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New Approaches to Sample Preparation and Integrated Spectroscopic Methods for the Identification of Polioxyethylene Triolate Sorbitane for Pharmaceutical Examination of Drugs

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Info Article	ABSTRACT		
Submitted: 07-06-2020	Due to the fast pace of development of spectroscopic research		
Revised: 01-07-2020	methods in the pharmaceutical expertise of drugs presented in the United		
Accepted: 01-09-2020	States Pharmacopeia (USP) and European Pharmacopoeia (Ph. Eur.), in this		
*Corresponding author Yerbolat Orazbekuly	paper, we examined complex methods for the identification and preparation for analysis of polyethylene sorbitan trioleate. Two new systems were identified for purification of 98% polyoxyethylene sorbitan trioleate from		
Email:	organic impurities in column chromatography: acetonitrile 100% and		
bolashik@mail.ru	acetonitrile/acetone 7.5/2.5. It was also revealed that a chamber with metallic iodine was the most suitable for selecting an eluent and controlling the cleaning by the TLC method. Proton NMR did not detect organic impurities. Identification of polyoxyethylene sorbitan trioleate was carried out on H1, C13, COSY and IR spectra. These research methods are characterized by simplicity in sample preparation, the availability of reagents, the effectiveness of identification and quantification analyzes, and the efficiency in labor and material costs.		
	Keywords: TWEEN-85, NMR, FTIR, Column Chromatography, sample preparation, pharmacopeia		

INTRODUCTION

Modern spectroscopy methods are increasingly used in the examination of pharmaceuticals (Holzgrabe, 2010). The FDA regards NMR and MS spectroscopy methods as the next generation USP (FDA and US Pharmacopeia, 2012). Proof of the development of spectroscopic methods in the examination of drugs can be attributed to a collection of works and a number of pharmacopeia methods of identification and in the analysis European pharmaceutical corporation (Holzgrabe, et al., 2008; European Pharmacopoeia, 2019). Also, the International Council for the Harmonization of Technical Requirements for the Registration of Medicines for Medical Use (ICH) by the working group of international experts "Development of analytical procedures Q14 and revision of the analytical validation procedure Q2 (R1)", where NCEM is one

of the members, considers the decision to include NIR, NMR, CE-MS, CE-ICP-MS, LC-NMR, GC-MS, LC-MS in this manual (ICH Quality guidelines, 2018).

In this paper, we examined the new complex methods of sample preparation for analysis and identification of polyethylene sorbitan trioleate (TWEEN-85), which is used in various industries, such as the food industry, pharmaceuticals, medicine, biotechnology, chemical industry, oil industry, etc. (Ema, et al 2008- Pandemic H1N1, 2009). The production of polyoxyethylene sorbitans in the world is constantly increasing. In Europe and America, people eat about 100 mg per day of polyoxyethylene sorbitan as a food supplement (Ema, et al 2008). Flu vaccines contain 25mcg of polysorbate 80 per dose (Pandemic H1N1, 2009). Polyoxyethylene sorbitans are nonionic surface active substances (nonionic surfactants), which by their nature are emulsifiers.

Nonionic surfactants weaken the surface tension at the separation of two phases - oil and water (Abramzon, 1979). There are several types of polysorbates: monolaurate is indicated by the number 20 (TWEEN -20), monopalmitate -40 (TWEEN-40), monostearate -60 (TWEEN -60), monooleate -80 (TWEEN -80) and trioleate (TWEEN -85). Polysorbates are of natural origin because they are based on sorbitol (in other words, sorbitol, glucite is a substance with a sweet taste and often used as a substitute for sugar in dietary products, it is obtained from fruits, often from fruit seeds) and fatty acids from basic oils: coconut -TWEEN -20, palm - TWEEN -40 and -60, olive -TWEEN -80 and 85.

In the countries of the European Union and in the USA, for the identification and quantification of polyoxyethylene sorbitan trioleate, mainly chromatographic methods are used, as well as the method of infrared absorption spectroscopy (European Pharmacopoeia, 2019; - United States Pharmacopeia, 2019), which include sample preparation, use and the constant presence of a high-quality reference standard (USP, EDQM) for each analysis of the drug and the shelf life of which often does not exceed 2 years.

The Republic of Kazakhstan, like other CIS countries, is a developing country. In our country, constant work is underway to harmonize the Kazakhstan Pharmacopoeia with the European Pharmacopoeia, which requires a lot of time, and therefore the Kazakhstan Pharmacopoeia is updated very lateFor the Republic of Kazakhstan, as well as other CIS countries, most of the reagents and reference standards are costly and often difficult to access due to frequent interruptions in the logistics of the necessary reagents from the countries of the European Union and the USA, which affects the quality and completeness of the examination of the drug entering the market, which is very relevant during the COVID-19 pandemic.

The presented new complex method of sample preparation is better than the previous ones in that it uses available chemicals, and the identification of polyoxyethylene sorbitan trioleate after the new method of sample preparation with the NMR spectrometer is characterized by the speed of analysis and low analysis costs. To confirm the validity of the assignment of NMR spectra, we also conducted studies on an FTIR spectrometer.

