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Synthesis, and Anti-Tumor Evaluation of Some New *Flurbiprofen* Derivatives against MCF-7 and WRL-68 Cell Lines

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ABSTRACT

A new series of flurbiprofen derivatives containing thiosemicarbazide moiety (3-7) was synthesized from flurbiprofen as parent nucleus by esterification, hydrazide formation, and heating with different aryl isothiocyanate substituents, respectively. Flurbiprofen was also treated with thiosemicarbazide in the presence POCl₃ as a catalyst, to produce 1,3,4 -thiadiazole -2-amine (8). Treatment of (8) with different aryl isothiocyanates produced thiourea derivatives (9-12). Also, the reaction of (8) with different benzoyl chloride substituents produced benzamide compounds (13-15). Eventually, treatment of (8) with ethyl acetoacetate (EAA) produced [1,3,4]thiadiazolo[3,2-a]pyrimidin-7-one (16) .The new compounds were characterized by spectroscopic techniques: FTIR, 1HNMR, and CHNS analysis. A molecular docking study for the synthesized compounds (3-16), against the Vascular Endothelial Growth Factor receptor (VEGFR-2) was applied, and it indicated that compounds 4,7,13, and 15, exhibited the optimum binding energy of -6.77,-6.12,-6.68,-6.43 kcal/mol, respectively. Target compounds were also assessed for their in vitro anticancer effects in a cell-line study. All of the compounds tested showed the most plausible anticancer activity, compared to a positive control (sorafenib), using in vitro MTT cytotoxic assay, against human breast tumor (MCF-7), and normal WRL-68 cell line. The in vitro results revealed that compounds 4,5,10,11,13, and 15 exhibited the highest inhibitory activity at their IC₅₀ concentrations, against MCF-7 cell lines, as follows (122.7,113.9,95,7. 109.1,40.32 and 112.29µg/mL, respectively. While their cytotoxic effect against normal WRL-68 cell line at their IC₅₀ concentrations, as follow 210.2,181.3,151.7,278.7,80.28, and 236µg/mL, respectively. Therefore, such compounds were considered more selective toward MCF-7 than normal WRL-68, and their selectivity index (SI): 1.71,1.59,1.59, 2.55,1.99, and 2.10, respectively. Among the synthesized compounds, the compound 15 was chosen to screen its effect in vitro through multiparameter cytotoxic assay against MCF-7 breast cancer implemented in High Content Screening (ArrayScan XTI, Thermo Scientific), which could be taken in consideration as a starting point for the development of new anticancer drugs.

Keywords: Thiosemicarbazide, anti-tumor activity, thiourea, aryl isothiocyanate, high content screening.

INTRODUCTION

New anti-cancer drugs that have already been developed and discovered are a significant strategy for the treatment of malignant tumors, an incredibly touching field for medicinal chemists to find compelling yet safer chemotherapeutic drugs that focus on different biological mechanisms responsible for the development of various cancers (Magalhaes et al., 2018). Among these targets, angiogenesis is one of the basic processes affecting the growth and development of cancerous cells (Nishida et al., 2006). Angiogenesis is a complex process managed by various growth factors and cytokines. Among such indicators, vascular endothelial growth factor (VEGF) is one of the most vital angiogenic factors' involved cancer development. VEGF" has two receptors, receptor 1 (VEGFR-1) and receptor 2 (VEGFR-2) (Sadeghinia et al., 2019). VEGFR-1 and VEGFR-2 are becoming strategic goals for the improvement of anticancer drugs. Inhibition of the signaling pathway of the VEGFR-2 has an important anti- angiogenic impact on tumors, hence the Food and Drug Administration (FDA) had also approved the small molecule VEGFR-2 kinase inhibitors sorafenib, sunitinib, and pazopanib for the management of serious renal cell carcinoma (Sadeghinia et al., 2019).

In this regard, thiosemicarbazide, 1,3,4thiadiazol-2-amine, thiourea, benzamide, and [1,3,4]thiadiazole[3,2-a]pyrimidine-7-one form significant components category compounds with a broad array of applications, one of which is anticancer. Thiosemicarbazide has an anticancer activity related to its ability to suppress the enzyme ribonucleotide reductase, as seen with potent anticancer medications, like triapine and methisazone (Arora et al., 2014). According to the previous studies 1,3,4-thiadiazol-2-amine has potent VEGFR-2 kinase inhibitory effects (Tripathy et al., 2007). 1,3,4-Thiadiazole is a unique five membered rings system that has acquired noticeable quality by exploring wide spectrum of biological activity, due to the existence of the -N=C-S component. Previous literature has already shown that 1,3,4-thiadiazole-derivatives possess diverse biological activities, such as, antibacterial (Jain et al., 2013, Mathew et al., 2007), antihepatitis B infections (Kushwaha et al., 2012), antitubercular (Kushwaha et al.,2012; Foroumadi et al.,2005), anti-leishmanial (Siddiqui et al.,2009), pain relieving, and anti-inflammatory (Kushwaha et al., 2012; Mathew et al., 2007), anti-cancer (Gomha et al 2015; Gomha et al., 2015; Kumar et al.,

2006), anticonvulsants (Siddiqui et al., 2009; Sharma et al., 2013), central nervous system (CNS) depressants (Siddiqui et al., 2009), anti-oxidant (Kushwaha et al., 2012), diuretics, and antihypertensive activities (Kushwaha et al., 2012; Bhattacharva *et al.*, 2005). Recently, some moieties containing 1,3,4-thiadiazoles were investigated for their potential anti-tumor and anti-microbial effects through the inhibition of dihydrofolate reductase (DHFR) (Riyadh et al., 2018). Thiourea induces apoptosis in cancer cell lines (Huangab et al., 2017). Benzamide derivative showed potential cytotoxic action against colon HCT-116 cells (Chan et al., 2018). The chemistry and biological research on pyrimidines and their analogs have drawn extensive attention because, in addition to their impressive anticancer activities, such a ring structure is the primary skeleton in alkaloids and nucleic bases (Yousif et al., 2017).

The current research aimed to synthesizes new flurbiprofen derivatives with interesting anticancer activity through the predicted VEGFR-2 kinase inhibitors from docking study, these derivatives containing 1,3,4-thiadiazol-2-amine, thiourea, benzamide, thiosemicarbazide, and [1,3,4]thiadiazolo[3,2-a]pyrimidin-7-one moieties, were synthesized and evaluated *in vitro* for their anticancer activities against MCF-7 and WRL-68 cell lines. Since VEGFR was over expressed in MCF-7cell, therefore these cell lines were chosen in this study (Dore-Savard *et al.*, 2016).

MATERIAL AND METHODS

R-Flurbiprofen was purchased from Sigma-Aldrich."FTIR" spectra were recorded using spectrophotometer on KBr" disk, (v = cm $^{-1}$). CHNS micro-analysis was carried out using Euro EA3000 elemental analyzer, and $^{1}HNMR$ spectra were recorded on Varian Inova Spectrometer, 500MHz utilizing (TMS) as an internal standard. The chemical 'shifts were expressed as (δ =ppm), DMSO_{d6}, CDCl₃ and CD₃OD were utilized as solvents.

Chemical synthesis

The synthesis of new derivatives (Figures 1 and 2).

Synthesis of methyl 2-(2-fluoro-[1,1'-biphenyl]-4-yl) propanoate ... (1) (Telehoiu *et al.*, 2020; Al-Wabli *et al.*, 2017)

Flurbiprofen (10mmol, 2.44g) was dissolved in 30mL of abs. MeOH, then few drops of conc. H_2SO_4 were added , the resulting mixture was refluxed for 3h, and monitored with TLC.

