Indonesian Journal of Biomedicine and Clinical Sciences

The profile of bacteria isolated from urine culture of adults with urinary tract infection in Yogyakarta 2007-2022

Abu Tholib Aman^{1,2*}, Titik Nuryastuti¹, Alia Hanifa Aman², Vitia Ajeng Nur Linda², Yuli Mawarti^{2,3}

¹Department of Microbiology, Faculty of Medicine, Public Health and Nursing, Universitas Gadjah Mada, ²INA-RESPOND Site 580 Faculty of Medicine, Public Health and Nursing, Universitas Gadjah Mada, ³Laboratory of Microbiology, Sardjito General Hospital, Yogyakarta, Indonesia

https://doi.org/10.22146/inajbcs.v56i3.15890

ABSTRACT

Submitted: 2023-08-28 Accepted : 2023-12-21 Local data regarding antimicrobial susceptibility patterns of bacteria from urine culture is limited in Indonesia, particularly in Yogyakarta. This study was conducted to provide epidemiology data of bacteria and their resistance profile, including the profile of bacteria that producing extended-spectrum beta-lactamase (ESBL) and carbapenemase in the urine of patients with urinary tract infection (UTI) in Yogyakarta. A descriptive retrospective study was conducted by assessing laboratory records of urine culture from adult patients at the Microbiology Laboratory, Department of Microbiology, Faculty of Medicine, Public Health, and Nursing, Universitas Gadjah Mada, Yogyakarta between 2007 and 2022. Of the 842 urine cultures, 464 (55.11%) isolates were recovered. Among these isolates, 50 (10.78%) were fungi, 67 (14.44%) were Gram-positive bacteria, and 347 (74.78%) were Gram-negative bacteria. Enterococcus sp. (41 (61.19%)) was the most bacteria found in the Gram-positive bacteria group, while Escherichia coli (38.90%) were the most bacteria found in the Gram-negative bacteria group. This study also identified Gram-negative bacteria producing ESBL enzymes (58.70%) and carbapenemases (27.94%). Gram-negative bacteria are the most common bacteria found in urine cultures of adult UTI patients in Yogyakarta, and the resistance profile of these bacteria is concerning.

ABSTRAK

Data lokal terkait pola kepekaan kuman terhadap antimikroba pada urin relatif jarang dijumpai di Indonesia, khususnya di Yogyakarta. Penelitian ini bertujuan untuk menyediakan data epidemiologi terkait bakteri dan profil kepekaannya, termasuk profil bakteri penghasil extended-spectrum beta-lactamase (ESBL) dan carbapenemase pada urin pasien dewasa dengan infeksi saluran kemih (ISK) di Yogyakarta. Penelitian retrospektif deskriptif dilakukan dengan menilai catatan laboratorium kultur urin dari pasien dewasa di Laboratorium Mikrobiologi, Fakultas Kedokteran, Kesehatan Masyarakat, dan Keperawatan UGM, Yogyakarta antara tahun 2007 hingga 2022. Dari 842 kultur urin, 464 (55,11%) isolat ditemukan. Diantara isolat tersebut, 50 (10.78%) adalah jamur, 67 (14,44%) bakteri Gram positif, dan 347 (74,78%) bakteri Gram negatif. Enterococcus sp. (41 (61,19%) merupakan bakteri terbanyak yang ditemukan pada kelompok bakteri Gram positif, sedangkan, sedangkan Escherichia coli (38,90%) merupakan bakteri terbanyak yang ditemukan pada kelompok bakteri Gram negatif. Penelitian ini juga mengidentifikasi bakteri Gram negatif penghasil enzim ESBL (58,70%) dan carbapenemase (27,94%). Bakteri Gram negatif adalah bakteri yang paling umum ditemukan pada kultur urin pasien ISK dewasa di Yogyakarta, dan profil resistensi bakteri ini mengkhawatirkan.

Keywords:

bacteria; infection; urine; urinary tract infection; Yogyakarta

INTRODUCTION

Urinary tract infections (UTIs) can cause significant public health problems. particularly severe infections which can be a major economic burden on the healthcare system and deter patients from an optimal guality of life.¹ It affects 150 million people each year worldwide with high recurrence rates and rehospitalization.^{1,2} UTIs are commonly treated with β-lactam antibiotics that raised a global concern for the emergence of antimicrobial resistance including extended-spectrum beta-lactamase (ESBL) and carbapenemase-producing bacteria.^{3,4} These resistance, in addition to the existing patient's comorbidities or risk factors, and the limited choice of effective antibiotics, increase the number of attributable deaths, disabilityadjusted life years, and the economic burden of the infections.^{1,4,5}

Careful assessment of signs, symptoms, and history of disease are important components to diagnose clinical UTI. However urine culture is crucial to identify the cause of infection and establish the diagnosis.6 The urine culture is a part of diagnostic stewardship that can supply information on the causative agents and their susceptibility patterns.⁶ Therefore, it will subsequently facilitate the most suitable antibiotic treatment, reduce cost, and improve patient outcomes.^{6,7} However, urine culture is not always available in Indonesia and many limited resources countries.⁸ It also required several days to produce a result.⁸

Local surveillance for UTIcausing pathogens and antimicrobial susceptibility is necessary in the absence of urine culture.⁹ It will also share benefits in predicting the cause of infection as well as the empiric antibiotic treatment of UTI patients in the same area or similar characteristics of environment.⁹ However, the data was scarcely available or published in limited-resources areas, particularly in Yogyakarta. This study was conducted to supply epidemiology data of bacteria and their resistance profile, including the profile of ESBL- and meropenemaseproducing bacteria from the UTI patients in Yogyakarta.

