: Methods of accelerating and optimizing process efficiency. *Processes*. Vol. 7, no. 8, pp. 1–19, doi: 10.3390/PR7080504.
- [16] Meegoda, J. N., Patel, B. Li, K. and Wang, L. B. 2018. A review of the processes, parameters, and optimization of anaerobic digestion. *International Journal of Environmental Research and Public Health*. Vol. 15, no. 10, doi: 10.3390/ijerph15102224.
- [17] Náthia-Neves, G., Berni, M., Dragone, G., Mussatto, S. I. and Forster-Carneiro, T. 2018. Anaerobic digestion process: technological aspects and recent developments. *International Journal of Environmental Science and Technology*. Vol. 15, no. 9, pp. 2033–2046, doi: 10.1007/s13762-018-1682-2.
- [18] Morales-Polo, C., del Mar Cledera-Castro, M. and Yolanda Moratilla Soria, B. 2018. Reviewing the anaerobic digestion of food waste: From waste generation and anaerobic process to its perspectives. *Applied Sciences*. Vol. 8, no. 10, doi: 10.3390/app8101804.

- [19] Bharathiraja, B., Sudharsana, T., Jayamuthunagai, J., Praveenkumar, R., Chozhavendhan, S. and Iyyappan, J. 2018. Biogas production – A review on composition, fuel properties, feed stock and principles of anaerobic digestion. *Renewable and Sustainable Energy Reviews*. Vol. 90, no. April, pp. 570–582, 2018, doi: 10.1016/j.rser.2018.03.093.
- [20] Zwain, H. M., Nile, B. K., Faris, A. M., Vakili, M. and Dahlan, I. 2020. Modelling of hydrogen sulfide fate and emissions in extended aeration sewage treatment plant using TOXCHEM simulations. *Scientific Reports*. Vol. 10, no. 1, pp. 1–11, doi: 10.1038/s41598-020-79395-8.
- [21] Tirta Nindhia, T. G., Surata, I. W. and Wardana, A. 2017. The effect of biogas desulfurization to acidity of lubricant oil of the biogas fuelled engine. *IOP Conference Series: Materials Science and Engineering*. Vol. 201, no. 1, 2017, doi: 10.1088/1757-899X/201/1/012021.
- [22] Yang, S., Liu, Y., Wu, N., Zhang, Y., Svoronos, S. and Pullammanappallil, P. 2019. Low-cost, Arduino-based, portable device for measurement of methane composition in biogas. *Renewable Energy*. Vol. 138, pp. 224–229, doi: 10.1016/j.renene.2019.01.083.
- [23] Nagahage, I. S. P., Nagahage, E. A. A. D. and Fujino, T. 2021. Assessment of the applicability of a low-cost sensor-based methane monitoring system for continuous multi-channel sampling. *Environmental Monitoring and Assessment*. Vol. 193, no. 8, doi: 10.1007/s10661-021-09290-w.
- [24] Iswanto, Ma'arif, A., Kebenaran, B. and Megantoro, P. 2021. Design of gas concentration measurement and monitoring system for biogas power plant. *Indonesian Journal of Electrical Engineering and Computer Science*. Vol. 22, no. 2, pp. 726–732, doi: 10.11591/ijeecs.v22.i2.pp726-732.
- [25] Fasake, V. and Dashora, K. 2020. Characterization and morphology of natural dung polymer for potential industrial application as bio-based fillers. *Polymers (Basel)*. Vol. 12, no. 12, pp. 1–16, 2020, doi: 10.3390/polym12123030.
- [26] Font-Palma, C. 2019. Methods for the Treatment of Cattle Manure—A Review. *Journal of Carbon Research*. Vol. 5, no. 2, p. 27, doi: 10.3390/c5020027.
- [27] de Nooijer, N., Gallucci, F., Pellizzari, E., Melendez, J., Tanaka, D. A. P., Manzolini, G. and van Sint Annaland, M. 2018. On concentration polarisation in a fluidized bed membrane reactor for biogas steam reforming: Modelling and experimental validation. *Chemical Engineering Journal*. Vol. 348, no. March, pp. 232–243, doi: 10.1016/j.cej.2018.04.205.
- [28] Achinas, S., Krooneman, J. and Euverink, G. J. W. 2019. Enhanced Biogas Production from the Anaerobic Batch Treatment of Banana Peels. *Engineering*. Vol. 5, no. 5, pp. 970–978, doi: 10.1016/j.eng.2018.11.036.
- [29] Pataro, G., Carullo, D., Bakar Siddique, M. A., Falcone, M., Donsi, F. and Ferrari, G. 2018. Improved extractability of carotenoids from tomato peels as side benefits of PEF treatment of tomato fruit for more energy-efficient steam-assisted peeling. *Journal of Food Engineering*. Vol. 233, pp. 65–73, doi: 10.1016/j.jfoodeng.2018.03.029.
- [30] de Andrade Lima, M., Charalampopoulos, D. and Chatzifragkou, A. 2018. Optimisation and modelling of supercritical CO₂ extraction process

- of carotenoids from carrot peels. *The Journal of Supercritical Fluids*. Vol. 133, pp. 94–102, doi: 10.1016/j.supflu.2017.09.028.
- [31] Sakayo, N. M., Mutuku, J. N. and Ngaruiya, J. M. 2019. Design and Calibration of a Microcontroller Based MQ-4 Gas Sensor for Domestic Cooking Gas System. *International Journal of Applied Physics*. Vol. 6, no. 2, pp. 31–40, doi: 10.14445/23500301/ijap-v6i2p106.
- [32] Kadir, A. and Kako, S. 2022. Comparative Investigation on the Quality of Sensitivity of Six Different Types of Thermocouples. *Al-Rafidain Engineering Journal*. Vol. 27, no. 2, pp. 117–126, doi: 10.33899/rengj.2022.132862.1157.
- [33] Stams, A. J. M., Teusink, B. and Sousa, D. Z. 2019. Ecophysiology of Acetoclastic Methanogens. *Biogenesis of Hydrocarbons*, pp. 1–14, 2019, doi: 10.1007/978-3-319-53114-4_21-1.
- [34] Negara, I. G. A., Nindhia, T. G. T., Surata, I. W., Nindhia, T. S. and Shukla, S. K. 2021. Method on utilization of low quality biogas as a fuel for 4 stroke spark ignition engine of electric generator. *Key Engineering Materials*. Vol. 877 KEM, pp. 147–152, doi: 10.4028/www.scientific.net/KEM.877.147.
- [35] Calbry-Muzyka, A., Madi, H., Rüsçh-Pfund, F., Gandiglio, M. and Biollaz, S. 2022. Biogas composition from agricultural sources and organic fraction of municipal solid waste. *Renewable Energy*. Vol. 181, pp. 1000–1007, doi: 10.1016/j.renene.2021.09.100.
- [36] Pirzadah, T. B., Malik, B., Bhat, R. A. and Hakeem, K. R. 2022. Bioresource Technology: Concept, Tools and Experiences. *John Wiley & Sons Ltd*. doi: 10.1002/9781119789444.