Compound	R
3	Н
4	Br
5	Cl
6	OCH_3
7	NO_2

Figure 1. Synthesis of compounds (1-7).

Compound	R'
9	Н
10	Br
11	Cl
12	NO_2
Compound	R
13	Н
14	OCH₃
15	NO_2

Figure 2. Synthesis of compounds (8-16).

Then cooled, and neutralized by $NaHCO_3$ (5%). The product (oil) was extracted twice with diethyl ether, and then, the extracted solvent was evaporated to form compound (1).

White powder, yield 78%, m.p: $98-101^{\circ}$ C, R_f value = 0.59, IR (KBr disc, v= cm⁻¹): 3068 and 3032 (Ar-CH) str, 2943 and 2881(CH) str of *aliph*. CH & CH₃, 1890, 1915, 1952 (Aromatic overtone/combination bands), 1736 (C=0) ester str, 1171. (C-0-C) *asym.*str, 1070 (C-0-C) 0CH₃ *sym* str, 1577, 1560 and 1483 Ar(C=C) str.

CHNS analysis: Calcd. for $(C_{16}H_{15}FO_2)$: C: 74.4, H: 5.85. Observed: C: 74.20, H: 5.32, ¹HNMR (500MHz, CDCl₃, δ =ppm); 7.26-7.03 (8H, m, Ar-H);

3.93 (1H, q, CH-aliph.); 3.69 (3H, s, OCH₃); 1.53 (3H, d, CH₃).

Synthesis of 2-(2-fluoro-[1,1'-biphenyl]-4-yl) propanehydrazide ... (2) (Parmar *et al.*, 2011; Koopaeia *et al.*, 2013)

(5.0mmol 1.3g) of compound (1) was dissolved in (20) mL of abs. EtOH, then (10.0 mmol, 0.33ml) of NH $_2$ NH $_2$ H $_2$ O was added slowly with stirring for 2h at RT, then the mixture refluxed for 6h. The solvent was evaporated, the mixture was allowed to cool, and the precipitate was already washed with ice-cold water several times, and recrystallized from MeOH.

White precipitate, yield 82%, m.p: 77-79°C, R_f value= 0.88. IR (KBr disc, (v=cm $^{-1}$): The Shoulder at 3210 and peaks at 3309, and 3178 (NH) *prim*. amine str, 3030 Ar (CH) str, 2978, 2933 and 2879 (CH) str of *aliph*.CH & CH₃, 1900, and1961 (Aromatic overtone/combination bands), 1635 (C=O) of amide hydrazide str, 1523, 1483 and 1415 Ar(C=C) str. CHNS analysis: Calcd. for (C₁₅H₁₅FN₂O): C: 69.75,H: 5.85, N: 10.85. Observed: C: 69.25, H: 5.36, N: 10.72.

¹HNMR (500MHz, CDCl₃, δ=ppm) 7.28-7.02 (8H, m, Ar-H); 3.87 (2H, *br*s, NH₂); 3.55 (1H, q, CH-*aliph*.); 1.55 (3H, d, CH₃).

General method for synthesis of the compounds (3-7)..... (Hui *et al.*, 1994; Al-Saad *et al.*, 2019)

(2.0 mmol, 0.517g) of compound (2) was dissolved in (20mL) abs. MeOH, then (2.0 mmol) of each different substituted aryl-isothiocyanates: (0.24mL) phenylisothiocyanate; (0.43g) p.bromo phenylisothiocaynate; (0.338g) p.chloro phenylisothiocyanate; (0.28mL) p. methoxy phenylisothiocyanate, and (0.360g) p.nitro phenylisothiocaynate, each was added separately with stirring. The temperature of the reaction was kept at 40-50°C. The precipitate was filtrated, then washed with EtOAc, and recrystallized from EtOH.

Compound (3): 2-(2-(2-fluoro-[1,1'-biphenyl]-4-yl)propanoyl)-N-phenylhydrazine-1-carbothioamide.

White precipitate, yield 72%, m.p: 163-165°C, R_F value = 0.37, IR (KBr disc,v= cm⁻¹): 3346, 3246 and 3136 (NH) str of (1stamide, 2nd amine and NH adjacent to thiocarbonyl), 3060 and 3032 Ar (CH) str, 2966, and 2879 (CH) str of *aliph*. CH & CH₃, 1944 (Aromatic overtone), 1676 (C=0) of amide hydrazide str, 1595, 1537, 1508, and 1448 Ar(C=C) str, 1190 (C=S) str. CHNS analysis: Calcd. for ($C_{22}H_{20}FN_3OS$) C: 67.16, H: 5.12, N: 10.68, S: 8.15. Observed: C: 67.86, H: 4.85, N: 11.00, S: 8.55. , HNMR (500MHz, DMSO_{d6}, δ =ppm); 10.15 (1H,s, NH); 9.51 (1H,s, NH); 7.70-7.05 (13H,m, Ar-H); 3.75 (1H,d, CH-*aliph*.); 1.40 (3H,d, CH₃).

Compound (4): N-(4-bromophenyl)-2-(2-(2-fluoro-[1,1'-biphenyl]-4-yl)propanoyl)hydrazine-1-carbothioamide.

White precipitate, yield 65%, m.p:179-181°C, R_f . value = 0.48, IR (KBr disc, v= cm⁻¹): 3239, 3184 br (NH) str of (1st amide, 2nd amine and NH adjacent to thiocarbonyl), 2978 (CH) str of aliph.CH & CH₃, 1886 (Aromatic overtone),1685 (C=0) str of

1stamide, 1593,1545,and 1487 Ar(C=C) str, 1176 (C=S)str, 766 (C-Br) str, 823 (Aromatic sp² CH) bend. of *para* position. CHNS analysis: Calcd. for (C₂₂H₁₉BrFN₃OS) C: 55.94, H: 4.05, N:8.90, S: 6.79 . Observed: C: 54.94, H: 3.31, N: 8.64, S: 6.74 . ¹HNMR (500MHz, DMSO_{d6}, δ=ppm); 10.15 (1H,s, NH); 9.69 (1H, s, NH); 7.51(2H, d, 2Ar-H); 7.48 (2H,d, 2Ar-H); 7.46-7.27 (m,8H,Ar-H); 3.76 (1H, q, CH-*aliph*.); 1.43 (3H, d, CH₃).

Compound (5): N-(4-chlorophenyl)-2-(2-(19-20)-2-(19-20)-4-yl) (2-carbothioamide).

Pale white precipitate, yield 45%, m.p. 190-192°C, R_f . value = 0.27, IR (KBr disk $v = cm^{-1}$): Shoulder at 3334, 3238, 3182 br (NH) str of (1st amide, 2nd amine and NH adjacent to thiocarbonyl), 2978, and 2883 (CH) str of aliph. CH & CH₃, 1866 (Aromatic overtone), 1685 (C=0) str of 1st amide; 1593, 1545, 1487 and 1415 Ar-(C=C) str, 1178 (C=S) str, 1072 (C-Cl) str, 823 (Aromatic sp² CH) bend. of para position. CHNS analysis: Calcd. for (C₂₂H₁₉ClFN₃OS) C: 61.75, H: 4.48, N: 9.82, S: 7.49. Observed: C: 61.32; H: 4.96, N: 8.94, S: 6.81, ¹HNMR (500MHz, DMSO_{d6}, δ=ppm); 10.16 (1H, s, NH); 9.68 (1H, s, NH); 7.51(d,2H,Ar-H); 7.46 (2H, d, Ar-H); 7.45-7.27 (8H, m, Ar-H); 3.76(1H,q, CH-aliph.); 1.43 (3H, d, CH₃).