MATERIAL AND METHODS

Design of study

A descriptive retrospective study was conducted by collecting data of urine culture from a register of laboratory (secondary records data) at the Microbiology Laboratory, Department of Microbiology, Faculty of Medicine, Public Health, and Nursing, Universitas Gadjah Mada, Yogyakarta during the period of 2007-2022. API (BioMerieux) was used for microorganism identification, then the susceptibility test was conducted with a disk diffusion method to evaluate their sensitivity against antibiotics on Mueller-Hinton agar. Result interpretation was performed following the Performance Standards for Antimicrobial Susceptibility Testing of Clinical and Laboratory Standards Institute (CLSI) M100.

The CLSI definition was used in this study to screen ESBL producer which can be identified from their phenotypic resistance against extended penicillin, monobactam (aztreonam), 3rd generation cephalosporinswithorwithoutresistance against 4th generation cephalosporins.¹⁰ Carbapenemase producer was identified from their resistant trait against one or more carbapenems (i.e. meropenem, imipenem, ertapenem, or doripenem) phenotypically.¹⁰ Meropenem is used to screen carbapenemase producers as it offers the best sensitivity and specificity features compared to other carbapenems.11

Data analysis

Data were collected using an anonymous data sheet to keep patient confidentiality and analyzed using STATA 17 ME. The species or genus of microbes that were recovered from the urine culture were tabulated, also their susceptibility profile against the antibiotics tested. Summary statistics was conducted for descriptive study using the command "tab" to obtain frequency distribution tables, crosstabulation, or two-way tables. The positive results were presented in the frequency distribution table [n (%)] with the detail of calculation in the footnote of TABLE 2 and 3. A total of 842 urine cultures from adult (18 y.o. or older) patients were documented during the study period. Before the study initiation, ethical approval was obtained from the Institutional Review Board (The Medical and Health Research Ethics Committee (MHREC)) of Faculty of Medicine, Public Health, and Nursing, Universitas Gadjah Mada, Yogyakarta (Reference number: KE/FK/0052/EC/2023).

RESULTS

Of the 842 urine cultures, 464 (55.11%) isolates were recovered during examination. Among those isolates, we identified 50 (10.78%) fungal isolates, 67 (14.44%) Gram-positive isolates, and 347 (74.78%) Gram-negative isolates.

Gram-positive isolates recovered from the urine culture

Among Gram-positive bacteria isolates, the most frequent bacteria identified in UTIs were *Enterococcus* sp. [41 (61.19%)], followed by *Staphylococcus aureus* [12 (17.91%)], *Enterococcus faecalis* [11 (16.42%)], *Streptococcus agalactiae* [2 (2.99%)], and *S. pneumoniae* [1 (1.49%)] (TABLE 1).

ESBL-producing and Gram-negative isolates recovered from the urine culture

This research elucidated that the most common Gram-negative bacteria identified from urine patients with UTIs were E. coli, accounted for 38.90% of total Gram-negative isolates (TABLE 2). The other Gram-negative bacteria recovered from those cultures were Pseudomonas sp. [60 (17.29%)], P. aeruginosa [42 (12.10%)], Klebsiella pneumoniae [38 (10.95%)], Enterobacter sp. [12 (3.46%)], Proteus mirabilis [10 (2.88%)], Klebsiella sp. [8 (2.31%)], K. aerogenes (also known as *E. aerogenes*) [4 (1.15%)], *P.* fluorescens [4 (1.15%)], Acinetobacter baumannii [3 (0.86%)], Citrobacter sp. [3 (0.86%)], Providencia rettgeri [3 (0.86%)], Proteus penneri (2 (0.58%)), Proteus sp. [2 (0.58%)], P. putida [2 (0.58%)], Serratia sp. [2 (0.58%)], and less common Gramnegative bacteria [17 (4.90%)].

TABLE 1. Gram-positive bacteria isolated from urine samplesof UTI patients in Yogyakarta 2007-2022

n (%)
41 (61.19)
12 (17.91)
11 (16.42)
2 (2.99)
1 (1.49)
67 (100.00)

Bacteria with "sp." means that the isolate identification was only up to genus level

TABLE 2. Gram-negative bacteria which isolated from urine samples of
UTI patients and ESBL-producing pathogens in Yogyakarta 2007-
2022