MATERIALS AND METHODS

The object of study is Tween-85 (polyoxyethylene sorbitan trioleate) (Figure 1) (li, et al 2009). The studies were conducted in the laboratory of Saint Louis University (USA). The study involved techniques such as thin-layer chromatography (TLC), column chromatography, IR spectrometry, and NMR. Column chromategraphy on alumina 60 was used to obtain pure Tween-85 for NMR and IR spectrometry studies. The solvent for the reaction product was selected by thin-layer chromatography. Solvents were distilled off by a vacuum pump with a trap of liquid nitrogen. Aldrich calibrated column with a 100mL solvent tank was used for column chromatography. Column chromatography samples were taken in 5mL tubes. Each tube was checked for impurities by thin layer chromatography on a silica plate with a pre-selected solvent. The bands on a pre-dried plate were observed under standard conditions in a TLC viewing chamber. 30min before analysis, a beaker with 2g of metallic iodine was placed in a TLC viewing camera equipped with 15W 254nm and 15W 365nm ultraviolet tubes to saturate it with iodine vapor.

NMR studies were performed on a Bruker AVANCE 3-400 spectrometer in 5mm Series 400 Thrift NMR tubes 0.01475 in wall thickness from Aldrich with deuterated dimethyl sulfoxide. To confirm the assignment of the NMR peaks, the IR spectra of polyoxyethylene sorbitan trioleate were recorded on a Shimadzu FTIR-8400S spectrometer on a PIKE Miracle nozzle with a ZnSe crystal. The spectrum of the desired sample is obtained by the ratio of scanning with the sample to scanning without a sample on the surface of the crystal. To prepare the crystal for analysis, it should be cleaned with a suitable mild solvent such as isopropyl alcohol. The sample must be in close contact with the crystal to obtain the FTIR spectrum. To obtain a spectrum, a drop of sample was placed on a crystal and data was collected.

For reliability of the result, the IR spectra of polyoxyethylene sorbitan trioleate were re-taken on a tablet from KBr. To prepare the test sample, a method of mixing a sample of 5% by weight with KBr and then compressing the tablet was used. Grinding and mixing was carried out using a mortar and pestle. A 0.5mm thick tablet obtained by compressing the prepared mixture with a hydraulic press was placed in a transmission holder for further scanning.



Figure 1. Molecular structure Tween-85 where n+w+y+z=19, some physical properties of Tween-85 are (Table I).

Table I. Key indicators of Tween-85

Indicators	Specifications
Density, g/cm ³	1,028
Refractive index	1.47-1.472
Flash point, °F	> 230
Viscosity, mPA	250-450
The content of the main	more 98%
substance	
The content of organic impurities	no more 2%

RESULTS AND DISCUSSION

For thin-layer chromatography, a silicon oxide plate was used. According to literary sources, it was found that for TLC, the acetone-water system from 8/2 to 5/5 is suitable. The authors propose to observe polyoxyethylene sorbitans on TLC in a chamber with metallic iodine (Iodine) or by spraying with 50% sulfuric acid (Katic et al., 1998). However, a system proposed from literature for TLC on a silica wafer did not perform well in alumina column chromatography. We also note that the method of spraying with 50% sulfuric acid to observe polyoxyethylen sorbitan also did not give the proper results (Katic *et al.*, 1998).

Further, for Tween-85, we found 2 systems: acetonitrile 100% and the acetonitrile-acetone 7.5/2.5 system. The systems for studying the obtained products on TLC behaved equally efficiently, both on a silicon oxide plate and on alumina column chromatography. To select an eluent and control the purification by the TLC method, a chamber with metallic iodine was used. For further purification of Tween-85 from impurities, an acetonitrile-acetone 7.5/2.5 system that we found was chosen for column chromatography from aluminum oxide. The acetonitrile-acetone 7.5/2.5 system, unlike the 100% acetonitrile system, requires less time for cleaning.

Solvent stripping was carried out by a vacuum pump with a liquid nitrogen trap. Acetone

was distilled off at room temperature (bp 56.1°C), and acetonitrile was distilled off by raising the temperature to 50° C (bp 82° C).

For investigation on an AVANCE 3-400 Bruker NMR spectrometer, 20% Tween-85 reaction solutions in deuterated dimethyl sulfoxide were taken and proton spectra (¹H), carbon spectra (¹³C) and two-dimensional COSY spectra were taken (Figure 2 and Figure 3).