Compound (6): 2-(2-(2-fluoro-[1,1'-biphenyl]-4-yl)propanoyl)-N-(4-methoxyphenyl) hydrazine-1-carbothioamide.

Pink precipitate, yield 62%, m p:215-218°C, R_f : value = 0.38, IR (KBr disc,v = cm⁻¹): 3336,3309 and 3138 (NH) str of (1st amide, 2nd amine and NH adjacent to thiocarbonyl),3066,3033 Ar(CH) str, 2964, and 2839 (CH) str of *aliph*. CH & CH₃, 1684 (C=O) str of 1st amide,1543, 1510 and 1450 Ar(C=C) str, 1250 *asym* (C-O) str, 1176 (C=S) str, 1030 *sym* (C-O) str, 837 (Aromatic sp² CH) bend. of *para* position. CHNS analysis: Calcd. for (C₂₃H₂₂FN₃O₂S) C: 65.23, H: 5.24, N: 9.92, S: 7.57. Observed: C: 64.90, H: 4.82, N: 9.36, S: 7.42., ¹HNMR (500MHz, DMSO_{d6}, δ=ppm); 10.11 (1H, s, NH); 9.46 (1H, s, NH); 7.50-7.23 (10H, m, Ar-H); 6.88 (2H, d, 2Ar-H); 3.75 (1H, q, CH-*aliph*.); 3.69 (3H, s, OCH₃); 1.43 (3H, d, CH₃).

Compound (7): 2-(2-(2-fluoro-[1,1'-biphenyl]-4-yl)propanoyl)-N-(4-nitrophenyl)hydrazine-1-carbothioamide.

White precipitate, yield 48 %, m.p: 192-195°C, R_f value = 0.26, IR (KBr disc, $v = cm^{-1}$): 3298 overlap (NH) str of (1st amide and 2nd amine); 3155

(NH) str adjacent to thiocarbonyl, 3055 Ar(CH) str; 2978, 2939 and 2833 (CH) str of *aliph*. CH & CH₃, 1685 (C=O) amide str; 1618 and 1574 Ar (C=C) str, 1184 (C=S) str, 1504 (NO) *asym*. str, 1333 (NO) *sym* str, 853 (Aromatic sp² CH) bend. of *para* position. CHNS analysis: Calcd. for ($C_{22}H_{19}FN_4O_3S$): C: 60.26, H: 4.37, N: 12.78, S: 7.37. Observed: C: 59.17, H:3.98, N:12.42, S:7.54. HNMR (500MHz, DMSO_{d6}, δ =ppm); 10.03 (1H, s, NH); 8.19 (2H, d, Ar-H); 7.86(d,2H,Ar-H); 7.52-7.27 (8H, m, Ar-H); 3.79 (1H, q, CH-*aliph*); 1.44 (3H,d, CH₃).

Synthesis of 5-(1-(2-fluoro-[1,1'-bi phenyl]-4-yl) ethyl)-1,3,4-thiadiazol-2-amine..... (8) (Tomi *et al.*,2014; Nahi *et al.*,2019)

A mixture of flurbiprofen (5.0mmol, 1.22g), thiosemicarbazide (5.0mmol, 0.455g,) and $POCl_3$ (5mL) were refluxed for 4h. Then the mixture was cooled and ice water (25mL) was added gradually and the mixture of reaction was refluxed for further 3h, then cooled and filtered. Neutralization of the filtrate with KOH solution (10%) and the precipitate was also filtered, and recrystallized from MeOH.

Brown precipitate, yield 62%., m.p 216-219°C, R_f .value= 0.38, FTIR (KBr, $v=cm^{-1}$): 3280 and 3118 str for *prim*. (NH₂), 3035 (Ar-CH) str, 2976 , 2933 and 2875 (CH) str. of *aliph*.-CH & CH₃,1952, 1892 (Aromatic overtone/combination bands), 1622,1608 (Ar-C=N)str, 1514,1485,1415 (Ar-C=C) str, 1130 (N-N)str, 766 and 725 C-S str. CHNS analysis: Calcd. for (C₁₆H₁₄FN₃S) C: 64.19 , H:4.71 , N: 14.04, S:10.71. Observed: C:63.85, H: 4.81, N: 13.80, S: 10.45. 1 H- NMR (500MHz,CD₃OD, 5 eppm); 7.46-7.07 (8H,m, Ar-H); 4.45 (1H,q, CH*aliph*.); NH₂(masked by CD₃OD-solvent); 1.68 (3H,d,CH₃).

General method for synthesis of 1-(4-chlorophenyl)-3-((5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-ylthiourea derivatives...(9-12) (Rahman *et al.*, 2014)

A mixture of compound (8) (0.375g, 1.25 mmol), and (1.25 mmol) of different substituted aryl isothiocyanates: phenyl isothiocyanate (0.15mL), *p*.bromo phenyl isothiocyanate (0.276g), *p*.chloro phenyl isothiocyanate (0.212g), *p*.nitro phenyl isothiocyanate (0.225g), each was added separately in acetone (20mL), and heated under reflux for 24h. The reaction mixture was then evaporated, left to cool, and the solid formed was already filtered, washed with acetone, dried and

recrystallized from EtOAc-MeOH to yield the desired compound.

Compound 9: 1-((5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)-3-phenylthiourea.

White precipitate, yield 49%, m.p:193-196 °C, R_f :value = 0.46, IR (KBr disc, $v = cm^{-1}$): 3456, 3323, 3261 and 3197 (NH) str, 3089 and 3028 (CH) str of aromatic CH, 2979, 2877 (CH) str, of *aliph*. CH and CH₃, 1597, 1550, 1489 and1435 Ar(C=C) str, and 1657Ar(C=N)str, 1234 (C=S) str, 1128 (N-N)str, 756 C-S str. CHNS analysis: Calcd. for (C₂₃H₁₉FN₄S₂) C: 63.57, H: 4.41 , N:12.89 , S:14.76 . Observed: C: 64.01, H: 3.98, N: 12.12, S: 13.93. 14 HNMR (500MHz, (CD₃)₂CO, δ =ppm); 7.67-7.09 (13H, m, Ar-H); 4.40 (1H, q, CH-*aliph*.); 1.42 (3H, d, CH₃).

Compound 10: 1-(4-bromophenyl)-3-((5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)thiourea

Pale white precipitate, yield 52%, m.p:202-205°C, R_f .value = 0.47, IR (KBr disc, $v = cm^{-1}$): shoulder at 3200, 3128 for (NH)str, 3060 (Ar-CH) str, 2987, 2935, (CH) str of *aliph*. CH and CH₃, 1512, 1481 and 1414 Ar(C=C), and 1601& 1624 Ar(C=N) str, 1263 (C=S) str, 1136 (N-N)str, 739 (C-S) str, 768 (C-Br) str, 854 and 833 (Aromatic sp² CH) bend. of *para* position . CHNS analysis: Calcd. for (C₂₃H₁₈BrFN₄S₂) C: 53.80 ,H:3.53, N:10.91 ,S:12.49 . Observed: C: 54.86, H:3.27 ,N:10.44,S:13.02-1HNMR (500MHz, DMSO_{d6}, δ=ppm); 10.38 (1H, s, NH); 7.73 (2H, d, Ar-H); 7.55-7.31 (10H, m, Ar-H); 4.55 (1H, q, CH-*aliph*.); 1.68 (3H, d, CH₃).