Name of bacteria/ spesies	Isolates recovered from urine samples [n (%)]*	Isolates tested for ESBL [n (%)]**	Isolates with ESBL phenotypes [n(%)]***
E. coli	135 (38.9)	135(100)	57 (42.22)
Pseudomonas sp.	60 (17.29)	55 (91.67)	49 (89.09)
P. aeruginosa	42 (12.1)	41 (97.62)	35 (85.37)
K. pneumoniae	38 (10.95)	38 (100)	20 (52.63)
Enterobacter sp.	12 (3.45)	12 (100)	7 (58.33)
P. mirabilis	10 (2.88)	9 (90)	1 (11.11)
Klebsiella sp.	8 (2.30)	8 (100)	5 (62.5)
K. aerogenes	4 (1.15)	4 (100)	4 (100)
P. fluorescens	4 (1.15)	4 (100)	3 (75)
A. baumannii	3 (0.86)	3 (100)	1 (33.33)
Citrobacter sp.	3 (0.86)	3 (100)	2 (66.67)
P. rettgeri	3 (0.86)	3 (100)	1 (33.33)
P. penneri	2 (0.58)	2 (100)	1 (50)
Proteus sp.	2 (0.58)	2 (100)	1 (50)
P. putida	2 (0.58)	2 (100)	1 (50)
Serratia sp.	2 (0.58)	2 (100)	1 (50)
S. maltophilia	1 (0.29)	1 (100)	0 (0)
Escherichia sp.	1 (0.29)	1 (100)	0 (0)
E. fergusonii	1 (0.29)	1 (100)	1 (100)
E. cloacae	1 (0.29)	1 (100)	1 (100)
A. caviae	1 (0.29)	1 (100)	1 (100)
B. pseudomallei	1 (0.29)	1 (100)	1 (100)
C. youngae	1 (0.29)	1 (100)	0 (0)
<i>Kluyvera</i> sp.	1 (0.29)	1 (100)	0 (0)
Leclercia sp.	1 (0.29)	1 (100)	0 (0)
P. alcaligenes	1 (0.29)	1 (100)	1 (100)
P. alcalifaciens	1 (0.29)	1 (100)	0 (0)
S. marcescens	1 (0.29)	1 (100)	1 (100)
S. odorifera	1 (0.29)	N/D	N/D
S. liquefaciens	1 (0.29)	1 (100)	1 (100)
Yersinia rohdei	1 (0.29)	1 (100)	1 (100)
Edwardsiella tarda	1 (0.29)	1 (100)	1 (100)
Edwardsiella sp.	1 (0.29)	1 (100)	1 (100)
Total	347 (100)	339 (97.69)	199 (58.70)

N/D: no data; (%)* is the number of Gram-negative isolates divided by total of Gramnegative isolates recovered from the urine culture; (%)** is the number of Gram-negative isolates which were tested for ESBL phenotypes divided by total of respective Gramnegative species or isolates recovered from the urine culture; (%)***) is the number of Gram-negative isolates with ESBL phenotypes divided by total of respective Gramnegative isolates or species which were tested for ESBL phenotypes; Bacteria with "sp." means that the isolate identification was only up to genus level

Name of bacteria/ Species	Isolates tested for carbapenem- resistant phenotype [n (%)] [¢]	Isolates with carbapenem- resistant phenotype [n (%)]**
Pseudomonas sp.	60 (100)	27 (45)
E. coli	135 (100)	23 (17.04)
K. pneumoniae	38 (100)	10 (26.32)
P. aeruginosa	40 (95.24)	10 (25)
Enterobacter sp.	12 (100)	5 (41.67)
Klebsiella sp.	8 (100)	4 (50)
P. fluorescens	4 (100)	3 (75)
Serratia sp.	2 (100)	2 (100)
Proteus sp.	2 (100)	1 (50)
Citrobacter sp.	2 (66.67)	1 (50)
P. rettgeri	3 (100)	1 (33.33)
A. caviae	1 (100)	1 (100)
B. pseudomallei	1 (100)	1 (100)
Edwardsiella sp.	1 (100)	1 (100)
E. tarda	1 (100)	1 (100)
Leclercia sp.	1 (100)	1 (100)
S. liquefaciens	1 (100)	1 (100)
Y. rohdei	1 (100)	1 (100)
P. mirabilis	10 (100)	1 (10)
A. baumannii	3 (100)	0 (0)
C. youngae	1 (100)	0 (0)
E. cloacae	1 (100)	0 (0)
K. aerogenes	4 (100)	0 (0)
E. fergusonii	1 (100)	0 (0)
Escherichia sp.	1 (100)	0 (0)
<i>Kluyvera</i> sp.	1 (100)	0 (0)
P. penneri	1 (50)	0 (0)
P. alcalifaciens	1 (100)	0 (0)
P. alcaligenes	1 (100)	0 (0)
P. putida	2 (100)	0 (0)
S. marcescens	1 (100)	0 (0)
S. odorifera	1 (100)	0 (0)
Total	342	95 (27.94)

TABLE 3. Gram-negative carbapenem-resistant bacteria from urine samples of UTI patients in Yogyakarta 2007-2022

(%)⁺ is the number of gram-negative isolates or species which were tested for carbapenemresistant phenotypes divided by total of respective gram-negative isolates which recovered from the urine culture; (%)⁺⁺ is the number of gram-negative isolates with carbapene- resistant phenotypes divided by total of respective gram-negative isolates which were tested for carbapenem-resistant phenotypes; Bacteria with "sp." means that the isolate identification was only up to genus level

During the study period, a total of 199 (58.70%) ESBL-producing gramnegative bacteria were identified from those isolates. We identified that 57 (42.22%) E. coli phenotypically exhibited the characteristic of ESBL-producing bacteria. Other gram-negative bacteria were also often phenotypically presented ESBL-producing bacteria; as those including Pseudomonas sp. (49 (89.09%)), *P. aeruginosa* (35 (85.37%)), and *K.* pneumoniae (20 (52.63%)). Enterobacter sp., Klebsiella sp., K. aerogenes, and P. fluorescens were less identified as ESBLproducing bacteria accounted for 7 (58.33%), 5 (62.5%), 4 (100%), and 3 (75%) isolates respectively. Other bacteria were also shown as ESBL producers, but they are limited in number (TABLE 2).