PPM 5.6 5.2 4.8 4.4 4.0 3.6 3.2 2.8 2.4 2.0 1.6 1.2 0.8 0.4 0.0

Figure 2. Proton NMR spectrum of Tween-85



Figure 3. Corresponding of proton spectrum of Tween-85 in formula, where n+w+y+z=19

Chemical shift	Values
δa	0.85ppm
Δb	1.2ppm
Δc	2.0 ppm
Δd	5.3ppm
Δe	1.5ppm
Δf	2.3ppm
Δg	4.15ppm
Δh	3.5ppm
Δi	3.4ppm
Δj	3.6ppm
Δk	4.5ppm

Table II. Chemical shifts of bond groups Tween-85

¹H NMR spectrum shows large peak (h) at δ =3.5ppm belonging to two methylene protons in (OCH2CH2-)n groups and peak (b) at δ =1.2ppm belonging to two methylene protons in CH2 groups. The data on the assignment of the spectra are confirmed by the authors (Huimeng *et al.*, 2008; Gordon *et al.*, 1976). As can be seen from the proton spectrum of impurities after sample preparation is not observed, which confirms the effectiveness of the selected methods for sample preparation. To confirm the assignments of proton signals in the ¹H NMR spectrum of Tween-85, studies were conducted on COSY and ¹³C.





Figure 5. Carbon spectrum of Tween-85

Having analyzed all the cross-peaks of the COSY Tween-85 spectrum, it was found that the peak in the 3.5ppm region related to (OCH₂CH₂-)n overlaps the quartet and triplet peaks in R1-CH2(R2)-CH2R3, where R1 is an oxygencontaining heterocyclic ring, R₂ and R₃ is polyoxyethylene oleate. The peak (p) at 177 ppm corresponds to C=O bonds in the ether groups. The band (l) in the region of 130 ppm belongs to -C=Clocated in alkenes. The four peaks at 62ppm, 68 ppm, 70ppm and 74ppm correspond to 4 different CO bonds of ether groups. The first peak (k) in the 74ppm region is connected by CO attached to an oxygen-containing heterocyclic ring. The second large band (j) in the region of 70 ppm belongs to the CO bonds of the ether group in (-CH₂-CH₂-O-CH₂-CH₂-O-)n. The third peak (i) in the region of 68 ppm belongs to the CO bonds in the -CO (O) C-. The carbon-oxygen bond band (h) located in the oxygen-containing heterocyclic compound is in the region of 62 ppm.



Figure 6. IR spectrum Tween-85

The band (g) at 60ppm refers to R-CH₂-OH bonds. Deuterated dimethyl sulfoxide is located in the region of 39.5ppm. The peak (f) in the 35ppm region refers to -CH- in the R₁-CH(R₂)-R₃ bonds in the oxygen-containing heterocyclic ring, and the band (e) in the 32ppm region refers to -CH- in the R_1 -CH(R_2)- R_3 bonds in the branch attached to the oxygen-containing heterocyclic ring. The band (d) at 29 ppm refers to the -C-C- first carbon in the group (-O-CH₂-CH₂-O-CH₂-CH₂-O-). The peak (c) at 26 ppm refers to $-CH_2$ - in the $-CH_2$ - CH = CHbonds, and the band (b) at 24ppm and 22ppm are (-CH₂-). The 16 ppm peak (a) belongs to the —CH₃ group located at the end of Tween-85 tails (Gordon, 1976). IR spectra of polyoxyethylene sorbitan trioleate were recorded on an FTIR-8400S Shimadzu IR spectrometer on a PIKE Miracle nozzle with a ZnSe crystal (Figure 6). For reliable results, the IR spectra of polyoxyethylene sorbitan trioleate were re-recorded on a KBr tablet.

After analyzing the literature (li *et al*, 2009; Karic et al., 1998; Huaimeng et al., 2008; Gordon, et al., 1976; Coates 2000), we can conclude that the bands 3483 cm⁻¹ are associated with O-H groups, and the bands at 1732 cm⁻¹ to -C (0) -O-. The bands at 2918 and 2851cm⁻¹ can be attributed to asymmetric and non-asymmetric aliphatic -CH₂ groups, respectively. Absorption bands at 1631 and 1454cm⁻¹ relate to vibrations of an oxygencontaining heterocyclic ring. The 1347cm⁻¹ bands relate to CH vibrations in -CH₃ and -CH₂ groups. In addition, there is a large peak in the region of 1101cm⁻¹, which can be attributed to C-O-C asymmetric stretching vibrations in the ether bond. The bands of 949cm⁻¹ relate to symmetric vibrations of the C-O-C valence groups. The peak in the region of 723 cm⁻¹ to vibrations of (-CH₂-) n.

CONCLUSIONS

Novel complex methods for the preparation for analysis and identification of polyethylene sorbitan trioleate was studied for the first time. New system of acetonitrile/acetone 7.5/2.5 for purification of 98% polyoxyethylene sorbitan trioleate from organic impurities in column chromatography was found. To select an eluent and control the purification by the TLC method, a chamber with metallic iodine was used. The structure of sample was studied and proven by different methods of NMR and additionally verified by FTIR spectroscopy. Proton NMR did not detect organic impurities. The advantages of this method are characterized by simplicity in sample preparation, the availability of reagents, the effectiveness of identification and quantification analyzes, and the efficiency in material costs.

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