Compound 11 : 1-(4-chlorophenyl)-3-((5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)thiourea

Pale white precipitate , yield 44%, m.p: 207-209°C, R_f .value = 0.47, IR (KBr disc, $v = cm^{-1}$): 3444,3323,3261 and 3197 str for (NH),3076 3026 (Ar-CH) str, 2978, 2929and 2883 (CH) str of *aliph*. CH and CH₃, 1658 Ar(C=N) str, 1597, 1545, 1489 and 1433 Ar(C=C) str, 1230 (C=S) str, 1124 (N-N)str, 768 (C-S)str, 1099(C-Cl)str, 831 (Aromatic sp² C-H) bend. of *para* position. CHNS analysis: Calcd. for $C_{23}H_{18}CIFN_4S_2$) C: 58.90 ,H:3.87 ,N:11.59 , S:13.67. Observed: C:58.84 ,H: 4.11, N:11.43, S:12.95. 1 HNMR (500MHz, DMSO_{d6}, δ=ppm); 10.38 (1H, s, NH); 7.72 (2H, d, Ar-H); 7.53-7.32 (10H, m, Ar-H); 4.55 (1H, q, CH-*aliph*.); 1.68 (3H, d, CH₃).

Compound 12: 1-((5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)-3-(4-nitrophenyl)thiourea

White precipitate, yield 44%, m.p. 207-209°C, R_f value = 0.65, IR (KBr disc, $v = cm^{-1}$): 3435, 3236 str of (NH), 3037 (Ar-CH) str, 2978, 2935and 2875 (CH) str of *aliph*. CH and CH₃, 1628, 1592 Ar(C=N) str, 1485 and 1446 Ar(C=C) str, 1275 (C=S) str, 1223 (N-N)str, 762 (C-S)str, 1535. (NO) *asym*. str, 1327 (NO) *sym*. str, 829 (Aromatic sp² CH) bend. of *para* position. CHNS analysis: Calcd. for (C₂₃H₁₈FN₅O₂S₂) C: 57.61, H: 3.78, N:14.60, S:13.37. Observed: C 58.01, H:3.91, N:14.12, S:12.89. ¹HNMR (500MHz, DMSO_{d6}, δ =ppm); 10.86 (1H, s, NH); 8.15 (2H, d, Ar-H); 8.05 (2H, m, Ar-H); 7.55-7.29 (8H, m, Ar-H); 4.59 (1H, q, CH-*aliph*.); 1.70 (3H, d, CH₃).

General method for synthesis of the compounds (13-15) (Issaca *et al.*, 2012)

(0.374g, 1.25mmol) of the compound (8) was dissolved in (20mL) dry benzene containing triethylamine (3drops) as a catalyst, 1.25mmol of different substituted benzoyl chlorides: benzoyl chloride (0.16mL), *p*-methoxy benzoyl chloride (0.17mL), *p*-nitro benzoyl chloride (0.232g), each was added separately, drop wise to an icecooled, stirred solution. The mixture was stirred at RT, then refluxed, until reaching the end after 4h, consequently, cooled at RT. The separated solid was filtered off, and recrystallized from chloroform to produce compounds (13-15).

Compound 13: N-(5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)benzamide

White precipitate, yield 67%, m.p: 185-187 °C, R_f . value = 0.56, IR (KBr disc, $v = cm^{-1}$): Shoulder at 3240, and peak at 3155 (NH) str of amide, 3053 and 3035 Ar (CH) str, 2976 and 2935 (CH) str of *aliph*. CH & CH₃, 1670 (C=0) of amide str, 1622 Ar(C=N) str, 1593., 1537, and 1485,Ar(C=C) str, 1228 (N-N)str, 744 (C-S)str, CHNS analysis: Calcd. for ($C_{23}H_{18}FN_3OS$) C: 68.47, H:4.50, N:10.41,S:7.95. Observed: C 67.97, H: 4.86, N:10.84, S:7.67. ¹HNMR (500MHz, DMSO_{d6}, δ =ppm); 12.98 (1H, s, NH); 8.08 (2H, d, Ar-H), 7.63-7.37 (11 H, m, Ar-H); 4.73 (1H, q, CH-*aliph*.); 1.77 (3H, d, CH₃).

Compound (14): N-(5-(1-(2-fluoro-[1,1'-biphenyl] -4-yl)ethyl)-1,3,4-thiadiazol-2-yl)-4-methoxybenzamide.

White precipitate, yield 70%,m.p:192-195°C, R_f = 0.68, IR (KBr disc, v = cm $^{-1}$): Shoulder at 3220 and peak 3163 (NH) str of amide, 3057,3035 Ar (CH) str, 2976, 2937 and 2800 (CH) str of *aliph*. CH & CH₃,1664 (C=O) of amide str, 1612 Ar(C=N)

str, 1533, 1510 and 1480 Ar(C=C) str, 1261 asym. (C-O) str, 1184 (C=S)str, 1041 (C-O) sym. str, 1130 (N-N)str, 742 (C-S)str, 837 (Aromatic sp² CH) bend. of para position. CHNS analysis: Calcd. for (C₂₄H₂₀FN₃O₂S) C: 66.50, H: 4.65,N:9.69 ,S:7.40. Observed: C 65.90, H: 4.88, N: 9.59, S:7.64. ¹HNMR (500MHz, DMSO_{d6}, δ =ppm); 12.89 (1H, s, NH); 7.99 (2H, d, Ar-H); 7.54-7.31 (10 H, m, Ar-H); 4.72 (1H, q, CH-aliph.); 2.36 (3H, s, OCH₃); 1.76 (3H, d, CH₃).

Compound (15): N-(5-(1-(2-fluoro-[1,1'-biphenyl]-4-yl)ethyl)-1,3,4-thiadiazol-2-yl)-4-nitrobenzamide

White precipitate, yield 65%, m p 197-200 °C , R_f . value = 0.47, IR (KBr disc, $v = cm^{-1}$): 3172, (NH) str of amide, 3118 ,3055 Ar (CH) str, 2976 , 2935 and 2870 (CH) str of aliph. CH & CH₃, 1678 (C=0) of amide str, 1604 Ar(C=N) str, 1483 Ar(C=C) str, and 1531 (NO) asym str, 1344 (NO) sym str, 1130 (N-N)str, 704 (C-S) str, 845 (Aromatic sp² CH) bend of para position. CHNS analysis: Calcd. for (C₂₃H₁₇FN₄O₃S) C:61.60 ,H:3.82, N:12.49, S:7.15. Observed: C 61.32, H:3.69, N:11.95, S:7.09. 1 HNMR (500MHz, DMSO_{d6}, δ =ppm); 8.34 (2H, d, Ar-H)); 8.27(d,2H,Ar-H); 7.53-7.31 (8 H, m, Ar-H); 4.73 (1H, q, CH-aliph.); 1.76 (3H,d, CH₃).

Synthesis of 2-(1-(2-fluoro-[1,1'-biphenyl]-4-yl) ethyl)-5-methyl-7H-[1,3,4]thiadiazolo[3,2-a]pyrimidin-7-one(16) (Rahman *et al., 2014*)

(1.25mmol, 0.374g) of compound (8) and ethyl acetoacetate (EAA) (1.25 mmol, 0.162g,) were heated at 160° C in oil bath for 6h, upon completion of the reaction, the mixture was therefore cooled at RT. The separated solid was filtered off and recrystallized from chloroform.

White powder, yield 78%, m. p 240-242°C, R_f . value = 0.68, IR (KBr disc, $v = cm^{-1}$): 3075,3060 Ar (CH) str, 2914 and 2804 (CH) str of *aliph*. CH & CH₃, 1711cm (C=O) str of amide, 1624 Ar(C=N)str, 1572, 1481 Ar(C=C) str, and 1161 (N-N)str,768 and 739 (C-S)str.