Carbapenem-resistant Gram-negative bacteria identified from the urine culture

This study recorded that among Gram-negative bacteria which isolated tested against carbapenem and (meropenem), resistance was identified in 27.94% of them (TABLE 3). Although some bacteria isolates revealed a relatively high percentage of carbapenem resistance, this research sample size is limited. Among Gram-negative bacteria isolates tested against carbapenem, Pseudomonas sp., E. coli, K. pneumoniae, P. aeruginosa, Enterobacter sp., Klebsiella sp., P. fluorescens, and Serratia sp. were relatively common as carbapenemase producers which accounted for 27 (45%), 23 (17.04%), 10 (26.32%), 10 (24.39%), 5 (41.67%), 4 (50%), 3 (75%), 2 (100%) isolates, respectively.

DISCUSSION

This study described the UTI's etiology and their resistance against antibiotics by observing their phenotypic characteristics as ESBL-producing and or carbapenemase-producing bacteria. This data can serve as local surveillance in Yogyakarta that enables benefits in diagnostic and antimicrobial stewardship programs.¹² This study showed of all recovered isolates from urine culture, Gram-negative bacteria were identified as the predominant (74.78%)uropathogens, whereas Gram-positive bacteria and fungi were represented only in 14.44% and 10.78%, respectively. This finding was similar to a study from East China (2021) that conducted urine culture from 1760 UTIs patients, and reported uropathogens which consisted of 90.5% Gram-negative bacteria, 9.3% Gram-positive bacteria, and 0.2% fungi.¹³ In Indonesia, a study of asymptomatic UTI in pregnant women revealed that Gram-negative bacteria (72%) were more frequently isolated from their urine culture, as compared to Gram-positive bacteria (28%).¹⁴ A study in Surabaya, Indonesia also reported similar result, Gram-negative (59.67%) and Gram-positive (14.51%) bacteria, as well as fungi (Candida sp.) (25.81%) were identified in the urine culture of diabetic patients with UTI.15

Gram - positive bacteria as uropathogens

The East China study reported E. faecalis as the most prevalent (31.7%) gram-positive bacteria, then followed by S. agalactiae (24.4%), S. saprophyticus (18.3%), E. faecium (9.1%) and others (16.5%).¹³ Our study has slightly different pattern, we reported *Enterococcus* sp., S. aureus, and E. faecalis as the majority (95.52% in total) of Gram-positive bacteria identified from urine culture, whereas S. agalactiae was only accounted for 2.99% of Gram-positive bacteria involved in UTIs. A study in Jakarta revealed that S. agalactiae (33.33%), E. faecalis (19.04%), and S. saprophyticus (14.28%) were the frequently identified gram-positive bacteria of asymptomatic UTI in pregnant women.¹⁴ In Surabaya, *E*.

faecalis (66.66%) was the gram-positive bacteria which frequently isolated from urine culture of diabetic patients with UTI.¹⁵

This study found that S. pneumoniae, which is commonly associated with respiratory or central nervous infection. was detected in a urine sample from a patient with UTI.¹⁶ This extraordinary finding came from a patient who also suffered from S. pneumoniae bacteraemia, as suggested by the positive results of two blood cultures (also yielded S. pneumoniae) which accompanied the urine culture. Although uncommon, in Munich, Germany, S. pneumoniae as urinary tract pathogen was reported in a 82-year-old man with pyelonephritis urosepsis.¹⁶ Pneumococcosuria and where S. pneumoniae identified as an agent of infection in urinary tract was scarce.¹⁷ A study at the Department for Infectious Diseases, University Hospital of Heidelberg, reported that 26 urine samples from 18 different patients (age of 3-72 years) contained S. pneumoniae between January 2010 and December 2014.¹⁷ The literature suggested that in children, S. pneumoniae is rarely identified from urine samples (less than 1%).18

Gram - negative bacteria as uropathogens

Escherichia coli was accounted for 75-95% of uropathogens in UTIs all over the world.¹⁹ A study in Japan which included a total of 2049 UTI patients reported that 1682 (82.1%) of UTIs were caused by gram-negative bacteria. It comprised *E. coli* (93.3%), *Klebsiella* sp. (6.2%), and *P. mirabilis* (0.5%).²⁰ Interestingly, a metaanalysis study in Iran found comparable finding among pregnant women, that *E. coli* and *Klebsiella* were the common gram-negative bacteria causing UTIs which accounted for 61.6% and 13.9% respectively.²¹ Similarly, the study in US Veterans Affairs medical centers (in Minnesota and Texas) highlighted that *E. coli* was a predominant uropathogen which accounted for 40.7%.²²

A study of asymptomatic UTI in pregnant women in Indonesia reported that the gram-negative bacteria which frequently isolated in the urine culture were E. coli (37.04%) and K. pneumoniae (27.78%).¹⁴ In Surabaya, a study of UTI in diabetic patients reported that the gramnegative bacteria which frequently isolated in the urine culture were E. coli (54.05%), A. baumannii (10.81%), and Enterobacter spp. (8.10%).¹⁵ Similarly, our study reported that *E. coli* was the most prevalent (38.9%) gramnegative bacteria recovered from UTIs, however the second and third most frequent gram-negative uropathogens were Pseudomonas sp. (17.29%) and P. aeruginosa (12.10%) respectively. We also found a various species of other gram-negative bacteria which involved in UTIs comprising K. pneumoniae (10.95%), Enterobacter sp. (3.45%), P. *mirabilis* (2.88%), and others (14.43%).