CHNS analysis: Calcd. for ($C_{20}H_{16}FN_{3}OS$) C: 65.74, H:4.41, N:11.50, S:8.77 . Observed: C: 64.83, H:4.23, N:10.92,S:7.97. ¹HNMR (500MHz, CDCl₃, δ =ppm); 7.29-7.02 (m, 8H, Ar-H +1H, CH=C-alkene); 4.53 (1H, q, CH-aliph.); 2.03-1.24 (6H,m, 2xCH₃).

Molecular docking study

All the molecular docking studies were carried out using MOE, 2014 software. With a force field tool, energy minimization was executed with (RMSD) gradient of 0.05kcal/mol (Brooks *et al.*, 2009), errors were fixed and the partial charges were computed. The protein data bank file (PDB:

(20H4) (El-Feky *et al.*, 2015) was selected for the pocket of Vascular Endothelial Growth Factor Receptor-2 (VEGFR-2). The enzyme was prepared for docking studies through the following steps (i) water molecules were removed (ii) adding of hydrogen atoms to the enzyme; (iii) the preparation of the reference ligand sorafenib (iv) the removal of the co-crystallized ligands and docking of the new targeted structures (Coutsias *et al.*, 2004).

Biological screening

The study of MTT and HCS was carried out using the procedure outlined earlier in the literature (Sebeka *et al.*, 2017; El-Far *et al.*,2009).

MTT assay

The MTT assay would be used to estimate the cytotoxicity of the synthesized compounds (3-7) and compounds (9-16) against MCF-7 and WRL-68 normal cell lines that use sorafenib as a reference drug.

Dimethyl-sulfoxide (DMSO.), was added to each of the synthesized compounds, to prepare a stock solution and serial dilutions (6.25-400μg/mL). 100μL of each of the samples and sorafenib (positive control), were added to MCF-7, and WRL-68 cell lines, and the cell culture was kept in the CO₂ incubator at (37°C) for 24h. Addition of 20mL of MTT stock solution (6μg./ mL) to each well and plates followed, then incubation for 1-4h ,when the media was removed, DMSO was already introduced per each well to dissolve the crystals of the formazan .The absorbance was validated at 575nm, through the use of a Hidex-Chameleonmicroplate reader (Lab Logic Systems- Ltd, Sheffield, United Kingdom). The small portion of the absorbance of the cells treated with the synthesized compounds compared to absorbance of untreated cells, (negative control) was calculated as a percentage of cell viability. The IC₅₀ values were calculated for each target compound (Abraham et al., 2008; Hmood et al. 2020).

High-Content Screening (HCS)

Three kits of Cellomics multi parameter cytotoxicity (Thermo Scientific, Japan) were also used to detect viable cell counts, nuclear, intensity, cell membrane permeability, mitochondrial membrane potential, and cytochrome C levels in the MCF-7 cell lines after treatment with compound (15). Briefly. 24h after compound's 15 treatment, mitochondrial membrane potential (MMP) dye, and the cell permeability dye were already added

to the cells and allowed to stand for 30min at 37°C. Cells were already fixed, permeabilized, and blocked with 1X blocking buffer prior probing with primary cytochrome C antibody and secondary DyLight TM 649 conjugated goat anti-mouse IgG for one h each. The plates were assessed by using the ArrayScan high content screening (HCS) system. A detailed study was carried out earlier in the literature (Looi *et al.*, 2013).

Caspases 7 and 9 Activity Assay

The caspase activity assay was performed with the use of of Caspase Glo7 and 9 kit (Promega-Madison-WI (Hamed et al., 2014). Briefly, MCF-7 cells were grown into 96-well plates, with various concentrations of compound (15) added for 24h. 100μL of Glo 9 reagent was added and allowed to stand for 30min at 25°C, the effect of the caspase in the treated cells was determined by the extent of amino-luciferin-labeled tetra peptide cleavage, and luciferase enzyme substrates release, using a Tecan-Infinite 200 Pro micro plate readers. Furthermore, time-dependent mode experiments were also performed to validate the expression degree of caspases and hence, MCF-7 cells had treated with compound heen (15)different concentrations. Then the assessment of the degree of expression of caspases 7 and 9 at different times was carried out (Paydar et al., 2014).

Statistical analysis

One way analysis of variances (ANOVAs) was used to compute whether the variances were statistically significant; data were calculated as the mean ± standard deviation, and statistically relevant results were analyzed using Graph Pad PrismV6 (Graph Pad program, Inc.).

RESULTS AND DISCUSSION Docking study Molecular docking processes

Molecular docking studies have been shown that the target compounds may be considered as potential inhibitors of the Vascular Endothelial Growth Factor (VEGFR-2) receptor. Vascular endothelial growth factor (VEGF) is over expressed in MCF-7 cell, and it is one of the most vital angiogenic factors associated with the development of the cancers.

All experimental models were produced using MOE 2014.0901 software. Throughout every experiment, cells that express VEGFR-2 were obtained from the Protein Data Bank (PDB ID 20H4) (El-Feky *et al.*, 2015).

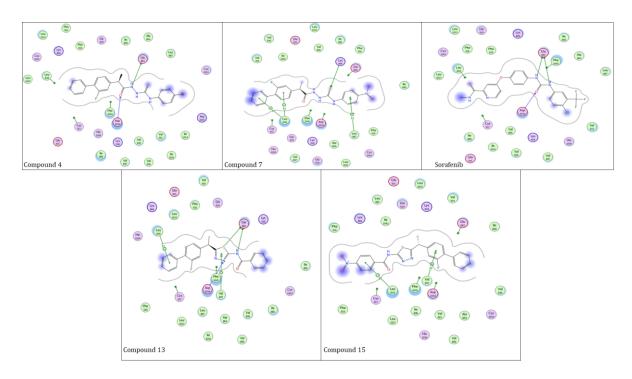


Figure 3. 2D Crystal ligand of some synthesized compounds (4,7,13 and 15), and sorafenib as potent inhibitor against VEGFR-2 pocket

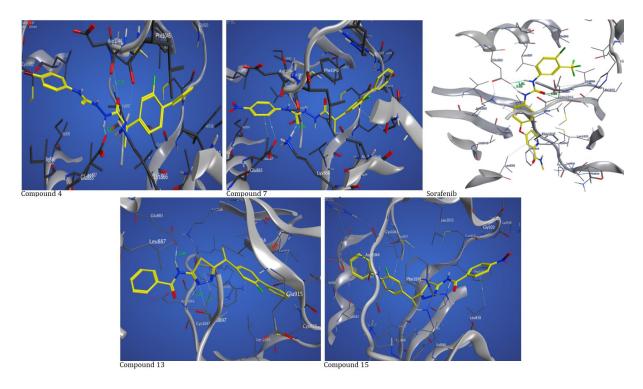


Figure 4. 3D binding mode of some synthesized compounds (4,7,13 and 15) and sorafinib (Yellow) docked in crystal protein of VEGFR-2 (PDB: 20H4) and the corresponding interaction diagram, hydrogen bonds with green color and pi-interaction with gray color.

Table I. Show ((AG) kca	I/mol of	the tested com	pounds against	(VEGFR-2)	target site PDB ID: 20H4.

Comp No	Bonds		Interaction with key	Score (ΔG)	RMSD	E-place	Bonds length
Comp. No.	H.B	<i>p</i> i	amino acids	Kcal /mol	value		range/Å
3	2	1	Glu883-Asp1044	-4.22	1.37	-96.61	1.90-1.98
4	2	0	Glu883-Asp1044	-6.77	3.58	-80.96	2.31-165
5	2	1	Glu883-Asp1044-Cys917	-5.02	1.03	-70.33	1.62
6	2	0	Glu883-Asp1044	-3.24	1.56	-60.52	2.95-2.85
7	1	3	Lys866-Leu887-Leu838	-6.12	1.09	-99.70	3.76
9	1	2	Leu838-leu887-Glu883	-3.48	1.91	-94.86	1.73
10	0	2	Val 897-Leu838	-4.65	1.75	-88.38	-
11	0	2 Val 897-Leu838		-5.09	1.70	-85.69	-
12	0	2	Phe1045-leu838	-3.89	0.88	-114.8	-
13	3	2	Glu883-Asp1044-Val897	-6.68	2.05	-89.9	2.09
14	1	0	Asp1044	-6.59	1.98	-97.69	1.71
15	15 0 2 Val897-1		Val897-Leu838	-6.43	2.16	-110.39	-
16	1	1 1 Cys917-Val 897		-1.17	0.79	-83.8	2.13
Sorafenib	3	0	Glu883-Asp1044	-9.71	1.22	-110.87	1.81-2.03

Table II. Predicted ADMET analysis for the target compounds (3-16).

Comp. No.	BBB level	HIA	CYTP450 3A4	CYP2D6	CYP2C9	PPB	Solubility level
3	4.120	94.59	0	0	0	98.20	1
4	5.720	95.24	0	0	0	91.60	1
5	5.550	93.03	0	0	0	95.50	1
6	3.220	94.87	0	0	Inhibitor	96.40	2
7	0.510	93.41	0	0	inhibitor	97.70	2
9	0.708	96.31	0	0	0	92.77	2
10	1.830	97.30	0	0	0	91.10	2
11	1.670	79.02	0	0	0	91.51	3
12	0.036	69.81	0	0	0	92.5	2
13	0.048	96.61	0	0	0	93.9	2
14	0.043	96.6	0	0	0	93.33	2
15	0.580	98.16	0	0	0	91.07	1
16	0.047	97.70	0	0	0	92.96	2

Solubility level: (4) high sol., (3) and (2) intermediate sol., (1) less sol., (0) poor sol; BBB: Blood-Brain Barrier; HIA: Human Intestinal Absorption; CYP2D6: Cytochrome P2D6, 0= non-inhibitor; PPB: Plasma Protein Binding.

The results showed that all ligands investigated had a quite comparable position and orientation within the defined binding site of VEGFR-2, which explains a vast area bounded by a membrane-binding domain.

The outcome of the free energy of binding (ΔG) stated that some of these compounds might have high binding affinity toward the receptor, and the computed values reflected the general pattern, (Table I). The key binding sites of (VEGFR-2) was reported in the literatures(El-Adl *et al.*, 2020), consisting of amino acids Tyr644, Ile619, Leu746, Thr692, Lys646, Glu706, Lys702, and Glu663.

At a distance of 1.65° A.The binding mode of compound **4**, revealed an energy binding of -6.77 kcal/mol. The nitrogen group at hydrazide moiety formed a hydrogen bond with Glu883. Also, the (C=0) group formed a hydrogen bond with Asp1044 at a distance of 2.31° A.

Compound 7 revealed binding energy of -6.12 kcal/mol. The sulfur group at thiourea moiety formed a H-bond with Lys866 at a distance of 3.76°A. While the hydrophobic phenyl rings formed three *Vander Waals* interactions, one with Leu887, and two bindings with Leu838.

Table III. Carcinogenicity prediction of the synthesized compounds (3-16).

Comp. No.	Ames test	Carcinogen on mouse	Carcinogen on rat	HERG inhibitor	Carcinogenicity	TA100-NA
3	Mutagen	Positive	Positive	High risk	1	-
4	Mutagen	Positive	Negative	-	0	-
5	Mutagen	Positive	Negative	High risk	0	-
6	Mutagen	Positive	Negative	Low risk	0	-
7	Non Mutagen	Negative	Negative	-	0	-
9	Mutagen	Positive	Negative	high risk	1	-
10	Mutagen	Positive	Positive	Med. Risk	1	-
11	Mutagen	Positive	Negative	Med. Risk	0	-
12	Mutagen	Negative	Negative	high Risk	1	-
13	Mutagen	Positive	Negative	Med. Risk	0	-
14	Mutagen	Negative	Positive	Med. Risk	0	-
15	Mutagen	Negative	Negative	Med. Risk	0	-
16	Mutagen	Positive	Positive	Med. Risk	0	-

carcinogen = 1; may be non-carcinogen = 0

Table IV. C log p, molecular volume (MV), topological polar surface area (TPSA), and percent of absorption of the synthesized compounds

Comp. No	Clogp	MV	n-Violation	TPSA	% *(Abs)
3	4.92	349.23	0	53.14	91
4	5.73	367.12	1	53.15	91
5	5.60	362.77	1	53.15	91
6	4.98	377.78	0	62.39	87.5
7	4.88	372.57	0	98.98	75
9	5.32	371.60	1	99.89	74.5
10	6.13	513.46	2	49.84	92
11	6.00	385.19	1	49.15	92
12	5.28	344.49	1	95.66	76
13	5.00	350.57	1	54.88	90
14	5.16	433.51	1	64.12	87
15	5.06	377.70	1	100.71	74.25
16	4.23	312.48	0	47.7	92.5

%*(Abs.) calculated according to a method described by (*Zhao,2002 et al.*,).

Compound **13** presented an energy binding of -6.68 kcal/mol. The thiadiazole ring formed two H-bonds with Glu883 and Asp1044 at a distance of 2.25 Å. Also, the secondary amido (NH) group formed a H-bond with Glu883 at a distance of 2.09° A. While the phenyl and thiadiazole rings formed two hydrophobic interactions with Leu838, and Val 897, respectively. The mode of binding of compound **15** exhibited an energy binding of 6.43kcal/mol. The *p*-nitro phenyl ring, and *m*-flouro phenyl ring formed two hydrophobic interactions with Leu838 and Val897, respectively. Finally, It is important to mention the binding mode of sorafenib, as a reference, which exhibited an energy binding of -9.71 kcal/mol. The

group at urea moiety formed two H-bonds with Glu883 at a distance of 1.81 –2.03 Å. Also, the (C=0) formed a H-bond with Asp1044 at a distance of 1.9 Å. All the above bindings mode 2D and 3D simulation attachment against (VEGFR-2) critical amino acids, length of hydrogen bonds/Å, are presented in (Table I and Figures 3 and 4).

In silico ADMET and carcinogenicity analysis

The pharmacokinetic profile, i.e. absorption, distribution, metabolism, and excretion was anticipated by ADMET descriptor module of the small molecules protocol of *pre* ADMET online software, http://preadmet.bmdrc.kr/adme/. (Table II).

The synthesized derivatives exhibited higher penetration to blood brain barrier (BBB). In addition, all of them possessed good human intestinal absorption (HIA) from the intestine following oral absorption, the HIA value ranged from 69.81 -98.16 (Eissa et al., 2018, Al-Saad et al., 2020). It was also noted that their aqueous solubility logarithmic level ranged from (1-3) in most of the compounds, reflecting low to moderate aqueous solubility. The plasma protein binding model showed good binding ability of the synthesized compounds to plasma proteins. All of the marked compounds were found to be non-inhibitors of CYTP450 3A4, CYP2D6 and CYP2C9, except for compounds 6 and 7 which were reported to be inhibitors of enzyme CYP2C9 (Table II).

Finally, (Table III) indicated that the series of the synthesized compounds are non–carcinogenic, except for compounds **3,9,10**, and **12(**carcinogenicity score=1) .While, the rest of the compounds are assumed to be safe.