ESBL producing Gram - negative bacteria

Our study highlighted that gramnegative bacteria were the major pathogen (74.78%) causing UTIs, and around 59% of those isolates shared similar phenotypic trait as ESBL producer. ESBLs are defined as a rapidly evolving group of enzymes produced by certain bacteria that can hydrolize extended spectrum cephalosporin.^{23,24} These enzymes found to be effective against one or more of third and fourth generation of cephems (such as ceftazidime, ceftriaxone, cefotaxime, cefepime), extended spectrum penicillin (i.e. piperacillin), and monobactam (i.e. aztreonam) but are inhibited by tazobactam.²³⁻²⁵ clavulanic acid or Therefore the presence of ESBLs in gram-negative bacteria warrant special attention due to the associated risks of antibiotic therapy failure.²⁶

ESBL-producing *Enterobacteriaceae* (i.e. E. coli, K. pneumoniae, Enterobacter sp., *Proteus* sp.) are the most prevalent causative agents of UTIs.²⁵ They can be a major threat to the global public as their resistance against *B*-lactam antibiotics leads to treatment failure in many infections.²⁵ While our study reported 42.22% ESBL-producing E. coli in the urine of UTI patients, a study in Jordan (2019) reported a higher proportion (62%).²⁷ A systematic review in Ethiopia reported a high rate of ESBL-producing gam-negative bacteria among clinical samples with a pooled rate of 50.1% among different species and varied in the groups of *Klebsiella* spp. (65.7%), Enterobacter spp. (62.2%), Salmonella spp. (48.4%), E. coli (47.0%), Citrobacter spp. 46.8%, Providencia spp. (43.8%), Proteus spp. (28.3%), P. aeruginosa (17.4%), Acinetobacter spp. (9.4%), and other Gram-negative bacteria (20.8%).²⁸

A five-year global surveillance "SMART program" (2015 - 2019)by reported that the prevalence of noncarbapenem-resistant ESBL-producing Enterobacteriaceae was 30% globally, and exceeded 50% in India, Thailand, Vietnam, China, Russia, Mexico, Kenya, and Kuwait.²⁵ The SMART program estimated that the prevalence of noncarbapenem-resistant ESBL-producing K. pneumoniae was 25.4% globally, and more than 40% in Portugal, Chile, Ecuador, Guatemala, Mexico, Israel, Morocco, Lithuania, Kenya, and Kuwait.²⁵ They also reported that the prevalence of non-carbapenem-resistant ESBL-producing *E. coli* increased significantly (p < 0.05) in Asia (excluding China), Australia, New Zealand, and Latin America.²⁵ Furthermore, the noncarbapenem-resistant ESBL-producing *K. pneumoniae* prevalence increased significantly (p< 0.05) in Latin America, USA, and Canada.²⁵ Unfortunately, no data from Indonesia was included in the SMART surveillance.²⁵

A retrospective study in Bali. Indonesia (2019-2020) revealed that ESBL-producing E. coli (56.32%) and K. pneumoniae (54%) were identified in the urine culture of patients with UTI and chronic kidney disease.²⁹ Another study in Medan reported that ESBL-producing *E. coli* was contributed to 8.4% of urine associated catheter infections in adult patients who admitted into the intensive care unit from July to August 2018.³⁰ Differences in clinical settings, study period, study population, and methods might modify the result, thus explaining the heterogeneity of the ESBL-producing bacteria prevalence worldwide.²⁸

ESBLs are enzymes encoded in plasmids and can be easily transferred to other bacteria.³¹ Apart from ESBLproducing Klebsiella and E. coli, our study also reported a relatively high prevalence of ESBL-producing Grambacteria, including negative nonlactose-fermenting bacteria such as P. aeruginosa, Pseudomonas spp., and a wide variety of other Gram-negative bacteria causing UTIs.³² These bacteria have also been reported as the causative pathogen in UTIs, especially in healthcare associated UTIs.³²

Carbapenem - resistant in Gram - negative bacteria

Carbapenems have а β-lactam ring that differs from penicillins by replacing the sulphur atom at C-1 with a carbon atom and adding a double bond between C-2 and C-3.³³ In addition to this characteristic, the side chain of carbapenemisinthetranspositioninstead of the cis- position, making this drug insensitive to the effect of β-lactamases.³³ Carbapenems were previously effective treatening-drug resistant (MDR) in bacteria, including ESBL-producing bacteria which resistant to penicillin cephalosporins.³⁴ Unfortunately, and the acquisition of carbapenemase genes causes these bacteria to be able to hydrolize carbapenem, leaving a limited choice of antibiotic treatment for MDR bacteria.³⁵

Carbapenemase emergence and spread have increased dramatically over the last decade following its discovery in K. pneumoniae in the US in 1996.^{34,36} The increasing prevalence of carbapenemresistant gram-negative bacteria as the cause of infections is a global threat, hence the WHO highlighted those pathogens, particularly carbapenem-Enterobacterales resistant (CRE), carbapenem-resistant *P*. aeruginosa (CRPA) and carbapenem-resistant A. baumanii (CRAB) in the global priority list of pathogens in 2017.³⁷ Carbapenemproducing bacteria surveillance is an important strategy to control its spread and enable a positive impact for public health.38