Physico-chemical properties of the synthesized compounds (3-16)

The values of Clogp, the topological polar surface area (TPSA) and molecular volume (MV) were calculated using Chem-informatics on the Web (http://www.molinspiration.com), (Table IV). The Clog p value gives an indication of the lipophilicity of the chemical compound. The increasing trend in this value leads to a rise in the lipophilic character of the compound tested (Rudraraju et al., 2014). It is worth mentioning that the Clog p values for the tested compounds ranged from 4.23 to 6.13. The tested compounds 3, 6, 7, and 16 are in clear violation of Lipiniski's rule of five. Further, the topological polar surface area (TPSA) of the new derivatives ranged from 47.70 to 100.71, with a reasonable oral bioavailability (Veber et al., 2002), and acceptable molecular volume (MV) values. The percentage of absorption (percent Abs.) was determined, according to the method outlined by Zhao et al. (Zhao et al., 2002) (Table IV).

Pre-metabolism analysis

The finding of the *pre*-metabolism investigation on compound (13) was taken as a prototype for the predicted metabolism analysis, revealing that CYTP450 3A4 liver microsomal enzyme had a small influence on the compound tested as CYP2C9 liver microsomal enzyme, with

the most metabolic reaction taking place to be benzene hydroxylation, whose product would be relatively easy to execrate it from the human body, (Table V).

Biological study MTT assav

The synthesized compounds (3-7) and (9-**16**) were assessed for cytotoxicity against MCF-7 (human, breast tumor), and one normal cell line WRL-68 (human hepatic cell line), by 3 -(4,5dimethyl thiazolyl-2)- 2,5- biphenyl tetrazolium bromide, (MTT) assay. Sorafenib was used as a reference drug. The results were recorded as (IC₅₀) (Figure 5). In order to explore the mechanism of action for compound (15) inhibition proliferation of MCF-7 cells, some conclusive factors, related to apoptosis were assessed by an in vitro Cytotoxic Multi-Parameter Assay against MCF-7 (human,breast cancer) undertaken in High Content Screening (Array Scan XTI, Thermo Scientific). Such factors include the effect of compound (15) on viable cell count, nuclear permeability. intensity, cell membrane mitochondrial membrane potential (MMP), and cytochrome C release, in addition to the five healthy cellular parameters, the effect of compound (15) was tested on caspase 9 activity, compared to a positive control (sorafenib).

The analysis revealed the cytotoxic effect of each of the compounds tested at concentrations [12.5 to 400µg /mL] on the viability of MCF-7 and WRL68 cells, after 24h of treatment, where all of these compounds (3-16) resulted in a substantial reduction in the survival rate of WRL68 and MCF-7 cells in concentration-dependence way, (P<0.0001) (Figure 6 and Table VII). Overall, the results showed that the synthesized compounds substantially improve the cytotoxicity against the breast cancer cell (MCF-7) with acceptable selectivity index, (SI) (Şenkardeş et al., 2020). Compounds (3,6 and7) were more toxic toward normal WRL-68 cells, their selectivity indices (SI) were 0.59, 0.69 and 0.87, respectively, compared to sorafenib with (SI) equal to 2.5. Compounds (11,13,15 and 16) were more selective toward MCF-7, and their (SI) were 2.55,1.99,2.10 and 3.0, respectively ,and possessed the highest potency (lowest IC₅₀ values), compared to their effect on normal cell (WRL-68), with the lowest potency (highest IC₅₀ values); therefore, these compounds are more selective for the type of breast cancer cell (MCF-7) (Table VI).

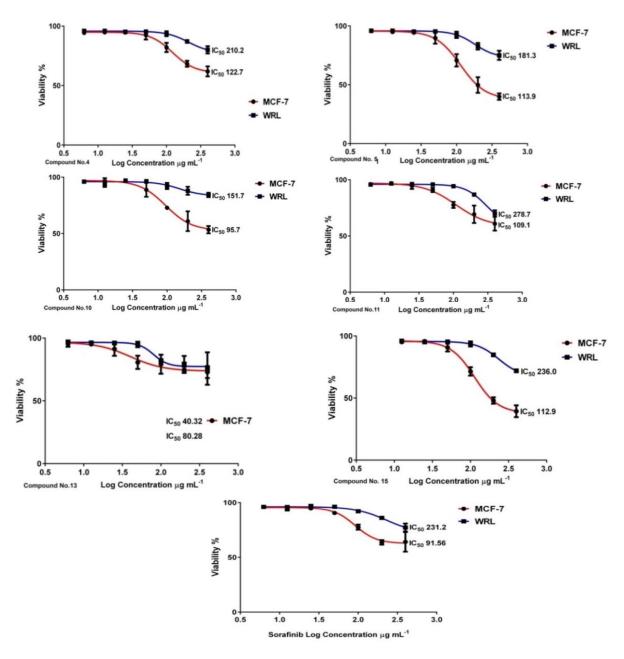


Figure 5. Dose-dependent cytotoxic effect of some synthesized compounds (4,5,10,11,13 and15), and sorafenib on MCF-7 and WRL-68 cells after 24h incubation at 37°C.

Multiple cytotoxic effects of compound (15) on MCF-7 cells *via* array scan high content screening

The cytotoxic effect of compound **(15)** on MCF-7 cell line over 24h was investigated by Array Scan High Content Screening (HCS). Six serial concentrations of compound **(15)** ranged from $(200\mu g/mL)$ to $6.25\mu g/mL$) were prepared and then used to observe the impact on MCF-7

cell viability, nuclear -intensity, membrane -permeability, mitochondrial- membrane potential (MMP), and cytochrome C. The change in such five parameters was shown as mean±SD (Figure 6 and Table VII).

The most major changes in the five parameters were seen at concentrations of $200\mu g/mL$ of compound (15), compared to sorafenib, ($200\mu g/mL$, p < 0.0001).

Table V. Show effect of liver microsomal enzymes on compound (13)

Enzyme	CYT. P450 enzymes effect on compound (13)	Reference scale bar
3A4	S N F	
2C9	NH NH F	0.5

Table VI. The IC₅₀ values determined by MTT assay against MCF-7 and normal WRL-68 cells.

Comm. No.	IC ₅₀ μ	g/mL	*Coloativite: Indox (CI)
Comp. No.	WRL-68	MCF-7	*Selectivity Index (SI)
3	136.5	231.5	0.59
4	210.2	122.7	1.71
5	181.3	113.9	1.59
6	136.5	198.6	0.69
7	175.5	200.8	0.87
9	190.0	152.6	1.25
10	151.7	95.7	1.59
11	278.7	109.1	2.55
12	69.1	67.53	1.02
13	80.28	40.32	1.99
14	185.1	130.8	1.42
15	236	112.29	2.10
16	392	129.9	3.00
Sorafenib	231.2	91.56	2.50

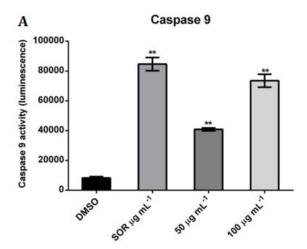
^{*}SI: IC₅₀ on normal WRL-68 /IC₅₀ on cancer MCF-7 cell (Şenkardeş et al.,2020)

Viable cell count

Compound (15) caused a significant reduction in viable cell count only at (200 and $100\mu g/mL$), (p<0.0001 and p=0.0003, respectively), while at concentrations ranging from (6.25-50 $\mu g/mL$), the MCF-7 cell lines were not considerably decreased in count, ($P\ge0.7003$). The reduction trend in cell viability was 27.25% at [$200\mu g/mL$], 21.6% at [$100\mu g/mL$], 9.7% at [$50\mu g/mL$], 2.9% at [$25\mu g/mL$], 3.3% at [$12.5\mu g/mL$], and 1.3% at [$25\mu g/mL$], compared to (sorafenib), 59% at [$200\mu g/mL$], and to the negative control (untreated cell) (Figure 6A and Table VII). Comparative study was previously achieved, following the treatment of sorafenib analogs to MCF7 cells (Wu *et al.*, 2015).