Carbapenem-resistant bacteria in this study were reported in 95 (27.94%) of gram-negative bacteria isolated from urine cultures of UTI samples. Our study identified various percentage of carbapenem resistance among different bacterial species isolated from UTI patients, including Klebsiella sp. (50%), K. pneumoniae (26.32%), E. coli (17.04%), Pseudomonas sp. (45%), Enterobacter sp. (41.67%), *P. fluorescens* (75%), and many more as described in TABLE 3. A review study comprising data mostly from Asia, North America, and Europe (2017) reported that proportion of carbapenem-Enterobacteriaceae in resistant community setting was ranged from 0 to 29.5% with the highest proportion of carbapenem- resistant Enterobacteriaceae was identified in Asia.³⁹ In Taiwan, the prevalence of carbapenem-resistant *P*. aeruginosa was increasing from 12% in 2012-2015 to 19-23% in 2018–2021.40 Surveillance of carbapenem resistance, especially in patients with UTI is remain scattered and fragmented, moreover various surveillance models were adapted to fit with the requirements and capacities of each setting or country.⁴¹ Furthermore, developing countries struggle with political and social dilemmas due to weak laboratory capacity, poor health systems governance, lack of health information systems, and limited resources.^{41,42} Despite the challenges, a local antibiogram or data regarding uropathogen and its susceptibility pattern is important to inform local treatment guidelines and promote antimicrobial stewardship program.43

Study limitation

This study has limitations, including the small sample size of each bacteria species, limited clinical information, and the passive nature of a secondary laboratory records.

CONCLUSION

This study highlighted the profile of bacteria associated with UTIs in Yogyakarta during 2007-2022 that could be useful for antimicrobial stewardship. The recovery proportion of pathogen of urine culture from UTIs patients was relatively high (55.11%), and like other studies in the world, gram-negative bacteria was the most prevalent pathogen isolated from urine of UTI cases. Escheriachia coli and bacteria in the genus Pseudomonas and Klebsiella were the most frequently identified in this population. In addition, this study revealed the high proportion of ESBLsand carbapenemase-producer among gram-negative bacteria associated with UTIs. Future study is required to refine the epidemiology of bacteria that cause UTI in developing countries or limitedresource region, especially in Yogyakarta.

ACKNOWLEDGMENTS

We would like to thank INA-RESPOND Site 580, Ms Mulyani, and everyone who gave support to our effort to write down this article. This study is partially funded by INA-RESPOND Site 580.

REFERENCES

- 1. Babich T, Eliakim-Raz N, Turjeman A, Pujol M, Carratala J, Shaw E, *et al.* Risk factors for hospital readmission following complicated urinary tract infection. Sci Rep 2021; 11(1):6926. https://doi.org/10.1038/s41598-021-86246-7
- 2. Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. Nat Rev Microbiol 2015; 13(5):269-84.

https://doi.org/10.1038/nrmicro3432

3. Zalmanovich A, Katzir M, Chowers M, Matar A, Rodrig J, Alon D. Improving urinary tract infection treatment through a multifaceted antimicrobial stewardship intervention in the emergency department. Am J Emerg Med 2021; 49:10-3.

https://doi.org/10.1016/j.ajem.2021.05.037

4. Zhang H, Liang B, Wang J, Cai Y. Non-carbapenem beta-lactam/ beta-lactamase inhibitors versus carbapenems for urinary tract infections caused by extendedspectrum beta-lactamase-producing Enterobacteriaceae: a systematic review. Int J Antimicrob Agents 2021; 58(4):106410.

h t t p s : // d o i . o r g / 10.1016/j. ijantimicag.2021.106410

5. Kang SW, Park S, Kim A, Han J, Lee J, Seo H, *et al.* Clinical characteristics of and risk factors for subsequent c a r b a p e n e m a s e - p r o d u c i n g enterobacterales (CPE) bacteremia in rectal CPE carriers. Int J Antimicrob Agents 2023; 62(5):106959.

h t t p s : // d o i . o r g / 1 0 . 1 0 1 6 / j . ijantimicag.2023.106959

6. Goebel MC, Trautner BW, Grigoryan L. The five Ds of outpatient antibiotic stewardship for urinary tract infections. Clin Microbiol Rev 2021; 34(4):e0000320.

https://doi.org/10.1128/CMR.00003-20

7. Lee ALH, Leung ECM, Lee MKP, Lai RWM. Diagnostic stewardship programme for urine culture: impact on antimicrobial prescription in a multi-centre cohort. J Hosp Infect 2021; 108:81-9.

https://doi.org/10.1016/j.jhin.2020.10.027

- Ginting F, Sugianli AK, Kusumawati RL, Parwati I, de Jong MD, Schultsz C, *et al.* Predictive value of the urinary dipstick test in the management of patients with urinary tract infectionassociated symptoms in primary care in Indonesia: a cross-sectional study. BMJ Open 2018; 8(8):e023051. https://10.1136/bmjopen-2018-023051
- 9. Bazaid AS, Saeed A, Alrashidi A, Alrashidi A, Alshaghdali K, Hammam SA, *et al.* Antimicrobial surveillance for bacterial uropathogens in Ha'il, Saudi Arabia: A five-year multicenter retrospective study. Infect Drug Resist 2021; 14:1455-65.

https://doi.org/10.2147/IDR.S299846

- Lewis JS, Weinstein MP, Bobenchik AM, Campeau S, Cullen SK, Dingle T, et al. CLSI performance standards for antimicrobial susceptibility testing M100. USA: The Clinical and Laboratory Standards Institute 2023. 30th ed.
- 11. (EUCAST) ECoAST. EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance. EUCAST. 2017; 1-43.
- 12. Sugianli AK, Ginting F, Kusumawati RL, Parwati I, de Jong MD, van Leth F, *et al.* Laboratory-based versus population-based surveillance of antimicrobial resistance to inform empirical treatment for suspected urinary tract infection in Indonesia. PLoS One 2020; 15(3):e0230489. https://doi.org/10.1371/journal. pone.0230489
- 13. Quan J, Dai H, Liao W, Zhao D, Shi Q,

Zhang L, *et al.* Etiology and prevalence of ESBLs in adult community-onset urinary tract infections in East China: A prospective multicenter study. J Infect 2021; 83(2):175-81. https://doi.org/10.1016/j.jinf.2021.06.004

- 14. Rosana Y, Ocviyanti D, Halim M, Harlinda FY, Amran R, Akbar W, et al. Urinary tract infections among Indonesian pregnant women and its susceptibility pattern. Infect Dis Obstet Gynecol 2020; 2020:9681632. https://doi.org/10.1155/2020/9681632
- 15. Norafika, Arbianti N, Prihatiningsih S, Indriani DW, Indriati DW. A retrospective cross-sectional study of urinary tract infections and prevalence of antibiotic resistant pathogens in patients with diabetes mellitus from a public hospital in Surabaya, Indonesia. Germs 2020; 10(4):157-66.

https://doi.org/10.18683/germs.2020.1201

 Dufke S, Kunze-Kronawitter H, Schubert S. Pyelonephritis and urosepsis caused by Streptococcus pneumoniae. J Clin Microbiol 2004; 42(9):4383-5.

h t t p s : // d o i . o r g / 1 0 . 1 1 2 8 / JCM.42.9.4383-4385.2004

17. Burckhardt I, Panitz J, van der Linden M, Zimmermann S. *Streptococcus pneumoniae* as an agent of urinary tract infections - a laboratory experience from 2010 to 2014 and further characterization of strains. Diagn Microbiol Infect Dis 2016; 86(1):97-101.

https://doi.org/10.1016/j. diagmicrobio.2016.06.009

- 18. Pougnet R, Sapin J, De Parscau L, Pougnet L. *Streptococcus pneumoniae* urinary tract infection in pedeatrics. Ann Biol Clin (Paris) 2017; 75(3):348-50. https://doi.org/10.1684/abc.2017.1241
- 19. Kubone PZ, Mlisana KP, Govinden U, Abia ALK, Essack SY. Antibiotic susceptibility and molecular characterization of uropathogenic *Escherichia coli* associated with

community-acquired urinary tract infections in urban and rural settings in South Africa. Trop Med Infect Dis 2020; 5(4):176.

h t t p s : / / d o i . o r g / 1 0 . 3 3 9 0 / tropicalmed5040176

20. Ohnishi T, Mishima Y, Naito T, Matsuda N, Ariji S, Umino D, et al. Clinical features and treatment strategies of febrile urinary tract infection caused by extendedspectrum beta-lactamase-producing Enterobacteriaceae in children: multicenter retrospective А observational study in Japan. Int J Infect Dis 2022; 125:97-102.

https://doi.org/10.1016/j.ijid.2022.09.033

21. Azami M, Jaafari Z, Masoumi M, Shohani M, Badfar G, Mahmudi L, *et al.* The etiology and prevalence of urinary tract infection and asymptomatic bacteriuria in pregnant women in Iran: a systematic review and meta-analysis. BMC Urol 2019; 19(1):43.

https://doi.org/10.1186/s12894-019-0454-8

- 22. Drekonja DM, Trautner B, Amundson C, Kuskowski M, Johnson JR. Effect of 7 vs 14 days of antibiotic therapy on resolution of symptoms among afebrile men with urinary tract infection: A randomized clinical trial. JAMA 2021; 326(4):324-31. https://doi.org/10.1001/jama.2021.9899
- 23. Rawat D, Nair D. Extended-spectrum beta-lactamases in Gram-negative bacteria. J Glob Infect Dis 2010; 2(3):263-74.

https://10.4103/0974-777X.68531

- 24. Ghafourian S, Sadeghifard N, Soheili S, Sekawi Z. Extended spectrum betalactamases: definition, classification and epidemiology. Curr Issues Mol Biol 2015; 17:11-21.
- 25. Karlowsky JA, Lob SH, DeRyke CA, Siddiqui F, Young K, Motyl MR, et al. Prevalence of ESBL non-CRE Escherichia coli and Klebsiella pneumoniae among clinical isolates collected by the SMART global

surveillance programme from 2015 to 2019. Int J Antimicrob Agents 2022; 59(3):106535.

https://doi.org/10.1016/j. ijantimicag.2022.106535

26. Apostolakos I, Mughini-Gras L, Fasolato L, Piccirillo A. Impact of selective and non-selective media on prevalence and genetic makeup of ESBL/pAmpC-producing Escherichia *coli* in the broiler production pyramid. Vet Microbiol 2020: 240:108536.