Nuclear intensity

The influence of compound (15) on nuclear intensity (MCF-7), (Figure 6B and Table VII), where this compound has been shown to raise the nuclear intensity of MCF-7, at concentrations of 200μg/mL, 100μg/mL, 50μg/mL, 25μg/mL, $12.5\mu g/mL$, and $6.25\mu g/mL$. The change in the mean nuclear intensity was 833.0±61.54; 676.7±48.99: 509.7±19.04: 464.0±11.14: 504.0±16.7 and 421.3±8.145, respectively, compared to sorafenib, that mostly produced 956.7±39.53 increase in the nuclear intensity, and against a negative control, (untreated cells) of 427.0±44.93.



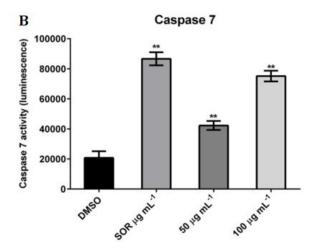


Figure 7 A. The Caspase-9 luminescence of MCF-7 treated with compound (15); B: Caspase-7 activity in MCF-7 cells treated with compound (15) after incubation for 24 h at 37°C.

Table VII. Multiple cytotoxic effects of compound (15) on five cell parameters of MCF-7 cell line via Array Scan high content screening (HCS).

Cytochrome C release	Mitochondrial membrane potential (MMP)	Cell permeability	Nuclear intensity	MCF-7 Cell inhibition %	Concentration (µg/mL)
215.0±22.91	428.3±41.20	79.33±14.05	427.0±44.93	0	Untreated cell
369.7±7.506 **	312.3±8.083 **	90.0±7.81	833.0±61.54 **	27.25 **	200
300.3±17.04 **	348.3±38.66 **	79.33±2.082	676.7±48.99	21.60 **	100
252.7±23.18	419±29.02	61.00±12.53	509.7±19.04	9.70	50
208.7±21.13	437±12.49	72.00±8.544	504.0±16.70	2.90	25
221.7±3.786	436±9.165	80.33±1.155	464.0±11.14	3.30	12.5
213.3±26.58	438±16.7	78.00±4.359	421.3±8.145	1.30	6.25
654.7±50.65 **	106±12.77 **	203.7±30.66 **	956.7±39.53**	59.00 **	Sorafenib

Table VIII. The caspases-9 and-7 activities in MCF-7 cells treated with compound (15) after incubation for 24h at $37^{\circ}C$.

Concentration µg/mL	Caspase 9 (mean± SD), MCF-7 cell line	Caspase 7 (mean ± SD),MCF-7 cell line
50	40855+949.8**	42288+2997***
100	73502+4344**	73169+3590***
Sorafenib	84652+4344**	86652+4356***
DMSO	8317+885	790+4391

^{**}refers to significant result at p < 0.0001; ***refers to significant result at $p \le 0.0004$

The effect of compound (15) on the MCF-7 cell line with a concentration of more than $50\mu g/mL$) was concentration-dependent, while at lower concentrations, (50, 25, 12.5 and $6.25\mu g/mL$), there was no real effect on nuclear intensity, ($p \ge 0.0657$). In the previous study, comparative data were obtained following treatment of MCF7 cells with the anastrozole

(Hassan et al., 2018).

Cell membrane permeability

The compound (15) effect on MCF-7 cells was shown with non-significant effects at all concentrations, therefore, compound (15) did not posses any effect on the cell membrane integrity, compared to sorafenib, as a positive control, that

resulted in a remarkable concentration effect at $(200 \mu g/mL)$, (203.7 ± 30.66) , (p<0.0001) (Figure 6C and Table VII).

Mitochondrial membrane potential (MMP)

The effects of compound (15), on the changes in the permeability of the mitochondrial membrane were evaluated in this experiment. The intensity of MMP was decreased with rising compound's (15) concentration in a concentration-dependent manner. At 200µg/mL there was a 100μg/mL, significant decrease in the mean of the intensity of MMP activity (312.3±8.083, 348.3±38.66) respectively, p<0.0001), compared to sorafenib at $(200 \mu g/mL, p < 0.0001)$, there was a decrease in the mean of the intensity of MMP to 106±12.77. Low concentrations of compound (15), less than <100µg/mL, resulted in a non-significant effect (p=0.9971) (Figure 6D and Table VII).

Cytochrome C release

The results of cytochrome release for compound (**15**)in MCF-7 cell line were a concentration-dependent. There was a marked raise in the cytochrome C release, (p<0.0001), with an increase in the concentrations, compared to a negative control, (215.0±22.91); where, the concentrations of compound (**15**) at 200 μ g/mL, 100 μ g/mL, caused a detectable changes in mean nuclear intensity as follows: (369.7±7.506, 300.3±17.04), respectively, (p<0.0001). While at low concentrations below <100 μ g/mL, there was a non-significant effect (p=0.3433).

Sorafenib also caused changes in mean nuclear intensity, 654.7 ± 50.65 (p<0.0001) (Figure 6 E and Table VII).

Effects of compound (15) on MCF-7 cell caspase 9

The outcomes of caspase-9 activity of MCF-7 treated with compound (**15**) (Figure 7); and there was a substantial increase in caspase 9 activity after 24h, which was (40855 ± 949.8 , 73502 ± 4344), (p<0.0001) in MCF-7 cells treated with $50\mu g/mL$, and $100\mu g/mL$ of compound (**15**), respectively, compared to sorafenib which was set to increase caspase 9 activity to 84652 ± 4344 , (p<0.0001), and in a non-treated cell (DMSO) to 8317 ± 885 , (Figure 7A and Table VIII).

Evaluation of caspase- 7 activity in MCF-7 cell treated with compound (15)

The results of caspase-7 activity showed that there is a significant increase in caspase 7 activity

after 24h, where the activity was, (42288±2997, and 73169±3590), (p<0.0001, and p<0.0004) at MCF-7 cells treated with 50μg/mL, and 100μg/mL of compound (15), respectively, compared to sorafenib that produced raise in caspase 7 activity to 86652±4356,(p<0.0001) ,and in a non-treated cell (DMSO), to 20790±4391 (Figure 7B and Table VIII).

These results showed that MCF-7 cells treated with compound **(15)** undergo apoptosis by disrupting the mitochondrial membrane and release of cytochrome C (Choo *et al.*, 2017). Since these two occurrences are closely correlated to caspase activation. Caspase 9 is a downstream initiate for the activation of another caspases (e.g. 3 and/or 7) leading to apoptosis (Loong *et al.*, 2014).

CONCLUSIONS

New compounds (3-16) were synthesized and approved by spectroscopic analysis including FTIR, ¹HNMR and CHNS. The defined compounds exhibited anticancer activity against the MCF-7 cell line with a maximum effect expressed by compounds 11 and 15 at their (IC_{50}) , 112.9 and 109.1μg/mL, respectively, and with low effect on (WRL-68) cells. Treated MCF-7 tumor cells at different concentrations with such a compound (15) observed statistically measurable reduction in valid cell count, loss of mitochondrial membrane potential (MMP), and a remarkable increase in total nuclear intensity, mitochondrial release of cytochrome C, and increase in caspase 7 and 9 activity. Experiments on molecular docking have shown that target compounds may be considered highly plausible inhibitors of the Vascular Endothelial Growth Factor (VEGFR-2) receptor.

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