https://doi.org/10.1016/j. vetmic.2019.108536

- 27. Al-Jamei SA, Albsoul AY, Bakri FG, Al-Bakri AG. Extended-spectrum betalactamase producing Escherichia coli in urinary tract infections: A two-center, cross-sectional study of prevalence, genotypes and risk factors in Amman, Jordan. J Infect Public Health 2019; 12(1):21-5. https://doi.org/10.1016/j.jiph.2018.07.011
- 28. Tufa TB, Fuchs A, Tufa TB, Stotter L, Kaasch AJ, Feldt T, et al. High rate of extended-spectrum beta-lactamaseproducing Gram-negative infections and associated mortality in Ethiopia: a systematic review and metaanalysis. Antimicrob Resist Infect Control 2020; 9(1):128. https://doi.org/10.1186/s13756-020-00782-x
- 29. Wijaya C, Eriata AH, Rustawan INT. Candra IKBA, Budayanti NNS. Prevalence of uropathogen producing extended spectrum beta lactamase (ESBL) at urinary tract infection in chronic kidney disease patient. J Clin Microbiol Infect Dis (JCMID) 2023; 3(1):12-5.

https://doi.org/10.51559/jcmid.v3i1.29

30. Anggi A, Wijaya DW, Ramayani OR. Risk Factors for Catheterassociated urinary tract infection and uropathogen bacterial profile in the Intensive Care Unit in Hospitals in Medan, Indonesia. Open Access Maced J Med Sci 2019; 7(20):3488-92.

https://doi.org/10.3889/oamjms.2019.684

- 31. Hosu MC, Vasaikar SD, Okuthe GE, Apalata T. Detection of extended spectrum beta-lactamase genes in Pseudomonas aeruginosa isolated from patients in rural Eastern Cape Province, South Africa. Sci Rep 2021; 11(1):7110. https://doi.org/10.1038/s41598-021-86570-y
- 32. Wagenlehner FME, Bjerklund Johansen TE, Cai T, Koves B, Kranz J, Pilatz A, et al. Epidemiology, definition and treatment of complicated urinary tract infections. Nat Rev Urol 2020; 17(10):586-600. https://doi.org/10.1038/s41585-020-0362-4
- 33. Aurilio C, Sansone P, Barbarisi M, Pota V, Giaccari LG, Coppolino F, et al. Mechanisms of action of carbapenem resistance. Antibiotics (Basel) 2022; 11(3):421. https://doi.org/10.3390/ antibiotics11030421
- 34. Hansen GT. Continuous evolution: perspective on the epidemiology of carbapenemase resistance among enterobacterales and other Gramnegative bacteria. Infect Dis Ther 2021; 10(1):75-92. https://doi.org/10.1007/s40121-020-00395-2
- 35. Chen Y, Marimuthu K, Teo J, Venkatachalam I, Cherng BPZ, De Wang L, et al. Acquisition of plasmid with carbapenem-resistance gene bla(KPC2) in hypervirulent Klebsiella pneumoniae, Singapore. Emerg Infect Dis 2020; 26(3):549-59. https://doi.org/10.3201/eid2603.191230
- 36. Chen L, Mathema B, Chavda KD, DeLeo FR, Bonomo RA, Kreiswirth Carbapenemase-producing BN. Klebsiella pneumoniae: molecular genetic decoding. and Trends Microbiol 2014; 22(12):686-96.

https://doi.org/10.1016/j.tim.2014.09.003

37. Nordmann P, Poirel L. Epidemiology and diagnostics of carbapenem resistance in Gram-negative bacteria. Clin Infect Dis 2019; 69(Suppl 7):S521-S8. https://doi.org/10.1093/cid/ciz824

38. Gomides MDA. Fontes AMS. Silveira A, Matoso DC, Ferreira AL, Sadoyama G. The importance of active surveillance of carbapenemresistant Enterobacterales (CRE) in colonization rates in critically PLoS ill patients. One 2022; 17(1):e0262554.

https://doi.org/10.1371/journal. pone.0262554

- 39. Kelly AM, Mathema B, Larson EL. Carbapenem-resistant enterobacteriaceae in the community: A scoping review. Int J Antimicrob Agents 2017; 50(2):127-34. h t t p s : // d o i . o r g / 1 0 . 1 0 1 6 / j . ijantimicag.2017.03.012
- 40. Karlowsky JA, Wise MG, Hsieh TC, Lu HC, Chen WT, Cheng MH, *et al.* Temporal and geographical prevalence of carbapenem-resistant *Pseudomonas aeruginosa* and the *in vitro* activity of ceftolozane/ tazobactam and comparators in Taiwan-SMART 2012-2021. J Glob

Antimicrob Resist 2023; 34:106-12. https://doi.org/10.1016/j.jgar.2023.06.013

- 41. Perez F, Villegas MV. The role of surveillance systems in confronting the global crisis of antibioticresistant bacteria. Curr Opin Infect Dis 2015; 28(4):375-83. https://doi.org/10.1097/ QCO.00000000000182
- 42. Iskandar K, Molinier L, Hallit S, Sartelli M, Hardcastle TC, Haque M, *et al.* Surveillance of antimicrobial resistance in low- and middleincome countries: A scattered picture. Antimicrob Resist Infect Control 2021; 10(1):63. https://doi.org/10.1186/s13756-021-

00931-w

43. Khatri D, Freeman C, Falconer N, de Camargo Catapan S, Gray LC, Paterson DL. Clinical impact of antibiograms as an intervention to optimize antimicrobial prescribing and patient outcomes: A systematic review. Am J Infect Control 2024; 52(1):107-22.

https://doi.org/10.1016/j.ajic.2023.08.013